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TRIM24 orchestrates metabolic reprogramming and EMT in Breast Cancer

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**TRIM24 ORCHESTRATES METABOLIC REPROGRAMMING AND EMT
IN BREAST CANCER**

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**TRIM24 ORCHESTRATES METABOLIC REPROGRAMMING AND EMT
IN BREAST CANCER**

**A
THESIS (DISSERTATION)**

Presented to the Faculty of
The University of Texas
Health Science Center at Houston
and
The University of Texas
MD Anderson Cancer Center
Graduate School of Biomedical Sciences
in Partial Fulfillment
of the Requirements
for the Degree of
DOCTOR OF PHILOSOPHY

By
Kaushik Thakkar, M.S.

Houston, Texas

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Dedication

To all my family, my mentors, my teachers, and my friends
for their love, teaching, and support.

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I would like to express my deepest gratitude to the many people who have helped me throughout my PhD education. Without their teaching, advice, encouragement and support, I would not have been able to complete my dissertation.

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TRIM24 orchestrates metabolic reprogramming and EMT in breast cancer

Kaushik Thakkar, MS

Advisory Professor: Michelle C. Barton, PhD

In this dissertation, I report the oncogenic functions of an epigenetic regulator Tripartite Motif Protein 24 (TRIM24) coupled with metabolic reprogramming and epithelial mesenchymal transition (EMT) in breast cancer. TRIM24 was first established by our laboratory as a previously unknown negative regulator of p53 via its RING domain, as a co-regulator of nuclear receptors and a PHD/Bromodomain reader of specific histone modifications. TRIM24 expression correlates with poor prognosis of breast cancer, but the mechanisms of TRIM24-mediated oncogenesis are unknown. In the first part of my thesis, I found that TRIM24 is aberrantly expressed in early stages of breast cancer progression. Ectopic expression of TRIM24 induces malignant transformation of immortalized human mammary epithelial cells (TRIM24-iHMECs) and efficient growth of intermediate to high-grade xenograft tumors.

TRIM24 induces gross metabolic changes in the iHMECs with increased glycolysis and TCA cycle along with glucose uptake. TRIM24 causes deregulation of c-Myc and p53, the two key players that regulate cancer metabolism. TRIM24 is directly recruited to the promoters of several metabolic genes such as *GLUT1*, *IDH1*, *IDH2* and *c-Myc*. Our hypothesis is that direct activation of various glucose metabolism genes by TRIM24 coupled with deregulation of key players of metabolism is critical for TRIM24-mediated oncogenic transformation in breast cancer.

TRIM24 is over-expressed in several cancers; however, a detailed analysis of how TRIM24 promotes tumorigenesis with progression to metastasis has not been elucidated. Prior to the metastatic progression of a tumor, cancer cells undergo a transition from an epithelial to a mesenchymal phenotype; termed as epithelial-mesenchymal transition (EMT). In the second part of my thesis, I found that TRIM24 is a driver of EMT in breast epithelial cells.

Upon TRIM24 over-expression in MCF10A cells, a mesenchymal-like change in the cellular morphology was observed. TRIM24-MCF10A cells exhibited marked decrease of E-cadherin and acquisition of various mesenchymal markers such as N-cadherin, fibronectin and vimentin. Similarly, upon TRIM24-knockdown in mesenchymal cells MDA-MB-231 and MDA-MB-468, there is reversal of EMT gene signature. Gene expression analysis of TRIM24-OE MCF10A cells further confirmed that EMT was among the top pathways that were enriched. Additionally, TRIM24 may be involved in direct transcriptional regulation of EMT genes as suggested by TRIM24-ChIP analysis and the presence of Histone H3 – K23 acetylation (H3K23ac) signature, for which the TRIM24-Bromodomain acts as a histone reader, at several EMT genes in MDA-MB-468 cells. We propose that aberrant expression of TRIM24 impacts a network of transcription regulatory factors and pathways to induce EMT and cancer metastasis.

For effective reduction of cancer mortality, it is very critical to intervene in the critical transition of non-invasive ductal carcinoma to extremely lethal invasive breast cancer. Our preliminary analysis suggests that TRIM24 plays a critical role in mechanisms involving both primary and metastatic cancer. Future studies will be focused to develop a better understanding of molecular mechanisms of TRIM24' oncogenic function and will pave the way for developing therapeutic opportunities by targeting TRIM24.

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List of Abbreviations

2-NBDG	2-(N-(7-nitrobenz-2-oxa-1,3-diazol-4-yl)amino)-2-deoxyglucose
ATCC	American Type Culture Collection
cDNA	complementary deoxyribonucleic acid
CMV	Cytomegalo virus
ChIP	Chromatin Immunoprecipitation
CIN	Chromosomal instability
DMEM	Dulbecco's Modified Eagle Medium
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
ECAR	Extracellular Acidification Rate
EMT	Epithelial-mesenchymal transition
ER	Estrogen receptor alpha
FCCP	Carbonyl cyanide 4-(trifluoromethoxy)phenylhydrazone
GSEA	Gene Set Enrichment Analysis
IHC	Immunohistochemistry
iHMEC	isogenic Human mammary epithelial cells
MDACC	M.D. Anderson Cancer Center
mRNA	messenger ribonucleic acid
NP-40	Nonidet P40
OCR	Oxygen Consumption Rate
OXPHOS	Mitochondrial respiration or oxidative phosphorylation
OE	Over-expression
qRT-PCR	quantitative Realtime Polymerase Chain Reaction

RNA	Ribonucleic acid
RBCC	RING- B-box and Coiled coil
shRNA	short hairpin ribonucleic acid
TCA	Tricarboxylic Acid cycle
TIF1	Transcription Intermediary Factor 1
TRAP	Transcription Factor Affinity Prediction analysis
TRIM	Tripartite motif containing family of proteins

Chapter 1: Introduction

Chapter 1: Introduction

1.1 Breast cancer:

Breast cancer is the most common malignant disease of Western women and it is the second leading cause of cancer-related deaths after lung cancer [1]. It is estimated that approximately 234,190 Americans will be diagnosed with invasive breast cancer and about 40,730 Americans will die of the disease in 2015-16. It is alarming to find out that the lifetime risk of breast cancer for American women has risen from 1 in 11 in the 1970s to 1 in 8 in 2013, which is attributed to shifting breast cancer risk factor demographics and the increased use of screening mammography [1].

Breast tumors are highly heterogeneous and are composed of multiple subtypes with distinct morphologies and clinical implications [2]. Breast tumors are classified based on three different criteria: (i) histology into ductal or lobular carcinomas, (ii) differentiation state/gene expression profiles into luminal and basal-like subtypes, and (iii) the expression of receptors such as estrogen (ER), progesterone (PR) and HER2 (human epidermal growth factor receptor 2) into ER+, HER2+, and ER– PR– HER2– (triple-negative breast cancer, TNBC) subtypes. Expression patterns of the ER+ and HER2+ tumors both have luminal features, however TNBCs show significant but not complete overlap with basal-like subtype [3]. One of the key reasons for a decrease in mortality rates of breast cancer patients is the widespread use of mammography that has led to increased diagnosis of early stage disease, including ductal carcinoma *in situ* (DCIS). Systematic characterization of breast cancer genomes has identified that somatic mutations in only three critical genes (TP53, PIK3CA and GATA3) occurred at more than 10% incidence across all breast cancers [3].

Classification of breast tumors based on hormone receptor and HER2 status and the use of antihormonal and HER2-targeted therapy, respectively, are among the first examples for molecular-based classification and personalized cancer treatment that made a significant difference in clinical outcomes. Three main groups of medications used for adjuvant therapy of breast cancer include hormone-blocking agents, chemotherapy and monoclonal antibodies [4]. Blocking of estrogen action can be achieved by using selective estrogen receptor modulators (SERMs) such as tamoxifen, which compete with estradiol for binding to ER α , and which induce an alteration of ER α 's conformation that is not conducive for activation. Selective ER α down-regulators (SERDs) such as fulvestrant induce destabilization and degradation of ER α . Finally, another way to block ER α action is to reduce the production of estrogen in peripheral tissues and within the tumor using inhibitors of aromatase, an enzyme which synthesizes estrogens from testosterone and androstenedione [5].

For almost three decades in both early and advanced breast cancer patients, tamoxifen has been the mainstay of endocrine therapy and remains to date the most successful targeted breast cancer therapy. However, it is unclear if aromatase inhibitors (AI) offer advantages over tamoxifen as adjuvant treatments for postmenopausal women [6]. The results of the Arimidex, Tamoxifen Alone or in Combination (ATAC) trial [7], and the International Breast Cancer Study Group's Breast International Group (BIG) 1-98 trial [8] showed small but significant improved efficacy of AIs, and in general fewer side effects.

Chemotherapy is primarily used for ER- breast tumors and for patients in advance stages of breast cancer. Combinatorial chemotherapy is the preferred option and it involves use of cyclophosphamide along with doxorubicin ('AC' regime) or

cyclophosphamide along with methotrexate and fluorouracil ('CAT' regime) [9]. For HER2+ breast tumors, trastuzumab, a HER2-specific monoclonal antibody has improved the 5-year disease free survival to about 87% [10].

1.2 Cancer metastasis and EMT:

Cancer progression involves the following steps: adenoma, carcinoma *in situ* and invasive carcinoma at the primary tumor site, (Fig. 1) [11]. A portion of cancer cells disseminate from the primary tumor, adapt and grow at a secondary site or organ other than the primary tumor sites, the phenomena is termed as 'cancer metastasis'. The distribution of tumors and the sites of tumor growth are not a chance event, but it is determined by the amount of 'seeds' (the cancer cells) on the 'congenial soil' (the destination organ for metastasis) as postulated by the Dr. Stephen Paget in 1989 [12]. Approximately 90% of breast cancer-related deaths are caused by metastasis not by primary tumor [13]. Hence, for effective reduction of cancer mortality, it is very critical to intervene in the critical transition of non-invasive ductal carcinoma to extremely lethal invasive breast cancer. The pathogenesis of cancer metastasis consists of sequential steps such as migration, invasion, proliferation and angiogenesis [14].

Epithelial to mesenchymal transition (EMT) is a fundamental biological process and plays a critical role in morphogenic processes of the developing embryo, wound healing and organ fibrosis [15]. Tumor cells frequently use EMT to gain migratory and invasive properties in order to relocate and eventually form distant metastasis [16]. During EMT, the cancer cells undergo aggressive behavioral changes; acquiring a spindle-shape, loss of desmosomes and adherens junctions (such as E-cadherin, ZO-1, occludins, and claudins), increased expression of mesenchymal markers (such as N-

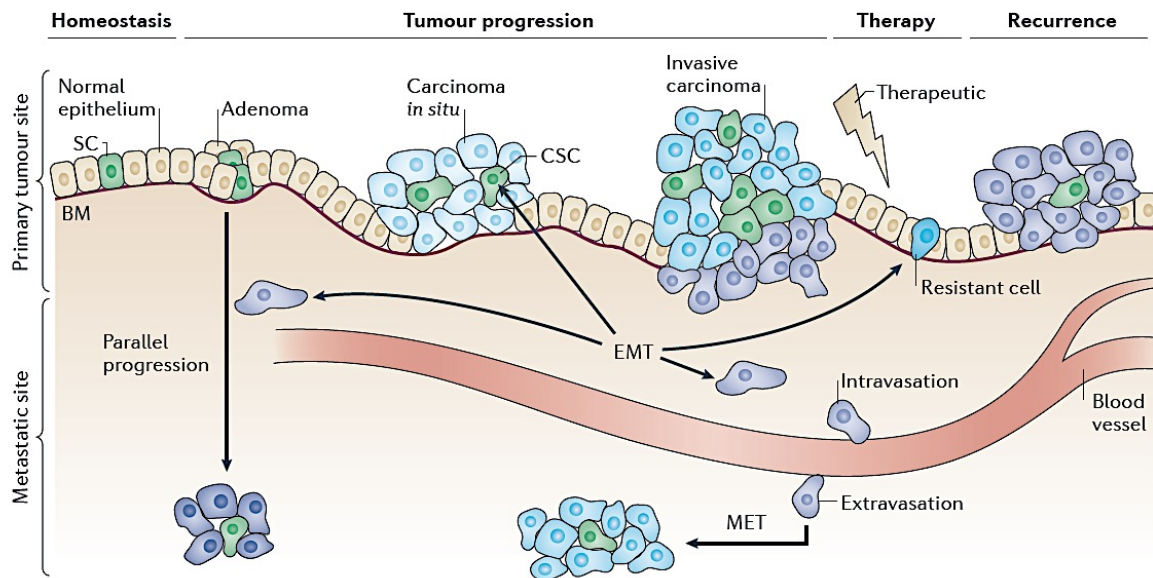


Figure 1: Role of EMT in cancer progression.

A primary tumor needs to undergo Epithelial to mesenchymal transition (EMT) in order to metastasize from a primary to a distant site.

(Figure is taken with permission from Craene B.D. and Berx, G. Regulatory networks defining EMT during cancer initiation and progression. Nat. Rev Cancer. 2013; 13: 97-110.) License Number: 3777250370389

cadherin, fibronectin and vimentin) and secretion of proteolytic enzymes, such as matrix metalloproteases, in order to clear the extracellular matrix (ECM) and exhibit increased migratory properties (Fig. 2) [15]. Of note, cancer cells undergoing EMT exhibit increased chemotherapy resistance, decreased apoptosis and have traits similar to stem cells [17].

Both intrinsic and extrinsic stimuli are known to induce EMT in cancer. Microenvironment, one of the key extrinsic stimuli, is composed of ECM, myofibroblasts, cancer-associated fibroblasts, immune cells and several secretory factors such as hepatocyte growth factor, epidermal growth factor, Wnt, transforming growth factor- β , Hedgehog and various cytokines such as interleukin-6 and tumor necrosis factor- β [18]. A cohort of transcription factors including those of the Snail, Twist and Zeb families are known to orchestrate the process of EMT [19]. These transcription factors have unique tissue-specific expression profiles and are known to control the expression of each other and cooperate functionally at target genes [20]. EMT transcription factors in concert not only repress epithelial genes but also activate mesenchymal genes, and often the same transcription factor is responsible for both activation and repression [11].

EMT transcription factors recognize E-box DNA sequences located near the transcription start site of *CDH1* gene, and recruit various co-factors and histone deacetylases to transcriptionally repress *CDH1*. Notably, SNAI1 induces recruitment of HDAC1 and HDAC2 to induce repressive histone modifications at the *CDH1* promoter and likewise, ZEB1 cooperates with deacetylase SIRT1 at the *CDH1* promoter leading to deacetylation of histone H3 and reduced binding of RNA polymerase II [11]. In addition, various histone modifiers are linked to SNAI1 activity such as the

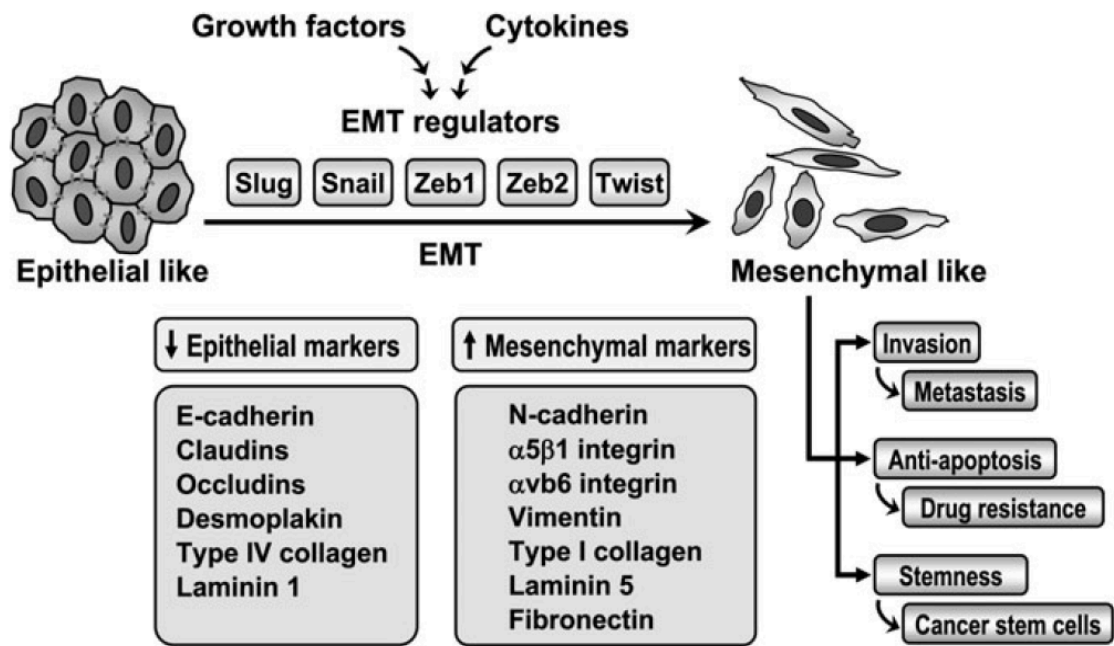


Figure 2: An overview of regulators of EMT

The EMT regulators transform the cancer cells from epithelial-cell like to mesenchymal-cell like with suppression of epithelial markers and expression of mesenchymal markers. The final effects on cancer cells are cancer metastasis, drug resistance and with features of cancer stem cells.

(Figure is taken with permission from Shih JY, Yang PC. The EMT regulator slug and lung carcinogenesis. Carcinogenesis. 2011 Sep;32(9):1299-304.) License Number:

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methyltransferase EZH2, SUZ12 and G9a (EHMT2) [11]. Similarly, TWIST induces another repressive histone mark H4K20 monomethylation by recruiting the methyltransferase SET8 at E-cadherin promoters [21]. On the other hand, in head and neck carcinoma, expression of BMI1 is activated by TWIST1 and they both subsequently coordinate to repression of E-cadherin and p16 by recruiting the PRC2 complex [22]. Interestingly under hypoxic conditions, the hypoxia inducible factor (HIF1a) induces EMT and tumor metastasis by enhancing TWIST expression [23]. The ZEB family of transcription factors recruits co-repressors such as C-terminal binding protein (CTBP) or chromatin remodeling protein, BRG1 [24]. Additionally, ZEB1 may also interact with p300/CBP-associated factor (PCAF) and p300 and form a transcriptional activation complex [25]. Therefore, similar to SNAI1 and TWIST, ZEBs function as transcriptional repressors or activators by binding to E-boxes and recruit corresponding complexes, ultimately repressing epithelial junction and polarity genes and activating mesenchymal genes that contribute to EMT.

1.3 Cancer cells and deregulated metabolism:

In the second cancer hallmarks review published by Drs. Hanahan and Weinberg, 'metabolic reprogramming' is recognized as one of the key hallmarks of cancer [26]. Multiple research groups are trying to understand how a tumor evolves metabolically in order to adapt to the uptake of novel nutrients or microenvironment. However, because of the complex wiring of metabolic circuits and the constant evolution of metabolic switching, it has been difficult to completely characterize cancer metabolism. Current research is focused on understanding how to restrict nutrient consumption in these cancer cells or ways to deplete the nutrient uptake to starve

these cells, which would ultimately cause reduced proliferation and growth of the tumor. Additionally, identification of novel regulators of cancer metabolism and uncovering their mechanism of action are critical.

Normal cells convert glucose to pyruvate that is later converted to acetyl-CoA to fuel the tricarboxylic acid cycle (TCA cycle). NADH and FADH₂ generated at the end of TCA cycle then provide electrons to the mitochondrial respiratory chain and ultimately each glucose molecule can produce up to 36 molecules of ATP. In normal cells, glycolysis is prioritized only when oxygen supply is limited. In contrast, cancer cells preferentially use glycolysis even in normoxic conditions; this phenomenon was discovered by Dr. Otto Warburg and is termed as 'Warburg effect' (Fig. 3) [27]. Hence, glycolysis in tumor cells is termed as 'aerobic glycolysis' to distinguish from the normal anaerobic glycolysis of healthy cells [28].

Glycolysis only generates 2 ATP per glucose molecule as compared to 36 ATP for each glucose molecule catabolized by mitochondrial respiration; hence, cancer cells have to compensate for the lower efficacy of energy production. They do this partly by up-regulating the expression of glucose transporters such as GLUT1 and GLUT4, to uptake more glucose and a majority of other glycolytic enzymes [29]. Oncogenes such as Ras, Myc, and HIF-1 α are reported to be master inducers of cancer glycolysis [30]. Elevated transcriptional activity of c-Myc and HIF-1 α along with p53 loss/mutation is responsible for enhancing expression of glycolytic enzymes. The majority of glycolytic gene promoters have consensus binding motifs of Myc and HIF-1 α . In normoxia, c-myc promotes expression of glycolytic genes, whereas when cells are in hypoxic HIF-1 α is activated that allows tumors to continuously drive glycolysis to not only support proliferation but also accelerated biosynthesis [31].

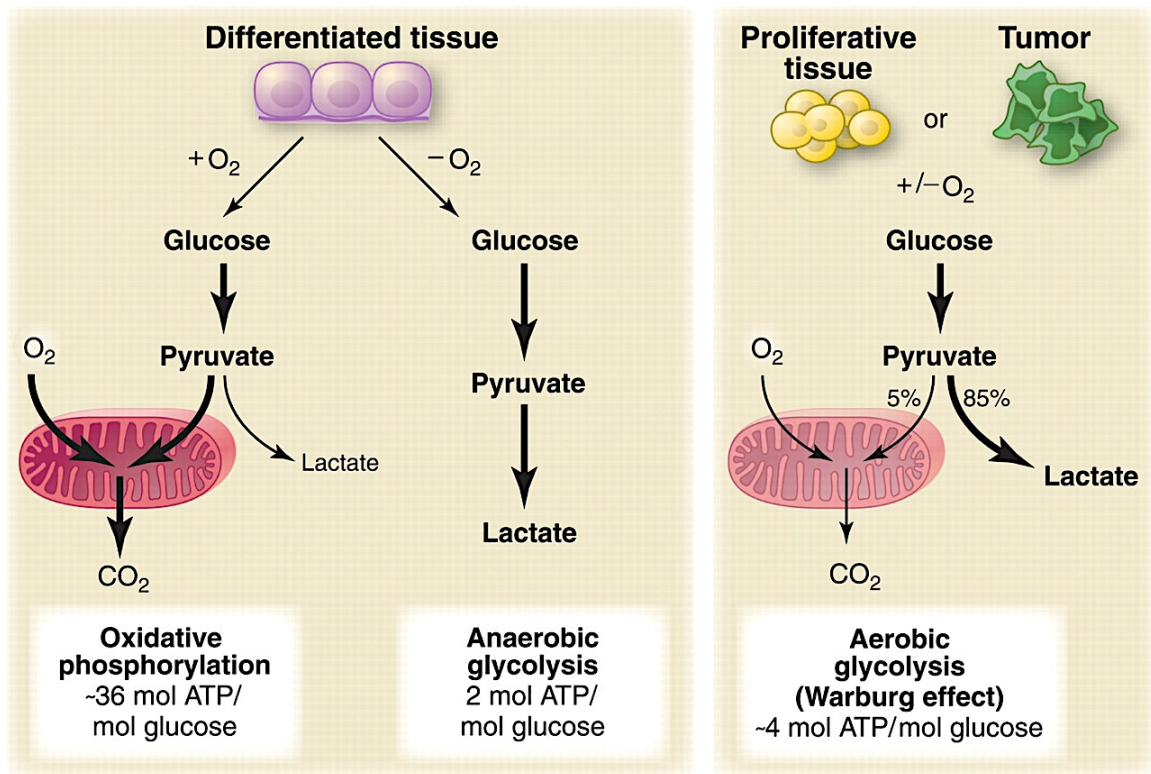


Figure 3: Tumor metabolism and Warburg effect

Normal differentiated tissues or cells, utilize glucose as an energy source through the glycolysis pathway and enter into Kreb's cycle to produce more energy through oxidative phosphorylation. On the contrary, cancer cells, consume increased amounts of glucose and switch to produce lactate without entering oxidative phosphorylation. Cancer cells are dependent on this switch to aerobic glycolysis for proliferation

(Figure is taken with permission from Vander Heiden, M.G., Cantley, L.C. and Thompson, C.B. Understanding the Warburg effect: The metabolic requirements of cell proliferation. Science. 2009; 324(5930):1029-1033.) License Number: 3777851053796

In contrast, the tumor suppressor p53 inhibits glucose uptake by directly inhibiting the transcription of glucose transporter GLUT1 and GLUT4 [32]. Fructose 2,6-bisphosphate functions as an allosteric activator of PFK1, a major glycolytic enzyme. Fructose 2,6-bisphosphate is produced by PFK2 from fructose 1-phosphate. p53 induces the expression of *TIGAR* and *TIGAR* significantly slows down tumor glycolysis by converting fructose 2,6-bisphosphate back to fructose 1-phosphate [33]. Hence, one of the key molecular interactions that define the status of cancer metabolism is that of p53, c-Myc and HIF-1 α .

As per the Warburg hypothesis, it was erroneously believed that mitochondrial metabolism is inconsequential for the metabolic demands of actively proliferating cancer cells. Recent studies, provide ample genetic evidence that mitochondria are indeed essential for tumorigenesis [34]. Further, it is proposed that tumors are metabolically heterogeneous (Fig. 4). The bulk of tumor cells, exhibit aerobic glycolysis (Warburg effect) and there is a small population of slow-cycling tumorigenic cells that are dependent on OXPHOS activity. The slow-cycling cells are proposed to be the cells that are resistant to various cytotoxic treatments [35].

1.4 TRIM family and TRIM24:

The TRIpartite Motif (TRIM) containing family of proteins consists of about 71 proteins that have the signature tripartite, 'RBCC', motif of RING-finger domain, one or two zinc binding motifs named B-boxes, and an associated Coiled-Coil region. On the basis of differences in their C-terminal domain structure, the TRIM proteins can be classified into 11 different sub-families (Fig. 5). Most of the TRIM family proteins act as

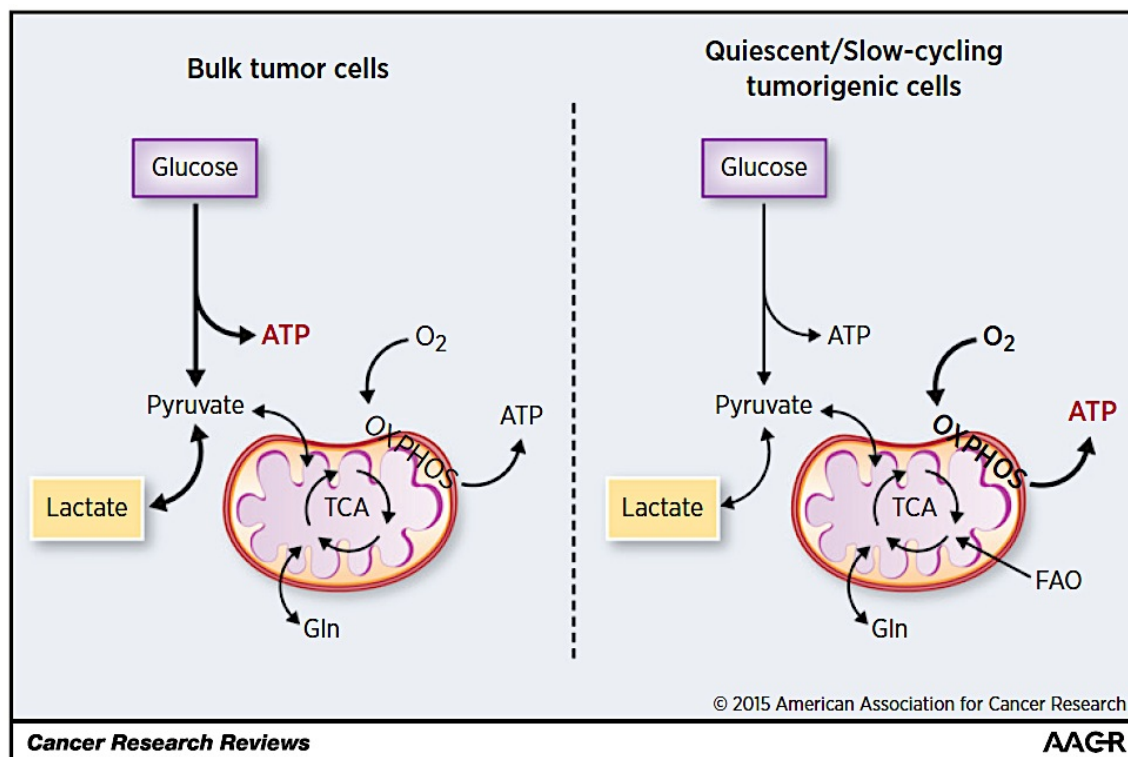


Figure 4: Diverse metabolism in different population of cancer cells

Different subpopulations of cancer cells coexist in tumors, contributing to functional heterogeneity. Rare tumorigenic cells exist in a quiescent/ slow-cycling state and are intrinsically resistant to cytotoxic treatments. Importantly, with respect to other cells inside the tumor, they seem to rely on a different metabolic program to maintain their energetics. In contrast to the bulk tumor, slow-cycling tumorigenic cells are more dependent on mitochondrial respiration (OXPHOS) and have impaired glycolytic capacity.

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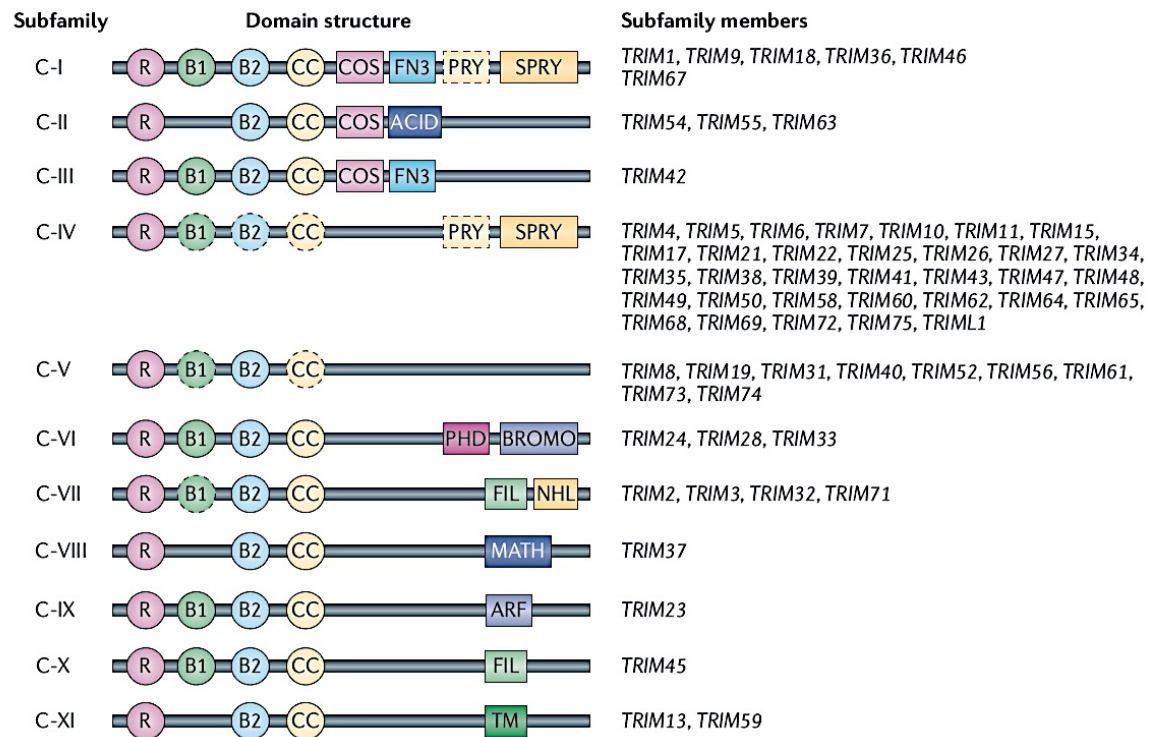


Figure 5: The TRIM family of proteins.

Based on protein structure the TRIM family of proteins is classified into nine eleven different sub-families. Almost all members have a RING-finger domain (R), one or two B-box domains (B) and a coiled-coiled domain (CC).

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E3-ubiquitin ligases and they function in various cellular processes, including regulation of protein stability, transcription, cell proliferation and apoptosis [36].

The Transcription Intermediary Factor (TIF1) sub-family is a unique sub-family of TRIM proteins that in addition to the N-terminal RBCC motif contains a tandem PHD-Bromodomain at the C-terminus. TRIM24 (TIF1 α) is the founding member of the TIF1 sub-family, which includes TRIM28 (TIF1 β), TRIM33 (TIF1 γ) and TRIM66 (TIF1 δ) [37]. Generally, a PHD-domain interacts with methylated lysines of histones, whereas the Bromo-domains interact with acetylated lysines of histones [38, 39]. Various members of TIF1 sub-family play key roles in regulating epigenetic mechanisms of transcriptional regulation that involve histone modifiers, nuclear receptors and heterochromatin binding proteins [37].

Oncoprotein T18 was one of the very first reports of TRIM24, in which it was found to be a fusion partner along with B-Raf protein. T18 had the first 332 amino acids of TRIM24 in its N-terminus and B-Raf protein at the C-terminus [40]. Using yeast two-hybrid screens, it was shown that mouse heterochromatin protein 1 alpha (mHP1a) interacts with TRIM24 via mHP1 chromo-shadow domain [41]. As the name suggests, HP1 proteins are associated with formation of heterochromatin and inhibition of gene expression. Using luciferase reporter assays, it was shown that interaction of TRIM24 and mHP1a associated with gene repression. In contrast, another group showed that TRIM24 exhibits its transcription repression activity in the absence its HP1-binding region in a similar reporter system [42]. Bonus, a *Drosophila* homolog of TIF1 proteins, is essential for development and interacts with various nuclear receptors such as betaFTZ-F1 and DHR3 [43]. Of note, the N-terminal RBCC motif of Bonus is essential for its transcriptional repression activity, but not the HP1-binding region and the C-

terminal PHD/Bromo domain. Hence, whether an HP1 interaction is essential for TRIM24-mediated gene regulation is not clear.

In contrast, TRIM24 localization studies reveal that TRIM24 associates with euchromatin of interphasic nuclei, and is not associated with condensed chromatin and metaphase chromosomes [44]. TRIM24 physically interacts with arginine methyltransferase 1 (CARM1) and glucocorticoid receptor-interacting protein 1 (GRIP1) to enhance transcriptional activation mediated by nuclear receptors [45]. TRIM24 knockdown is reported to diminish transcriptional activation mediated by GRIP1 and AR. Additionally, TRIM24 also reportedly interacts with several chromatin remodeling proteins. In mouse development during the zygote stage, TRIM24 is localized to transcriptional active sites co-occupied by SNF2H and BRG-1 that are known chromatin-remodeling proteins [46]. Notably, TRIM24 depletion caused mislocalization of RNA Pol-II, BRG-1 and SNF-2 and down-regulation of their target genes in the zygote. Hence, these results suggest that TRIM24 may play an active role in transcriptional activation.

TRIM24 acts a transcriptional co-regulator by virtue of its LXXL motif by which it interacts with various nuclear receptors such as RAR α , Vitamin D3, androgen and estrogen receptors (Fig. 6). TRIM24 modulates the transcriptional activity of various nuclear receptors either positively or negatively in a ligand dependent manner. In prostate cancer cells, TRIM24 directly interacts with a histone acetyltransferase, TIP60, a coactivator of androgen receptor (AR) [47]. Additionally, TRIM24 also binds to bromodomain-containing 7 (BRD7), a corepressor of AR and negative regulator of cell proliferation. Thus, TRIM24 regulates AR-mediated transcription through crosstalk between TIP60 and BRD7 in prostate cancer [47].

TRIM24: A multifunctional transcriptional regulator

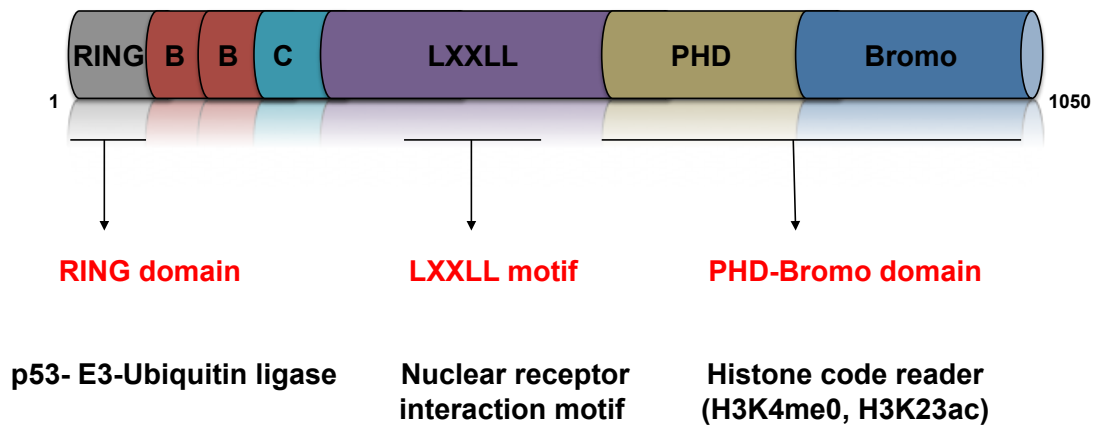


Figure 6: TRIM24 protein structure

Diagram of TRIM24 protein structure and domains, along with its known functions.

In a mouse model targeting exon 4 of TRIM24 for germline deletion *Trim24*^{dlE4/dlE4}, it was discovered that RAR-mediated signaling was reactivated, through re-expression of RAR α and repressed genes in the liver, leading to eventual HCC [48]. Additionally, TRIM24 was shown to suppress HCC progression by inhibition of the IFN/STAT pathway via RAR dependent regulation of the Stat1 promoter [49]. Based on analysis of this mouse model, it was suggested that TRIM24 is a liver-specific, retinoid-dependent tumor suppressor [48]. However, recent evidence suggests that *Trim24*^{dlE4/dlE4} mice retain normal levels of *Trim24* RNA lacking exon 4 and are not null of TRIM24 in liver [50]. Our laboratory generated a new mouse model for complete TRIM24 knockdown by targeting exon 1 of TRIM24 (*TRIM24*^{dlE1/dlE1}). It was found that *TRIM24*^{dlE1/dlE1} mice have a significant depletion of visceral fat and spontaneously develop liver steatosis, fibrosis and ultimately hepatocellular carcinoma. In the murine liver, TRIM24 acts as a transcriptional co-repressor by repressing hepatic lipid accumulation, inflammation, and damage [51].

Based on multiple studies it can be concluded that TRIM24 plays a dual role of transcriptional co-activator and transcriptional co-repressor, and its role is dependent on the tissue type and associated complexes. Interestingly, it has been reported that TRIM24 may phosphorylate various transcription factors by acting as a protein kinase *in vitro* [52]. However, based on protein sequence analysis TRIM24 contains no conserved characteristics of protein kinase domains and there are no reported *in vivo* substrates.

Our laboratory identified TRIM24 as a p53-interacting protein, upon purification of an endogenous TAP-tagged version of p53 in embryonic stem cells (ES cells) [53]. Knockdown of TRIM24 ortholog, *bonus*, in *Drosophila* induces apoptosis. Further,

mosaic analysis in *Drosophila* depleted in the TRIM24 ortholog, *bonus*, shows that p53 could rescue apoptosis of imaginal disc cells. Similarly, TRIM24 knockdown in breast cancer cell lines (MCF7) results in p53-dependent apoptosis [53]. Additional studies in MCF7 breast cancer cell lines, which harbor wild-type p53, showed that TRIM24 responds to stress and damage by negatively regulating p53 through an autoregulatory feedback loop. It was found in response to DNA-damage the TRIM24-p53 interaction is disrupted due to ATM-mediated TRIM24 phosphorylation. In turn, the damage-activated p53 binds to p53 response elements on TRIM24 gene and induces TRIM24 gene expression. Finally, the newly synthesized TRIM24 terminates the DNA damage response by targeting phosphorylated-p53 for proteasomal-mediated degradation. This feedback loop was found to be independent of MDM2 and dependent upon direct binding of p53 to p53 response elements (p53 REs) in the distal promoter region of TRIM24 [54].

The aforementioned studies provide genetic evidence that TRIM24 acts as a tumor suppressor in murine liver, but there are multiple studies that provide evidence that TRIM24 acts as an oncogene in various cancers such as in prostate cancer [47], breast cancer [55, 56], non-small cell lung cancer [57], head and neck carcinoma [58], colorectal cancer [59], gastric cancer [60], glioblastoma [61], bladder cancer [62] and even human hepatocellular carcinoma [63] (Table 1). Expression of TRIM24 has been reported to correlate negatively with survival of breast cancer patients [55, 56]. Molecular and three-dimensional structural analysis showed that TRIM24 via its tandem PHD-Bromo domain acts as a dual histone code reader of H3K4me0 and H3K23ac. It was found that TRIM24 binds estrogen receptor and chromatin (at specific

Cancer	Human/ Mouse	Oncogene/Tumor Suppressor	Clinical data	Mechanism	Ref.
HCC	Mouse	Tumor Suppressor	No	RAR-a corepressor	[48, 51]
Breast	Human	Oncogene	Yes	ER-a coactivator	[55]
NSCLC	Human	Oncogene	Yes	NA	[57]
HNSCC	Human	Oncogene	Yes	NA	[58]
CRC	Human	Oncogene	No	NA	[59]
Gastric	Human	Oncogene	Yes	NA	[60]
GBM	Human	Oncogene	Yes	PIK3CA/Akt	[61]
Bladder	Human	Oncogene	Yes	NA	[62]
HCC	Human	Oncogene	Yes	NA	[63]

Table 1: List of various studies of TRIM24 in different cancers

histone modifications mentioned above) to activate estrogen-dependent genes associated with tumor development and cell proliferation [55]. Hence, by reading a non-canonical histone signature the PHD-Bromo domain of TRIM24 provides a structural rationale for gene regulation, establishing a novel route by which histone readers may influence pathogenesis of cancer.

Similar to other correlations in human cancers, it was found that TRIM24 expression positively correlates with glioma malignancy and it is a marker of poor prognosis for patients with newly diagnosed glioblastoma (GBM) [61]. TRIM24 expression is critical for efficient cell proliferation, cell cycle progression, and *in vivo* tumor growth in GBM. These authors showed that TRIM24 directly activates *PIK3CA* gene expression in a PHD-Bromo domain dependent manner. Intriguingly, TRIM24-mediated regulation of the PI3K/AKT/NF- κ B pathway induces the expression of DNA repair gene O⁶-methylguanine-DNA methyltransferase (MGMT) and induces chemoresistance to temozolomide in GBM cells [61].

Although, it has yet to be determined if TRIM24 can target mutant p53 for proteasomal-based degradation, in a very elegant study it was found that TRIM24 may be part of a protein complex responsible for conversion of mutant p53 conformation to its WT form [64]. The studies were done in mouse embryonic stem cells harboring mutant p53 (ES-cells^{mutp53}) that remained pluripotent and had a normal karyotype as compared to ES-cells knocked out of p53. In ES-cells^{mutp53}, the mutant p53 was transcriptionally active in response to DNA-damage and activated canonical p53-target genes such as p21 and *btg2*. Among the network of proteins identified by MS-based interactome analysis of p53, in addition to TRIM24, was CCT complex, USP7, Aurora kinase and Nedd4 [64].

To summarize, there are several reports of TRIM24 association with poor prognosis and survival in various cancers [55, 56]. TRIM24 regulates gene expression by acting either as transcriptional activator or repressor. However, whether TRIM24 plays a causal role in malignant transformation during tumor development and progression has yet to be studied. Further, the mechanisms involved in potential TRIM24-mediated tumor initiation and progression are unknown.

Chapter 2: Materials and methods

Chapter 2: Materials and methods

The contents of this chapter are based on Pathiraja TN, Thakkar KN, Jiang S, Stratton S, Liu Z, Gagea M, Shi X, Shah PK, Phan L, Lee MH, Andersen J, Stampfer M, Barton MC. TRIM24 links glucose metabolism with transformation of human mammary epithelial cells. *Oncogene*. 2015 May 28; 34(22): 2836-45.

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2.1 Cell culture

Previously established HMEC cultures, 184D (normal pre-stasis [65]), 184B-7p (abnormal, post-stasis [65]), 184A1 (Immortal non-malignant[66]) and 184AA2 (Immortal malignant [67]) were cultured in M87A medium with 0.1% AlbuMAX bovine serum albumin (Invitrogen) and 0.1 nmol/L oxytocin (Bachem) as described [65]. MCF10A cells were cultured as described [68]. Human breast cancer cell lines (MCF-7, MDA-MB-231, MDA-MB-468, and SKBR3) from ATCC were cultured according to the supplier's protocols. For transient knockdown experiments, TRIM24 (L-047483-00) and non-silencing control (D-00120-01-20) small interfering RNAs (siRNAs) (Dharmacon) were transfected at 100 nM for 48 hours using Lipofectamine RNAiMAX reagent (Invitrogen) as per manufactures instructions.

2.2 Cell proliferation, soft agar colony formation, and cell-cycle analysis

For cell proliferation (7500 cells in 24 well plate, in triplicates) and XTT assays (500 cells in 96 well plates, in quadruplets), respective cell numbers were seeded and counted every 2 days for upto 6 days (Beckman Coulter counter). For soft agar colony formation, 5000 cells were seeded in 1.5 ml of respective medium with 0.35% low-melting-temperature agarose (BD Biosciences) overlying 1.5 mL of 0.7% low-melting agarose in 6-well plates and incubated at 37°C for 14 days. For cell-cycle analysis, cells were washed and fixed with 70% ethanol for 1 hour at 4°C and stained with propidium iodide at a concentration of 50 µg/mL, along with RNase A (100 U/mL), prior to analysis using Flow cytometry and ModFit software program (Verity Software House).

2.3 Generation of stable cells

TRIM24 full-length coding sequences were amplified from the vector pCMV-XL4-hsTRIM24 (OriGene Technologies) using gene-specific primers (Appendix 1), digested with *Xba*I and *Pml*I and cloned into pEntry4-FLAG vector [69]. TRIM24 coding sequences and FLAG-only sequences were transferred to pLenti-PGK-Neo-DEST [69] using Gateway cloning (Invitrogen). Lentiviruses containing culture supernatants were prepared as described [55]. Briefly, 800 ng of clone/vector DNA were transfected into HEK293T cells with packaging vectors pPAX2 and pMD2 (800 ng each) using Effectene (Qiagen) according to manufacturer's instructions. After 48 hours, target cell line was transduced using virus-containing culture supernatants with 8 µg /mL Polybrene (Sigma). After 48 hours, cells were selected using 700 µg/mL G418

(Stratagene) for 10 days. Stable cell lines, ectopically expressing *TRIM24* or FLAG only were maintained in regular culture medium containing 200 µg/ml G418.

2.4 Quantitative real-time PCR (qRT-PCR)

Total RNA was isolated with TRIzol reagent (Invitrogen) and qRT-PCR analysis was performed as described [55]. Primers used are listed in Appendix 1.

2.5 nCounter analysis

A panel of 420 breast cancer associated genes was used for nCounter analysis (NanoString Technologies) [70]. Three biological replicates of RNA from TRIM24 and Control iHMECs (100ng each) were processed according to the manufacturer's recommendations (<http://www.nanostring.com>). Data were analyzed using the nSolver digital analyzer software program (<http://www.nanostring.com/products/nSolver>). Differentially expressed genes were identified using the Student *t*-test: genes with expression significantly ($p < 0.05$) up- (fold change ≥ 1.2) or down-regulated (fold change ≤ 0.8) were identified. Unsupervised hierarchical clustering of differentially expressed genes was performed using the GENE-E software program (<http://www.broadinstitute.org/cancer/software/GENE-E/>) and a Euclidean distance matrix. Gene Ontology (GO) category (<http://www.geneontology.org/>) enrichment analysis and KEGG pathway analysis were done using the Database for Annotation, Visualization and Integrated Discovery (DAVID) (<http://david.abcc.ncifcrf.gov/>) [71].

2.6 Immunoblotting and Immunohistochemistry

Immunoblotting of proteins in TRIM24 and Control iHMECs was performed using standard techniques with antibodies listed in Appendix 2. Immunohistochemistry (IHC) of breast tumor arrays, BR2082 (US Biomax) and BR1503 (US Biomax), was performed, as described [55], using a VECTASTAIN Elite ABC Kit and a DAB Detection Kit (Vector Laboratories) with an anti-TRIM24 antibody (1:200 dilution; Proteintech). IHC staining data for ER, PR, and ErbB2 were available and TRIM24 staining was quantified using the Allred scoring method (for details of the Chi-square method and calculations please see Appendix 3) [72].

2.7 Metabolic experiments and glucose uptake

The extracellular acidification rate (ECAR) and oxygen consumption rate (OCR) of TRIM24 and Control iHMECs were measured using an XF^e96 Extracellular Flux Analyzer (Seahorse Bioscience) as described [73]. Briefly, 50,000 cells were seeded into XF^e96 tissue culture plates in quadruplicate and ECAR and OCR measured under basal conditions and after sequential treatment of the cells with Oligomycin (1 μ M), FCCP (Carbonylcyanide p-trifluoromethoxyphenylhydrazone, 500 nM) and Rot + Ant [Rotenone (1 μ M) + Antimycin A (1 μ M)]. Cells were trypsinized and counted using a Coulter counter (Beckman Coulter) and the cell count used for normalization purposes. For glucose uptake assays, a non-metabolic fluorescent-labeled glucose (2- NBDG) was used. iHMECs were seeded in the regular HMEC medium for 24h and just prior to performing the assay were washed with glucose-free medium. Cells were incubated with glucose-free medium, containing 120 μ M 2-NBDG (Invitrogen) for 0.5h, 1.0h, 1.5h and 2h, followed by FACS analysis.

2.8 Xenograft experiments

Female athymic Nu/Nu mice (age 6–8 weeks) were housed under pathogen-free conditions. 1 million TRIM24 iHMECs or Control iHMECs suspended in 50 μ l of serum free DMEM were co-injected subcutaneously with 50 μ l of high concentration Matrigel (BD Biosciences); 10-15 mice were injected for each cell line. Tumor sizes were measured weekly by caliper for up to 64 days and xenograft volume calculated as previously described [74]. All procedures were approved by the University of Texas M D Anderson Cancer Center Institutional Animal Care and Use Committee and performed with veterinary supervision.

2.9 The Cancer Genome Atlas data and statistical methods

The Cancer Genome Atlas – Breast invasive carcinoma (TCGA-BRCA) data on RNA expression (Level 3) of Breast invasive carcinoma patients (1008) and normal tissues (92) in terms of RSEM (RNA-seq by Expectation-Maximization) were downloaded from the Broad Institute TCGA Firehose Genome Data analysis center pipeline (standard data run 2013/11/14). 950 of 1008 BRCA samples had PAM50 subtype information available. Gene set enrichment analysis (GSEA) was run with default parameters with TRIM24 expression levels as phenotype and C2 (canonical pathways) gene sets. GSEA with patient samples limited to Basal subtypes and TRIM24 expression as phenotype gave very similar results.

2.10 Invasion and migration assays

Cells were resuspended in 500 μ l serum-free media and seeded in a 24-well Transwell or Matrigel plate (BD Biosciences, pore size 8 μ m) at a concentration of $(5 \text{ to } 7.5) \times 10^4$

per well. 750 μ l of DMEM/ RPMI supplemented with 10% FBS was added to the lower chamber and cells were allowed to migrate or invade for 16 h in 5% CO₂ at 37°C. Migrating cells were stained with 0.1% crystal violet. Non-migrating cells were removed using a cotton swab. Migrated cells were quantified based on five microscopic fields at a 4X magnification and results were represented as mean \pm standard deviation and student's t-test was performed for statistical significance. Each assay was performed in triplicates.

2.11 Acini formation assay

The overlay method of acini formation assay for MCF10A cells was performed as described [68]. Briefly, 40ul of growth-factor reduced matrigel (BD Biosciences – No. 354230) was added to each well of an eight-well glass chamber slide (Fisher – 1256518) and spread evenly in the well using the P-200 tip (the slides can be assembled on ice). MCF10A cells were trypsinized and resuspended in 2 ml of Resuspension medium and spun down for 3 min at 150g in a tissue culture centrifuge. The cells were then mixed in assay medium without EGF and mixed 10-25 times to obtain a single-cell suspension. 25,000 cells per ml cells were mixed 1:1 with assay medium containing 4% matrigel and 10 ng/ml of EGF (usually 1ml of each). 400 μ l of this mixture was plated on the top of solidified matrigel in each well of the chamber slide. Final concentration of cells is 5000 cells per well in assay medium containing 2% matrigel and 5 ng/ml EGF. Cells are allowed to grow in 5% CO₂ humidified incubator at 37°C for 10-12 days and the cells are refed with fresh assay medium containing 2% matrigel and 5 ng/ml EGF every 4 days.

2.12 Experimental lung metastasis studies

To determine the experimental metastatic potential of cells, 5×10^5 MDA-MB-231 control-shRNA (GIPZ) and TRIM24-shRNA (GIPZ) cells labeled with RFP-firefly luciferase were injected into NOD/SCID mice via the tail vein. SCID Mice were assessed weekly for presence of lung metastasis after the intraperitoneal injection of D-Luciferin (Fisher – PI88293). *In vivo* bioluminescence was assessed using the IVIS imaging system 200 series (PerkinElmer), approximately 10 mins post-injection. Until about 6-7 weeks, mice were monitored for lung metastasis weekly and were euthanized after 7 weeks. Lungs were removed surgically and fixed using Bouin's fluid and the number of lung tumor nodules was counted and subjected to hematoxylin and eosin staining thereafter. Lungs were also stained for RFP and TRIM24 to ascertain the presence of MDA-MB-231 cells and TRIM24 expression, respectively. All procedures were approved by the University of Texas M D Anderson Cancer Center - IACUC and performed under proper supervision of veterinary personnel.

2.13 HTG EdgeSeq analysis

mRNA expression profiling of 2,560 genes was done using the HTG EdgeSeq Oncology Biomarker Panel assay. The assay uses an NGS-based gene expression system powered by HTG EdgeSeq chemistry. Samples were processed according to the manufacturer's recommendations (<http://www.htgmolecular.com/science/sample-prep>). Differentially expressed genes were identified using the Student t-test: genes with expression significantly ($p < 0.05$) up- (fold change ≥ 1.2) or down-regulated (fold change ≤ 0.8) were identified. Gene Ontology (GO) category enrichment analysis and KEGG pathway analysis were done using the molecular signatures database (MSigDB) [75].

2.14 Chromatin immunoprecipitation (in breast cancer cells)

Chromatin immunoprecipitation (ChIP) is routinely used in our laboratory [55]. Briefly for chromatin immunoprecipitation, cells are harvested and then cross-linked with formaldehyde for 10 min and neutralized with glycine. Cells are lysed with 1 mL Cell Lysis Buffer (5mM PIPES pH 8.0, 85mM KCl, 0.5% NP-40, fresh protease inhibitors) for 15 minutes. After sonication, the tubes are spun. Supernatant is transferred to a new tube to check for fragment size. Lysates are divided and diluted using ChIP Lysis Buffer. Immunoprecipitation is performed overnight with specific antibodies. The next day, Protein A Sepharose beads (GE Health) are incubated with antibody/protein bound complexes for 2 hr at 4°C. Then, Protein A beads are washed and re-suspended and incubated with RNase A for 30 min at 37°C and then Protease K for 2 hours at 55°C. The crosslinks are reversed by incubating the samples at 65°C overnight. The next day, protein /antibody bound DNA fragments are extracted with Phenol/Chloroform. qPCR analyses are performed to analyze specific antibody- and protein-bound.

2.15 Co-immunoprecipitation

Cells were lysed in NTEP buffer (25mM Tris-HCl pH 7.5, 150mM NaCl, 5mM EDTA, 0.5% NP-40) for 30 minutes at cold room. Using high-speed centrifugation, 14,000rpm for about 10 minutes, the cell lysate is spun down. Protein concentration was measured with BCA assay. 500 µg of cell lysate was pre-cleared by incubating with Protein A sepharose beads (GE health). 500 µg cell lysate was used to incubate per respective antibody overnight in the cold room. Then, protein A Sepharose beads (GE Health) were added for additional 2 hours incubation. After extensive washing, 35 µl SDS

loading dye was added into the beads, the beads was boiled for 5-10 minutes to elute the protein complex. Elute was then subject to western blot in various percentage of SDS-PAGE gels.

2.15 Karyotyping

a) Protocol for Chromosome preparation:

Exponentially growing cells were exposed to Colcemid (0.04 µg/ml) for one hour at 37°C and to hypotonic treatment (0.075 M KCl) for 20 minutes at room temperature. Cells were fixed in a methanol and acetic acid (3:1 by volume) mixture for 15 minutes, and washed three times in the fixative. Slides were prepared by dropping 50-100 µl the cell suspension on wet slides and air-dried.

b) Protocol for G-banding:

Slides were baked at 90° C for 3 hours. They were then treated with trypsin (200µl of 40X trypsin +1X HBSS, pH 7) for 2.5 ~ 3.5 minutes. Stained in 8% Giemsa staining solution for 3 ~ 4 minutes. Images were captured using Nikon 80i microscope equipped with Spectral Karyotyping software from Applied Spectral Imaging (ASI, Vista, CA). 15-20 metaphases were karyotyped from each cell line.

*Karyotyping was performed at MD Anderson Cytogenetics Core

2.16 Immunofluorescence

Cells were grown on a sterile glass cover slips or slides overnight in 37°C/5% CO₂ incubator. Slides were then rinsed briefly with PBS and cells were fixed by incubation with 1% formalin in PBS for 10 minutes. Slides were then rinsed three times

with PBS. Incubate with primary antibody for 1 hour at room temperature or overnight at 4°C. Optimal antibody concentration is usually from 1-10ug/ml in PBS-BSA. Wash three times with PBS. Incubate with conjugated secondary antibody for 45 minutes. Again, optimal antibody concentration is usually from 1-10ug/ml in PBS-BSA. Wash three times with PBS. Mount aqueous coverslip

Chapter 3: TRIM24 links glucose metabolism with transformation of Human mammary epithelial cells

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The contents of this chapter are based on Pathiraja TN, Thakkar KN, Jiang S, Stratton S, Liu Z, Gagea M, Shi X, Shah PK, Phan L, Lee MH, Andersen J, Stampfer M, Barton MC. TRIM24 links glucose metabolism with transformation of human mammary epithelial cells. *Oncogene*. 2015 May 28; 34(22): 2836-45.

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3.1 Introduction and Rationale

Recent studies indicate that specific members of the tripartite motif (TRIM) protein family, characterized by conserved amino (N)-terminal zinc-finger domains of a RING-type E3 ubiquitin ligase, B-boxes and coiled coil, are important regulators of carcinogenesis [36]. Among these, we identified TRIM24, as a previously unknown E3-ubiquitin ligase of p53 in embryonic stem cells and breast cancer cell lines, and as a histone reader that activates estrogen-dependent genes associated with cellular proliferation and tumor development [53, 55, 76]. Of note, we and others reported that high expression of TRIM24 is associated with poor prognosis and survival in breast cancer patients [55, 56]. However, whether TRIM24 plays a causal role in malignant transformation of breast epithelial cells during breast tumor development and progression has yet to be studied. Further, the mechanisms involved in potential TRIM24-mediated breast tumor initiation and progression are unknown.

3.2 Aberrant expression of TRIM24 during breast cancer progression

To determine whether TRIM24 expression in breast tissues was deregulated during breast cancer progression, we performed IHC-staining to detect TRIM24 protein expression in a human tissue microarray (BR2082, US Biomax) consisting of samples of normal breast tissue, atypical ductal hyperplasia, intraductal breast carcinoma, and invasive breast carcinoma. We detected low TRIM24 protein expression in normal breast tissue but high expression in atypical ductal hyperplasia and carcinoma (Fig. 7A), suggesting that TRIM24 expression is deregulated in breast cancer and likely early in progression. We then examined the expression of TRIM24 in 1008 breast cancer patients and 92 normal samples from The Cancer Genome Atlas – Breast invasive carcinoma (TCGA-BRCA) dataset. We found the TRIM24 was significantly upregulated in breast invasive carcinoma patients (p-value: $1e-16$, Fig. 7B) and its expression in paired samples was greater than 1.5 fold in 40 out of 106 (37.8%) patients (Fig. 8A).

Next, we assessed whether high levels of TRIM24 expression were associated with any specific breast cancer sub-type by using the TCGA-BRCA dataset and by performing TRIM24-IHC in an array of tissue samples from 72 breast cancer cases. In the TCGA-BRCA dataset, the PAM50 (Prediction analysis of Microarray – 50 genes expression signature[77]) breast-cancer subtypes showed different distribution in TRIM24 high expressing versus low expressing patients (p-value: $1.32e-07$). The basal subtype (Odds ratio: 1.98, p-value: $3.8e-04$) was significantly over-represented in the TRIM24 high expressing patients, followed by HER2 (Odds ratio: 1.78, p-value: 0.03) and Luminal B subtype (Odds ratio: 1.4, p-value: 0.047); whereas, the Luminal A subtype (Odds ratio: 0.43, p-value: $1.46e-08$) was significantly under-represented (Fig. 7C). For details of the intersection of PAM50 and TRIM24-expression analysis, please

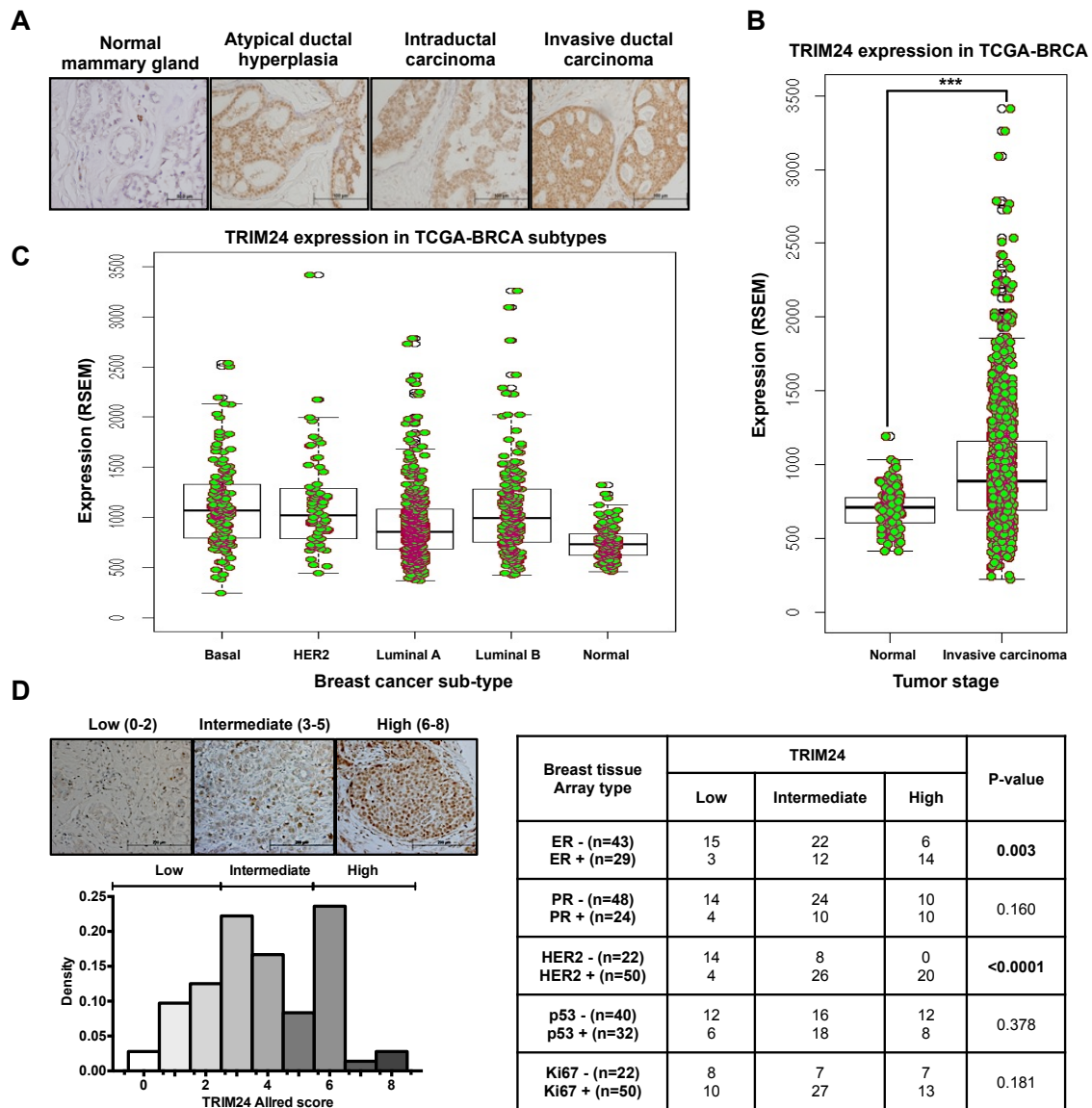
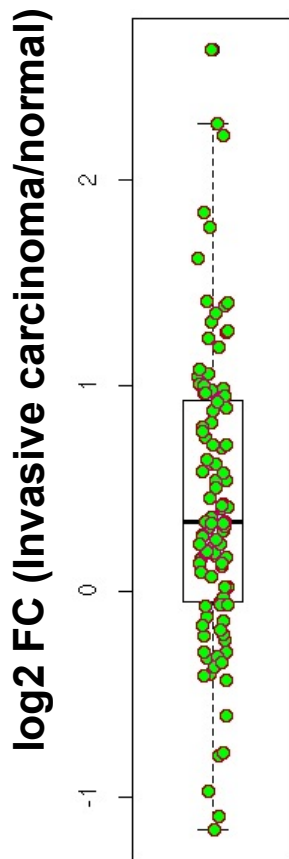


Figure 7: Aberrant expression of TRIM24 during breast cancer progression.

A, TRIM24-Immunohistochemical (IHC) staining was performed on a breast tissue array (BR2082, US Biomax) with tissues from different stages of breast cancer progression such as normal mammary gland, atypical ductal hyperplasia, Intraductal carcinoma and invasive ductal carcinoma, B, Distribution of TRIM24 expression (p-value:1.32e-07) in TCGA breast invasive carcinoma patients (1008) and normal tissues (92) and C, in PAM50 breast cancer subtypes (950). D, TRIM24 expression in breast tissue array (BR1503, US Biomax) by Allred score (left), Correlation of TRIM24

expression with ER, PR, HER2, p53, and Ki67 (right and bottom left, as assessed by IHC). *** $p < 0.001$.

A TRIM24 expression in TCGA BRCA



Paired Samples

- Fold change > 0 for 78 of 106 (73%) patients with paired profiles
- 40 out of 106 (37.8%) patients have fold change > 1.5

B Top 30% vs. Bottom 70%

TRIM24 =>	High Exp	Low Exp
LumA	107	325
LumB	77	117
Basal	66	74
Her2	31	36

p-value: 1.32e-07

TRIM24 =>	High Exp	Low Exp
LumA	107	325
Not LumA	174	227

p-value: 1.46e-08; Odds ratio: 0.43

TRIM24 =>	High Exp	Low Exp
Lum B	77	117
Not LumB	204	435

p-value: 0.047; odds ratio: 1.40

TRIM24 =>	High Exp	Low Exp
Basal	66	74
Not Basal	215	478

p-value: 3.8e-04; Odds ratio: 1.98

TRIM24 =>	High Exp	Low Exp
Her2	31	36
Not Her2	250	516

p-value: 0.03; Odds ratio: 1.78

Key:

LumA: ER+, PR+, HER2-; **LumB:** ER+, PR+, HER2+

HER2: ER-, PR-, HER2+; **Basal:** ER-, PR-, HER2-

Figure 8: Aberrant expression of TRIM24 in paired samples of invasive carcinoma and classification based on breast cancer-subtypes

A, Distribution of TRIM24 expression in TCGA breast invasive carcinoma patients in 106 paired samples; B, Distribution of TRIM24 expression in TCGA breast invasive carcinoma patients based on PAM50 breast cancer subtypes (950). TRIM24 high-expression patients were defined as 30% of patients with high TRIM24 expression. The

p-values suggest enrichment of a particular subtype in TRIM24 high-expressing patients

see Fig. 8B. In the breast cancer tissue array (BR2082, US Biomax), TRIM24 expression stratified into three classes: low (score, 0-2), undetectable to low expression in few foci (25%); intermediate (score, 3-5), abundant foci with expression in nuclear and cytoplasmic compartments (47%); and high (score, 6-8), abundant foci with high expression in nuclei (28%, Fig.6D). Of note, chi-square testing identified a statistically significant positive correlation of TRIM24 expression with ErbB2 (HER2) expression ($p < 0.0001$) and ER ($p = 0.003$) – Appendix 3.

To estimate a timeline for deregulation of TRIM24 expression in breast cancer cells during malignant transformation, we used an isogenic human mammary epithelial cells (iHMECs) model that facilitates assessment of molecular changes from the earliest stages of human breast carcinogenesis (Fig. 9A). In this model, the transformation of normal, finite lifespan iHMECs to malignant cells requires overcoming two distinct senescence-associated barriers to immortality [78, 79]. The first, a stress-associated barrier called stasis, is overcome or bypassed in cultured iHMECs by inactivation of the RB pathway, commonly through loss of p16^{ink4a} expression [65, 66, 80, 81]. iHMECs that escape stasis undergo further proliferation before encountering the second, more stringent barrier to immortality due to critically shortened telomeres [81]. In rare instances, cells that gain telomerase expression escape this barrier and acquire immortal potential. Additional perturbations, such as insertional mutagenesis of p53 [67] or transduction of specific oncogenes [82] can confer malignant properties to the immortally transformed cells. We found that endogenous TRIM24 expression increases at both RNA and protein levels early in the transformation process, after stasis is overcome (184B-7p [65, 78]), and continues to be highly expressed in the immortalized HMEC lines 184A1[66] and 184AA2[67] (Fig. 9B). Strikingly, TRIM24 expression was even more highly up-regulated in MCF-7 cells (p53 wild type and ER positive),

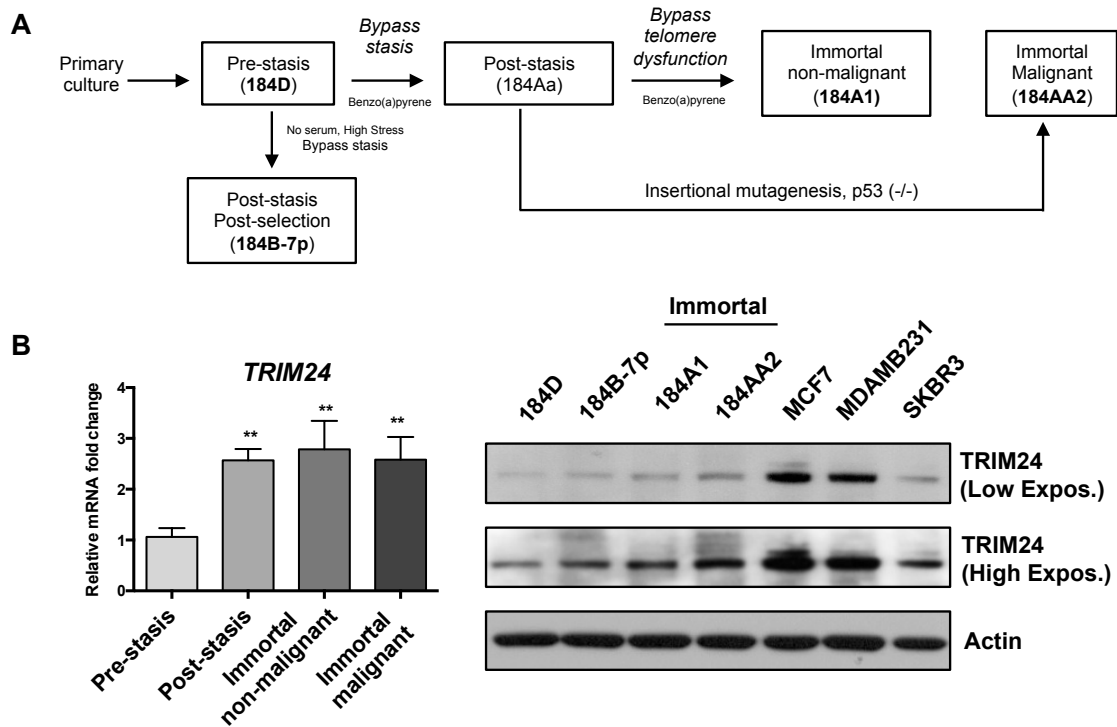


Figure 9: Aberrant expression of TRIM24 in an isogenic HMEC model

A, Schematic of the isogenic HMEC model showing transition of the cells from finite-lifespan cells to immortal and malignant cells. B, TRIM24 mRNA (left) and protein (right) expression in HMEC lines transitioning from “normal” finite-lifespan cells before stasis to immortal and malignant cells. Data are averages from three biological replicates \pm standard deviation (SD). ** $p < 0.01$.

MDAMB231 (p53- mutant and triple negative) and SKBR3 (p53-mutant and HER2 positive) cultured lines of breast tumor-derived cells (Fig. 9B). Thus, the progressive stages of breast cancer in human patient tumor samples and immortalization of isogenic HMEC lines support a correlation between TRIM24 expression and loss of normal, cellular homeostasis.

3.3 TRIM24 drives iHMEC transformation and survival

To answer the critical question of whether TRIM24 causes malignant transformation of iHMECs, we induced ectopic expression of TRIM24 in the non-malignant, immortal 184A1 HMEC (iHMEC) line. Analysis of stable control- (FLAG-only) and TRIM24-expressing, lentiviral-transduced iHMEC pools from two independent experiments (TRIM24-1 and TRIM24-2) showed increased TRIM24 expression at both the RNA and protein level, compared to control pooled cells (Fig. 10A). Of note, we see a corresponding decrease in p53 protein expression in both TRIM24-1 and TRIM24-2 iHMECs suggesting that the ectopically expressed TRIM24 is enzymatically active [53]. The growth rates of TRIM24-1 and TRIM24-2 iHMECs were significantly higher than control vector-transduced iHMECs, suggesting increased proliferation (by cell count see Fig. 10B; by XTT assay see Fig. 10C). Consistent with this, both TRIM24-1 and TRIM24-2 iHMECs exhibited significant enrichment of cells in the S and G₂/M phases of the cell cycle, compared to control cells enriched in G₁, indicating rapid cell-cycle progression as a result of TRIM24 expression (Fig. 10D, for flow data see Fig. 11).

To assess the oncogenic potential of TRIM24-iHMECs, we examined their ability to grow in an anchorage-independent manner in soft agar. Our control iHMEC line failed to form colonies in soft agar, whereas TRIM24-iHMECs formed colonies comparable in

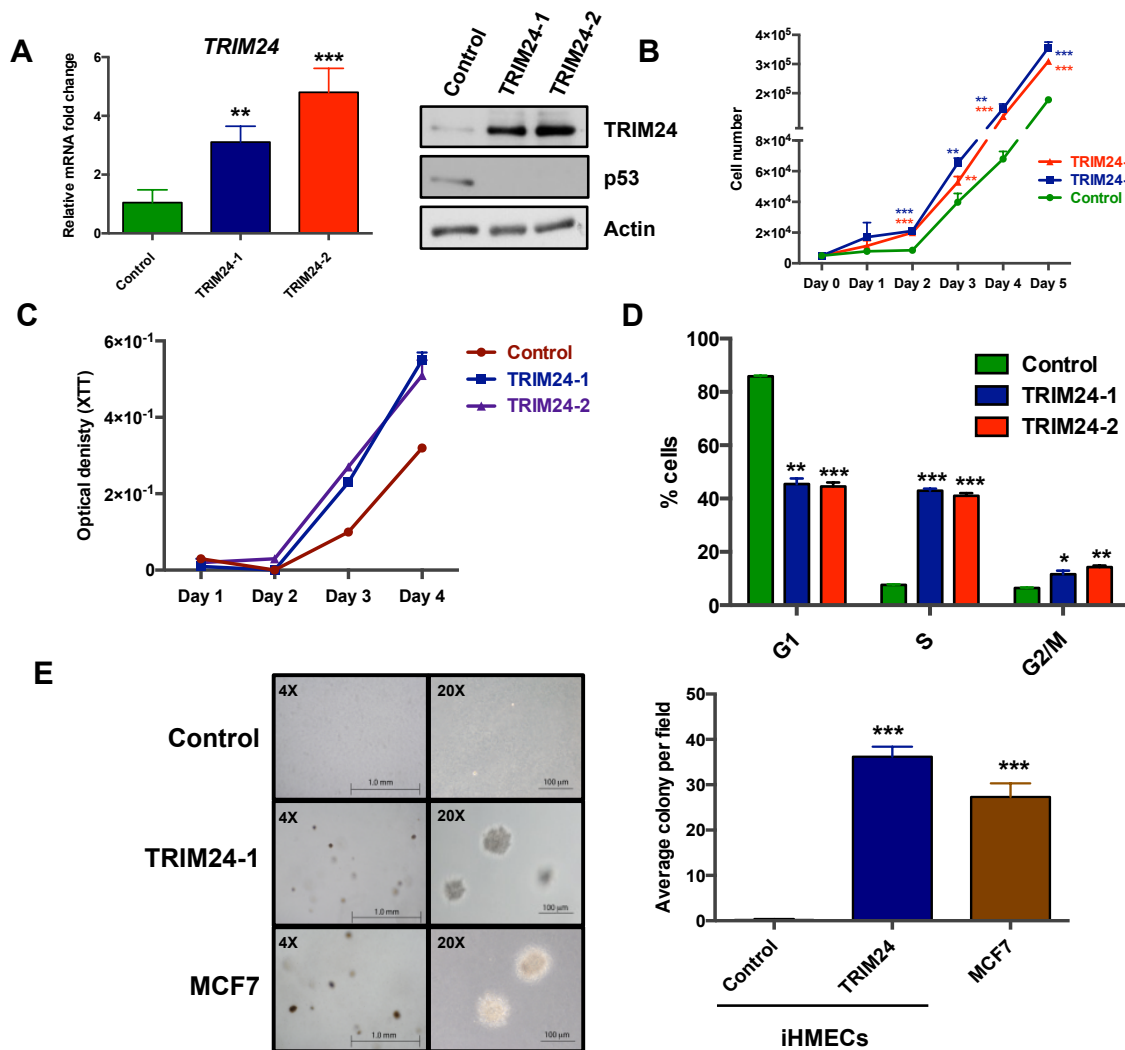
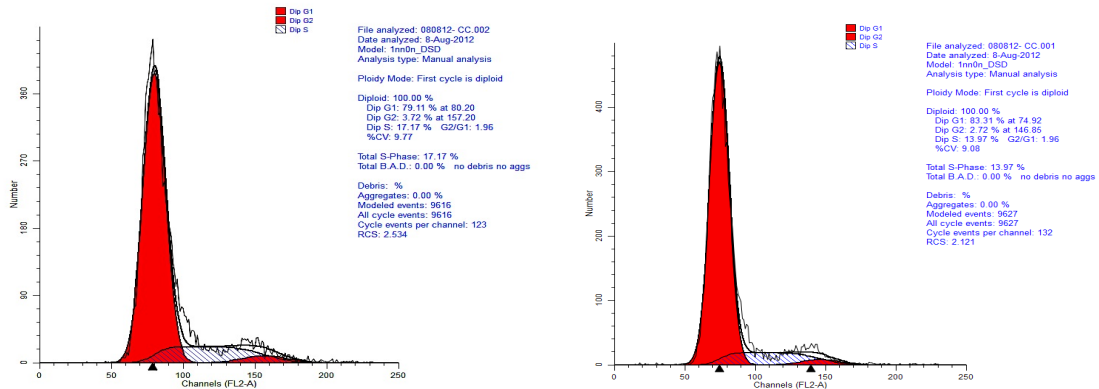


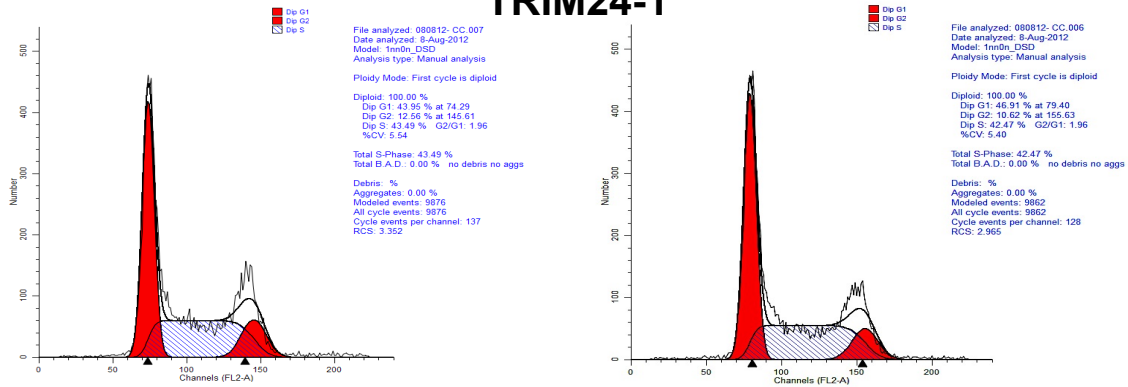
Figure 10: TRIM24 drives iHMEC transformation and survival.

A, qRT-PCR analysis of TRIM24 mRNA and Immunoblot of TRIM24 and p53 in Control, TRIM24-1, and TRIM24-2 iHMECs. B & C, Cell proliferation analysis of Control, TRIM24-1, and TRIM24-2 iHMECs by cell count and XTT-based analysis. C, Cell-cycle analysis of Control, TRIM24-1, and TRIM24-2 iHMECs. D, Anchorage-independent proliferation of Control iHMECs, TRIM24-1 iHMECs, and MCF-7 cells as determined using a soft agar colony formation assay (left) and quantification of the colonies per field \pm SD (right). The data are averages from two replicates \pm SD. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

Control



TRIM24-1



TRIM24-2

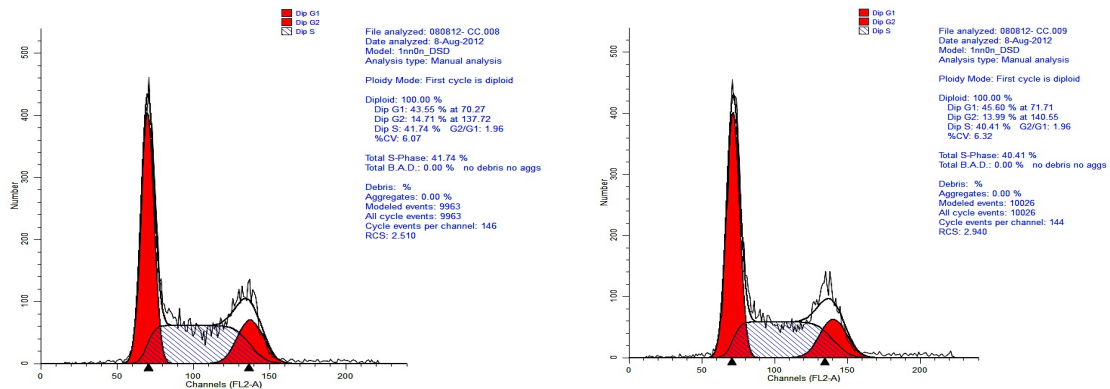


Figure 11: Graphical representation of cell cycle changes in control vs TRIM24-ihMECs

Flow data for cell cycle analysis of Control, TRIM24-1 and TRIM24-2 ihMECs.

size and number to colonies of MCF-7 cells used as a positive control (Fig. 10E). Thus, ectopic expression of TRIM24 confers anchorage-independent growth to iHMECs, suggesting that TRIM24 over expression is sufficient to induce malignant transformation of immortal, nontumorigenic HMECs.

3.4 TRIM24 regulates multiple cancer-associated pathways and promotes a glycolytic and TCA-cycle gene signature in HMECs

To understand the mechanisms of TRIM24-induced transformation of iHMECs, we analyzed the gene expression profiles of 420 cancer-associated genes in TRIM24-iHMECs and control iHMECs using the nCounter analysis system. This analysis identified 233 genes (55% of the total) that were significantly changed suggesting that TRIM24 mediated deregulated expression of cancer-relevant genes. Expression of 127 (30%) of these genes was upregulated, whereas 106 genes (25%) were downregulated (Appendix 4). Unsupervised hierarchical clustering of the 233 differentially expressed genes clearly distinguished the control and TRIM24-iHMECs (Fig. 12A), supporting TRIM24-mediated transformation of iHMECs via effects on cancer-associated genes and pathways.

TRIM24 induced expression of genes frequently altered in cancers, including *PTEN*, *NRAS*, *KRAS* and others that enrich amongst multiple cancer pathways (see Appendix 5 and 6). Additionally, TRIM24 up-regulated genes are significantly enriched ($p \leq 0.05$) in functional pathways that impact metabolism and growth, including the citrate cycle (TCA), ErbB, insulin, mitogen-activated protein kinase (MAPK) and mTOR signaling and cell cycle regulation (Fig. 12B). The highly significant pathways downregulated by

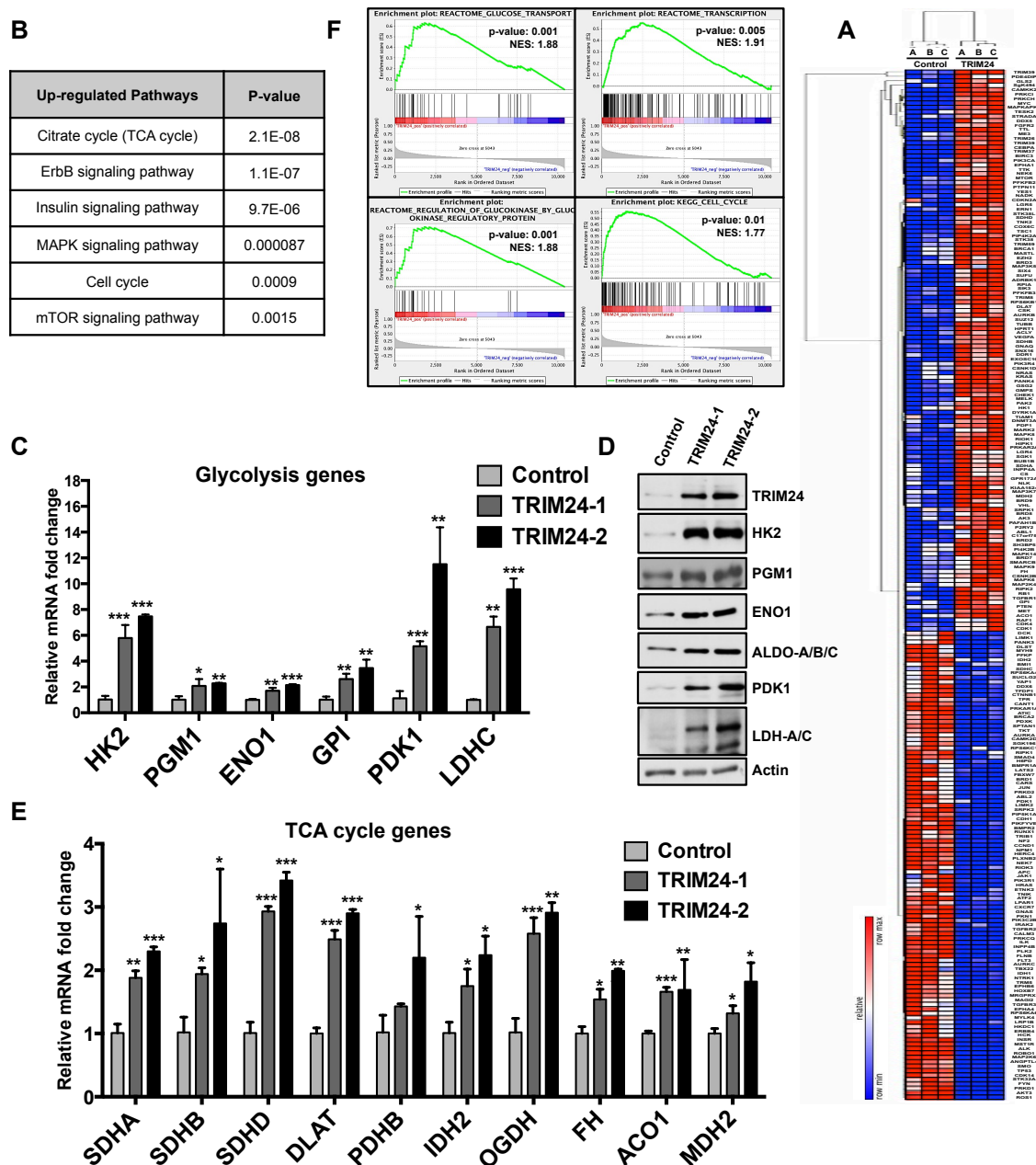


Figure 12: Regulation of multiple cancer-associated pathways in iHMECs by TRIM24.

A, Heat map showing unsupervised hierarchical clustering of 233 differentially expressed genes in TRIM24-1 compared to the Control. B, Table showing the list of Upregulated pathways associated with Growth and Metabolism in TRIM24-iHMECs C, qRT-PCR analysis of Glycolytic genes in Control, TRIM24-1, and TRIM24-2 iHMECs showing upregulation of their expression. D, Immunoblot for various Glycolytic proteins

in Control, TRIM24-1, and TRIM24-2 iHMECs. E, qRT-PCR analysis of TCA-cycle genes in Control, TRIM24-1, and TRIM24-2 iHMECs. The data are averages from three biological replicates \pm SD. F, GSEA profiles of different pathways positively correlated to TRIM24 expression in TCGA-BRCA dataset. NES: Normalized Enrichment Score. *p < 0.05; **p < 0.01; ***p < 0.001.

TRIM24 expression includes TP53, representing a breadth of cancer types and pathways, as well as members of the ErbB pathway (see Appendix 5 and 6).

Analysis of individual glycolytic regulatory genes uncovered a glycolytic signature, specific to TRIM24-iHMECs versus control, marked by increased Hexokinase 2 (*HK2*), Phospho-glutamase 1 (*PGM1*), Enolase 1 (*ENO1*), Glucose-6-phosphate isomerase (*GPI*), Pyruvate dehydrogenase kinase 1 (*PDK1*), and Lactate dehydrogenase C (*LDHC*) mRNA expression (Fig. 12C). Gene expression changes were reflected in elevated HK2, PGM1, ENO1, PDK1, and LDHC protein expression, along with elevated Aldolase A (Aldo) protein (Fig. 12D). HK2 is a key enzyme involved in tight regulation of glycolysis via catalysis of individual, irreversible steps of glucose to glucose-6-phosphate and phosphoenolpyruvate to pyruvate [83].

Additionally, we found that expression of TCA-cycle genes was also higher in TRIM24-1 and TRIM24-2 iHMECs compared to control iHMECs. Succinate dehydrogenase complex, subunits A, B, and D (*SDHA*, *SDHB* and *SDHD*); dihydrolipoamide S-acetyltransferase (*DLAT*); isocitrate dehydrogenase 2 (*IDH2*); oxoglutarate dehydrogenase (*OGDH*); aconitase 1 (*ACO1*); malate dehydrogenase 2 (*MDH2*) and fumarate hydratase (*FH*) were among the TCA-associated genes with up-regulated expression (Fig. 12E). Collectively, these results suggest that TRIM24 expression in iHMECs and subsequent transformation rely on increased glucose metabolism to meet the higher energy demands of increased growth and proliferation. This metabolic shift is marked by up-regulation of both glycolysis and expression of TCA-cycle genes.

To determine if these findings are relevant to human breast disease, we assessed whether TRIM24 expression correlated with altered expression of glucose metabolism genes in human breast tumors using TCGA-BRCA expression data and Geneset

enrichment analysis (GSEA) on invasive breast carcinoma samples. Consistent with our *in vitro* findings, we found that the glucose transport pathway (p-value: 0.001, Normalized Enrichment Score (NES[75]): 1.88) was among the top 10 pathways positively correlated with TRIM24 expression (Fig. 12F). For a complete list of enriched pathways please see Appendix 7. Thus, TRIM24 has a significant and/or strong tendency to be co-expressed with genes that regulate glucose metabolism in breast tumors, supporting clinical relevance of our findings.

3.5 TRIM24 expression results in diverse metabolic states and increased glucose uptake in iHMECs

To determine the functional impact that TRIM24 expression has on cellular metabolism, we examined the two main bio-energetic pathways: oxidative phosphorylation (OXPHOS) and glycolysis. The rate of oxygen consumption by cells (OCR) is an indicator of mitochondrial respiration [73]. Additionally, cells generate ATP via glycolysis (the conversion of glucose to lactate) independently of oxygen. Measurement of the lactic acid produced indirectly via protons released into the extracellular medium, which causes acidification of the medium surrounding the cells, provides the extracellular acidification rate or ECAR [73]. Previous studies suggest that maintenance of high reserve mitochondrial capacity is a major factor that defines the vitality and/or survival of cancer cells [84].

To examine the bio-energetic phenotype of TRIM24-iHMECs and control iHMECs, we measured respiration, glycolysis rate and ATP turnover in the cells in response to pharmacological modulators of glucose metabolism (Fig. 13A and B). TRIM24-1 and TRIM24-2 iHMECs exhibited higher basal ECARs and OCRs compared to control

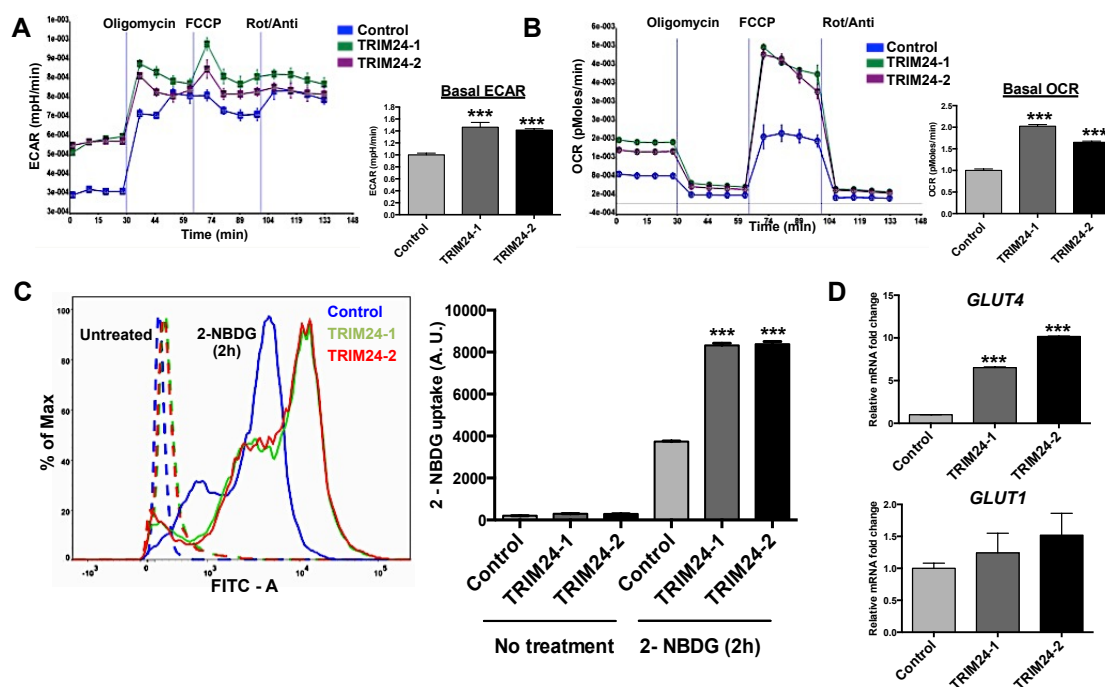


Figure 13: TRIM24 leads to diverse metabolic states and increased glucose uptake.

A and B, Extracellular Acidification rate (ECAR) and Oxygen Consumption Rate (OCR) in the presence of Oligomycin (1 μ M), FCCP (Carbonylcyanide p-trifluoromethoxyphenylhydrazone, 500 nM) and Rot + Ant [Rotenone (1 μ M) + Antimycin A (1 μ M)] measured using an XF^e96 Extracellular Flux Analyzer (Sea-horse) for Control, TRIM24-1 and TRIM24 iHMECs. Both ECAR and OCR were normalized using cell number. C, 2-NBDG (Florescent Glucose) uptake assay for Control, TRIM24-1, and TRIM24-2 iHMECs for 0 hours (untreated) and 2 hours. Florescent intensity curves of the 2-NBDG uptake are shown. D, qRT-PCR analysis of GLUT4 and GLUT1 mRNA expression in Control, TRIM24-1, and TRIM24-2 iHMECs. The data are averages from three biological replicates \pm SD. *** p < 0.001.

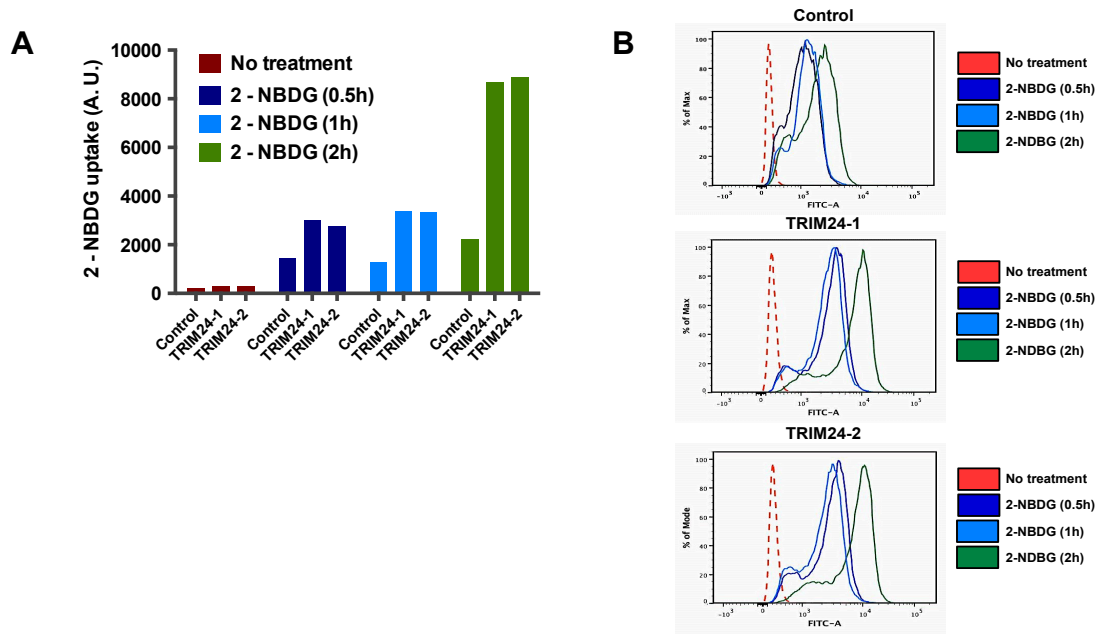


Figure 14: Glucose uptake time course for control vs TRIM24-iHMECs as measured by FACS

A and B, 2-NBDG (glucose) uptake assay for Control, TRIM24-1, and TRIM24-2 iHMECs for 0h (untreated), 0.5h, 1h and 2 h respectively. Bar graphs (A) and Florescent intensity curves (B) of the 2-NBDG uptake are shown.

iHMECs, suggesting increased basal glycolysis and mitochondrial respiration, respectively. We then metabolically perturbed the cells by treating the cells with compounds that shift the bio-energetic profiles of the cells. Oligomycin, an inhibitor of the mitochondrial ATP synthase, triggered a robust increase in lactic acid production (Fig. 13A), expressed as changes in ECAR levels in control and TRIM24-iHMECs. After uncoupling the proton gradient in the electron transport chain from oxidative phosphorylation by treating the cells with FCCP, TRIM24-1 and TRIM24-2 iHMECs had much higher reserve mitochondrial capacities than did control iHMECs (Fig. 13B). With these effects on metabolism, TRIM24-iHMECs are more responsive to stress under conditions of increased energy demand, compared to control iHMECs. Taken together, our findings suggest that ectopic expression of TRIM24 in iHMECs leads to increased basal glycolysis and basal mitochondrial respiration in parallel with an increased reserve of mitochondrial capacity.

Next, we studied whether the altered bio-energetic states of TRIM24-iHMECs result in increased glucose uptake by these cells. We compared control and TRIM24-iHMECs using the fluorescent 2-deoxyglucose analog (2-NBDG) staining. Our results indicated higher glucose uptake in both TRIM24-1 and TRIM24-2 iHMECs, compared to control iHMECs, further confirming the increased glycolytic state of TRIM24-transformed cells. (Fig. 13C and Fig. 14A-B). Consistent with this, we observed significantly increased glucose transporter protein *GLUT4* gene expression and measurable *GLUT1* activation in TRIM24-iHMECs (Fig. 13D).

3.6 TRIM24 causes deregulation of two key players of cancer metabolism

p53 and c-Myc are two important regulators that determine the fate of cellular metabolism. The tumor suppressor p53 regulates energy metabolism by lowering rates of glycolysis and inducing mitochondrial respiration. *GLUT1* and *GLUT4* are direct p53 targets of transcriptional regulation [85]. p53 indirectly represses *GLUT3* by repressing NF- κ B gene expression [86]. p53 induces the expression of *TIGAR* (TP53-inducible glycolysis and apoptosis regulator) and TIGAR significantly slows down tumor glycolysis by converting fructose 2,6-bisphosphate back to fructose 1-phosphate [33]. Fructose 2,6-bisphosphate functions as an allosteric activator of PFK1, a major glycolytic enzyme. Fructose 2,6-bisphosphate is produced by PFK2 from fructose 1-phosphate. Additionally, p53 indirectly represses activation of several glycolytic enzymes by activating miR-34a [87]. The net result of constitutive p53 activation is primarily to direct the glycolytic flux towards pyruvate that can then enter the TCA cycle.

In contrast, the oncogene c-Myc is known to upregulate glycolysis, glutaminolysis and mitochondrial biogenesis in cancer cells. c-Myc directly upregulates the expression of HK2, PFK1, TP11, LDHA and several others in the glycolytic pathway. The majority of glycolytic enzymes contain c-Myc binding motifs in their gene promoters [88]. Two major glutamine reporters, ASCT2 and SN2, are trans-activated by c-Myc in several tumors [89]. GLS1, one of the major enzymes responsible for glutaminolysis, is indirectly upregulated by c-Myc suppressing miR-23a/b [90]. c-Myc regulates expression of several nuclear-encoded genes critical for mitochondrial growth and biogenesis. Mitochondrial transcription factor A (TFAM), a critical regulator of mitochondrial DNA replication and transcription, is trans-activated by c-Myc [91].

Interestingly, in TRIM24-iHMECs there is a significant induction of c-Myc mRNA and protein expression with concomitant decrease in p53 protein levels (Fig. 15). Induction of c-Myc and degradation of p53 in TRIM24-iHMECs may explain the dual induction of glycolysis and TCA cycle in these cells. Whether or not TRIM24 directly regulates expression of genes associated with metabolism or does indirectly by regulating p53 and c-Myc needs to be determined.

3.7 TRIM24 regulates expression of several genes associated with glucose metabolism in multiple breast cancer lines

We asked whether TRIM24-mediated regulation of glycolytic and TCA-associated genes is widespread amongst various cellular models of breast cancer by assessing multiple, established cell lines derived from breast tumors. Transient knock down of *TRIM24* (siTRIM24) expression in MCF-7 (luminal), MDA-MB-231 (triple-negative), and SKBR3 (*HER2*-positive) indicated that depletion of *TRIM24* down-regulated *GLUT1*, *ACO1*, *IDH1*, *IDH2*, *PGM1*, and *OGDH* expression in MCF-7 cells (Fig. 16A & Fig. 17). Moreover, expression of *GLUT1*, *ACO1*, *IDH1*, and *IDH2* was likewise decreased in MDA-MB-231 and SKBR3 cells treated with TRIM24 siRNA, suggesting that regulation of specific metabolic genes by TRIM24 is conserved, albeit to varying levels. We then checked for expression of the aforementioned metabolic genes in the isogenic panel of HMECs. Interestingly, we found that the endogenous expression pattern of *ACO1* and *IDH2* parallels endogenous TRIM24 expression and follows an upward trend along the transformation process, and *OGDH* and *HK2* gain highest expression in the immortalized malignant HMECs (Fig. 16B). Taken together, these results suggest that

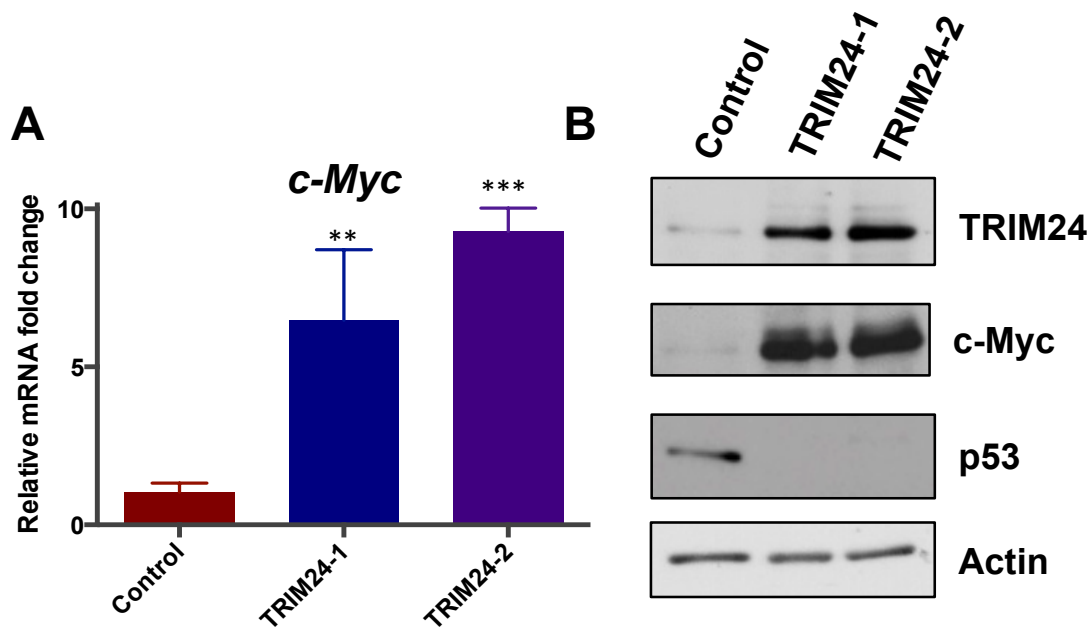


Figure 15: TRIM24 induces deregulation of two key players of cancer metabolism

A & B, qRT-PCR analysis of cMyc mRNA (A) and Immunoblot of TRIM24, c-Myc and p53 in Control, TRIM24-1, and TRIM24-2 iHMECs. The data are averages from three biological replicates \pm SD. * $p < 0.05$ and ** $p < 0.01$.

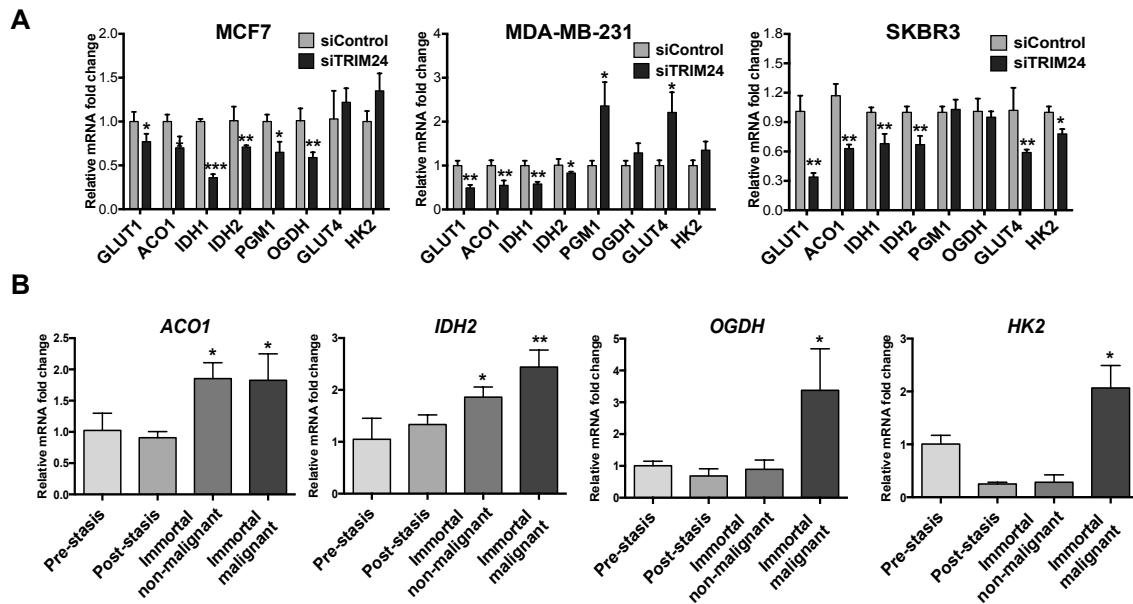


Figure 16: TRIM24 regulates expression of several genes associated with glucose metabolism in multiple breast cancer subtypes

A, MCF-7, SKBR3 and MDA-MB-231 cells were transfected with siControl or siRNA against TRIM24 (siTRIM24) and harvested 48 hours later. Effect of TRIM24 knockdown on expression of a glucose metabolism gene panel (by qRT-PCR analysis) is shown. The data are averages from three replicates \pm SD. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

B, mRNA expression for ACO1, IDH2, OGDH and HK2 in HMEC lines transitioning from normal finite-lifespan pre-stasis cells to abnormal finite post-stasis, non-malignant immortal and malignant cells. The data are averages from three biological replicates \pm SD. * $p < 0.05$ and ** $p < 0.01$.

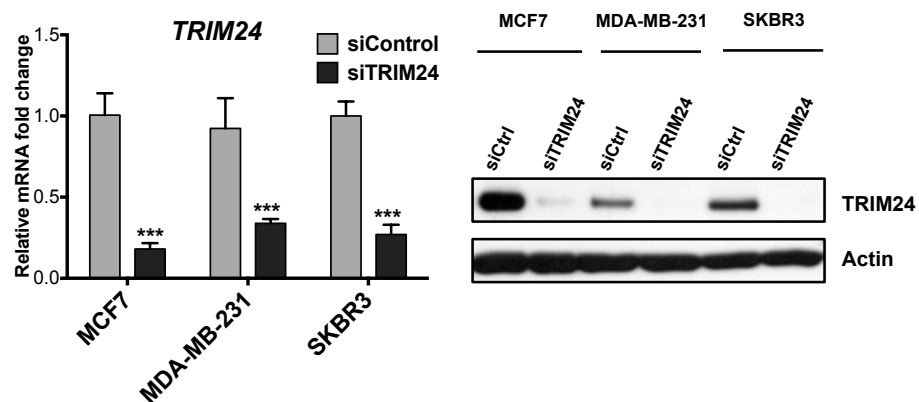


Figure 17: siRNA based TRIM24 knockdown in multiple breast cancer cell lines

TRIM24 RNA and protein levels in various breast cancer cell lines MCF-7, SKBR3 and MDA-MB-231 cells were transfected with siControl or siRNA against TRIM24 (siTRIM24). TRIM24 RNA (by qPCR) and protein expression (by Western) are shown.

TRIM24-mediated deregulation of glucose metabolism is a relatively global phenomenon that occurs in multiple breast cancer subtypes.

3.7 TRIM24-HMECs form intermediate to high-grade tumors in nude mice

We assessed the impact of *TRIM24* expression in iHMECs by *in vivo* xenograft formation. To this end, we injected female nude mice subcutaneously with either Control or TRIM24-iHMECs in 50% high concentration matrigel. TRIM24-1 and TRIM24-2 xenografts displayed significantly higher xenograft volume as compared to their control counterparts (Fig. 18A and Fig. 19A-C). H&E staining showed that xenograft tumors from TRIM24-iHMECs contained intermixed neoplastic epithelial cells and fibrous tissue in various proportions (Fig. 18B, left panel), whereas no epithelial cells were detected in xenografts from Control-iHMECs (Fig. 18C and Fig. 19D-E). TRIM24-expressing, neoplastic epithelial cells formed tumors that reflect a spectrum of low, intermediate and high-grade malignancy (Fig. 18B, left panel). The expression levels of TRIM24 in these tumors correlated with the malignancy grade, as shown by TRIM24-IHC (Fig. 18B, right panel). The grade of malignancy was based on the degree of cellular atypia, anisokaryosis and degree of vascularization (Fig. 19F-H and Appendix 8). Of note, about 60-70% of the tumors formed with TRIM24-iHMECs were either intermediate or high grade (Fig. 18C) with the proportion of epithelial cells in the subcutaneous tumors at 30-40% (Fig. 18D and Fig. 19E), compared to xenografts of Control-iHMECs that consisted of fibrous tissue only (Fig. 19E and Appendix 8). Taken together, these data indicate that over expression of TRIM24 can confer *in vivo* tumorigenic growth of iHMECs.

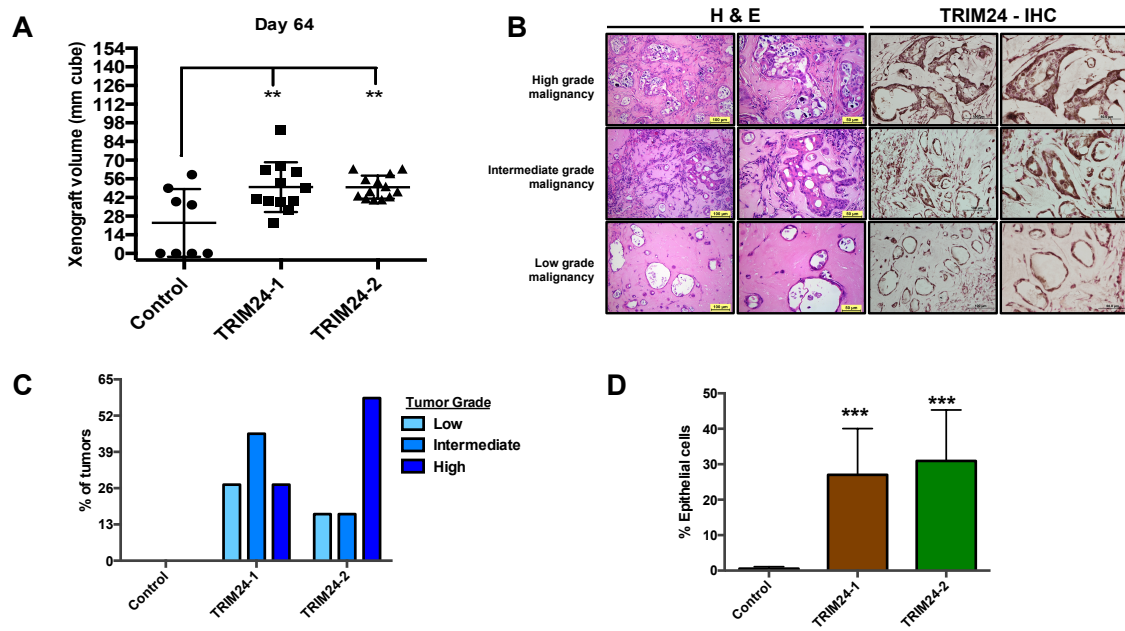


Figure 18: TRIM24-HMECs form intermediate to high-grade tumors in nude mice

A, Tumor volume after 65 days post-injection of Control and TRIM24-iHMECs in nude mice. ** $p < 0.01$ (all compared to Control). B, H&E Staining and TRIM24-IHC for various grades of tumors obtained from nude mice injected with TRIM24-iHMECs. C, Distribution of various grades of tumors from mice injected with either Control or TRIM24-iHMECs. D, Distribution of epithelial cells in tumors from mice injected with either Control or TRIM24-iHMECs.

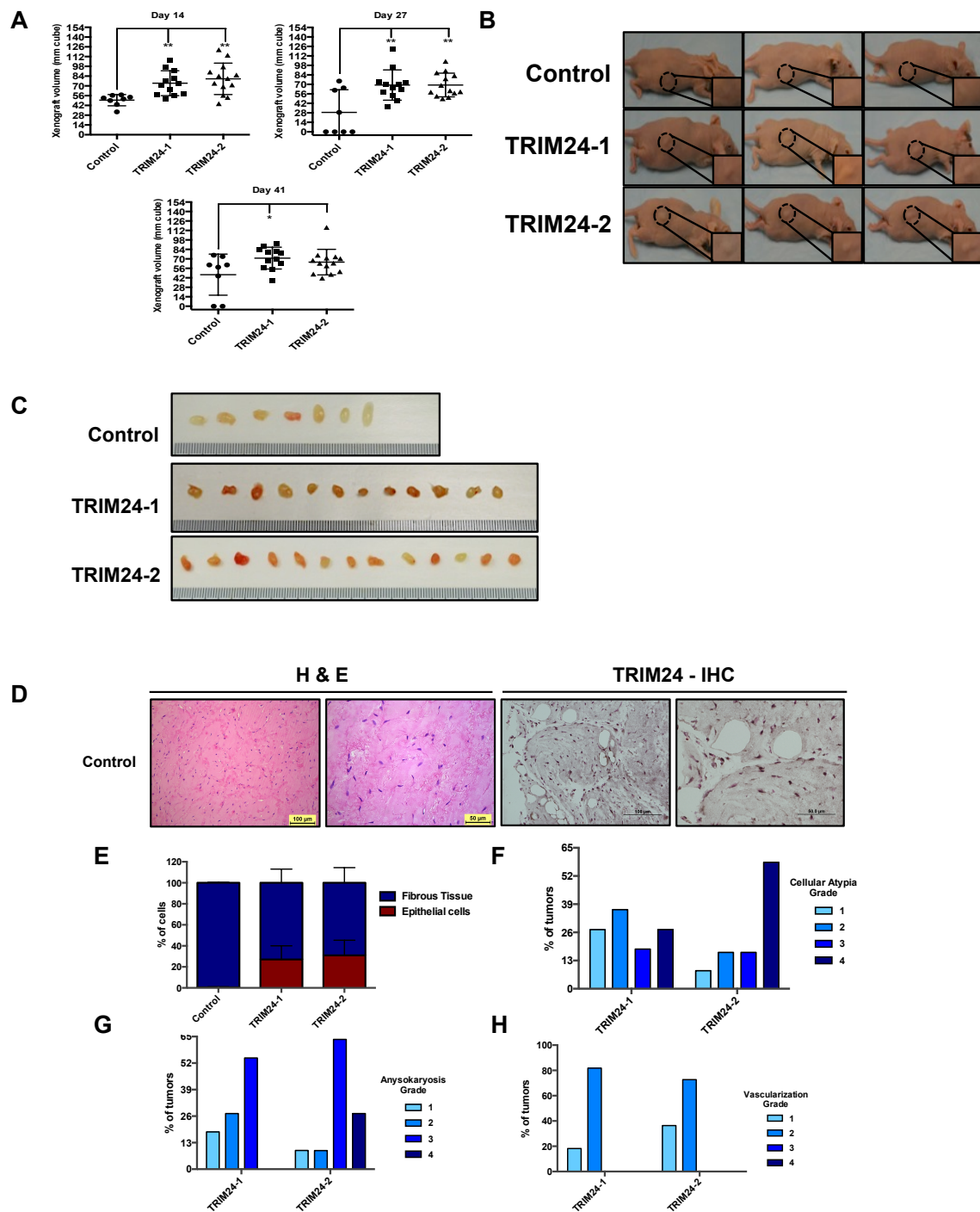


Figure 19: Characterization of xenograft tumors formed by control and TRIM24-HMECs

A, Tumor volume post-injection of Control and TRIM24-iHMECs in nude mice after Day 14, Day 27 and Day 41. * $P < 0.05$, ** $P < 0.01$ (all compared to Control). B, Images

from mice injected with Control or TRIM24-iHMECs (showing tumors). C, Tumors obtained from mice injected with Control or TRIM24-iHMECs

D, H&E Staining and TRIM24-IHC for a representative tumor obtained from mice injected with Control-iHMECs. E, Percentage of Epithelial cells v/s fibrous tissue in tumors obtained from mice injected with Control or TRIM24-iHMECs. F, G and H, Cellular atypia grade, Anysokaryosis grade and Vascularization grade of tumors obtained from mice injected with Control of TRIM24-iHMECs.

3.8 TRIM24 directly regulates expression of various metabolic genes

To elucidate critical targets of TRIM24-induced transformation, previously published TRIM24 ChIP-seq (in MCF7 cells) was intersected with the transcriptional signature obtained after TRIM24-overexpression in iHMECs. Interestingly, several metabolic genes were bound by TRIM24. Recruitment of TRIM24 at several of these metabolic genes was confirmed by TRIM24-ChIP followed by qPCR in MCF7 cells. As shown in Fig. 20, TRIM24 is recruited to the promoters of several metabolic genes such as *GLUT1*, *IDH1* and *IDH2*. Hence, it is clear that *GLUT1*, *IDH1* and *IDH2* are direct TRIM24 targets.

Next, in order to determine which transcription factor(s) may help recruit TRIM24 at these metabolic genes, transcription factor affinity prediction (TRAP) analysis was performed. In TRAP analysis, it was found peroxisome proliferator activated receptor- γ (PPAR γ) was among the top-three transcription factors to be significantly enriched at TRIM24-binding sites (Fig. 21). One of the key players that regulates adipogenesis and whole-body lipid metabolism is PPAR γ [92]. In addition, it is also crucial for controlling gene networks involved in glucose homeostasis including increasing the expression of GLUT4 and c-Cbl associated protein (CAP)[92]. Intriguingly, in breast cancer cells ER- α physically associates with PPAR γ and functionally interferes with PPAR γ signaling [93]. Future studies will investigate the crosstalk between TRIM24, PPAR γ and ER- α by co-immunoprecipitation and ChIP assays to confirm their recruitment and mechanism of action at various metabolic genes.

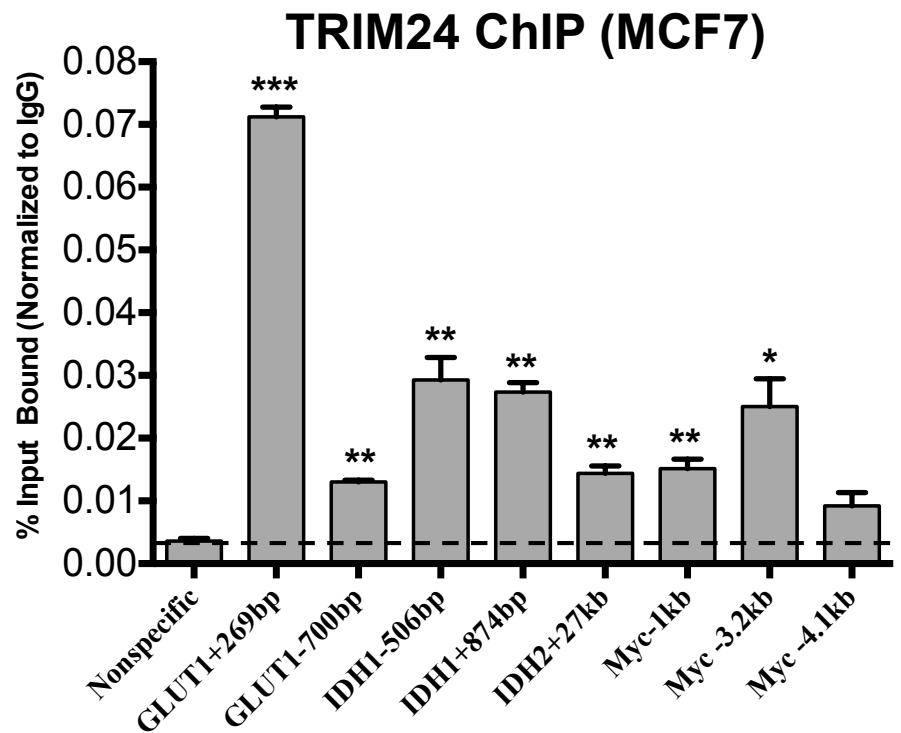


Figure 20: TRIM24 ChIP in MCF7 cells

TRIM24 ChIP confirms recruitment of TRIM24 at various metabolic genes such as *GLUT1*, *IDH1* and *IDH2*, and c-Myc along with previous assessment of gene expression to show direct impact of TRIM24 binding. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ (TRIM24 ChIPs done by Sabrina Stratton).

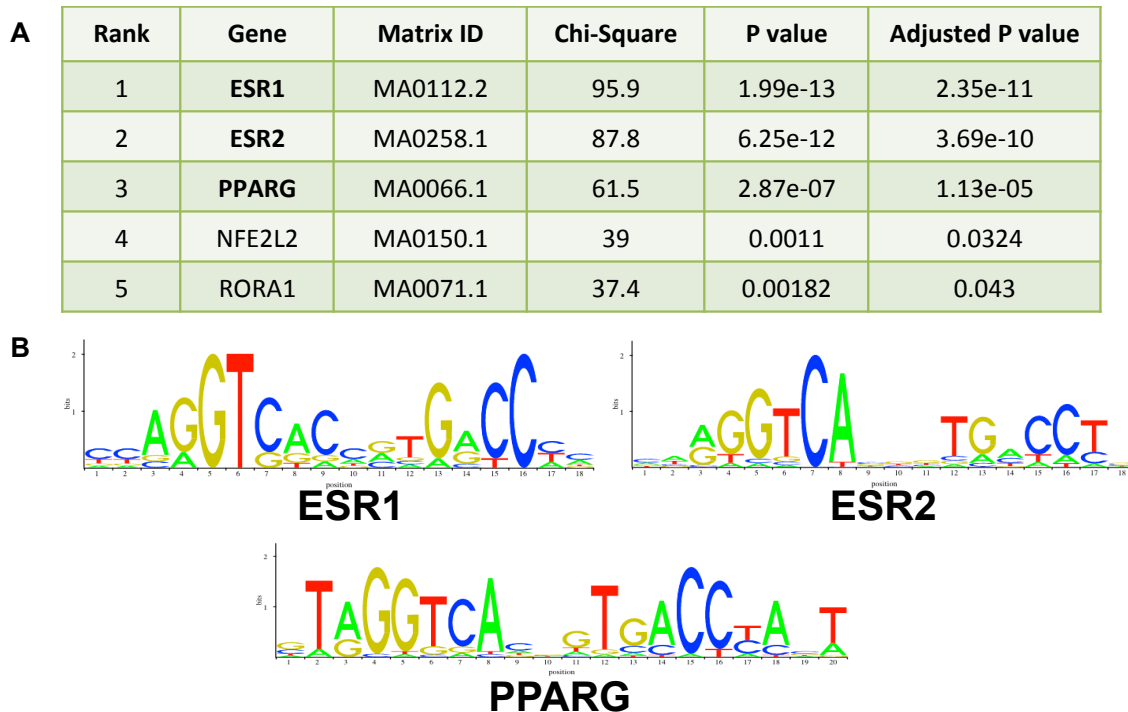


Figure 21: TRAP analysis of TRIM24-binding peaks in MCF7 cells

A. Table showing Top 5 hits of Transcription factor affinity prediction of TRIM24 binding peaks in MCF7 cells. B. Binding motifs of Top 3 hits viz. ESR1, ESR2 and PPARG. (Bio-informatic analysis done by Whijae Roh).

3.8 Effect of TRIM24 over-expression on HMEC cell size and karyotype

Interestingly, H&E staining of TRIM24-iHMECs, showed that TRIM24-iHMECs were much bigger in size (Fig. 22A) and the overall nuclear to cytoplasmic ratio was very high in these cells. Also, when an equal number (4 million) of control and TRIM24-iHMECs were spun down in a 15ml tube, the pellet formed by TRIM24-iHMECs occupied much more area (Fig. 22B).

Since, TRIM24-iHMECs are much bigger in size and exhibit all of the hallmarks of transformed cells, we determined if there was any change in the chromosome number upon TRIM24 overexpression. We performed a karyotype analysis in control and TRIM24-iHMECs and found that the majority of TRIM24-iHMECs were aneuploid with an average of 74-76 chromosomes per cell; the control-iHMECs had a normal karyotype with close to 46 chromosomes per cell (Fig. 23). In both TRIM24-iHMECs clones, increased chromosome copies (tri-, tetra- and penta- somey) and chromosomal rearrangements and translocations were observed with high frequency. A high frequency of mutated chromosomes (M) was also observed in TRIM24-iHMECs.

3.9 Effect of dual TRIM24-BRPF1 Bromodomain inhibitor on glucose metabolism

As discussed previously, TRIM24 contains a tandem PHD and Bromo-domain in the C-terminus that reads a non-canonical histone signature (H3K4me0 and H3K23ac). Because of the druggability of bromodomain-containing proteins, the TRIM24-bromodomain offers a potential therapeutic for epigenetic regulation [94]. As part of the drug discovery program at the MD Anderson Cancer Center, a highly potent and selective dual TRIM24-BRPF1 bromodomain inhibitor (IACS-9571) was developed with an ITC K_d = 31 nM for TRIM24 and ITC K_d = 14 nM for BRPF1 [95].

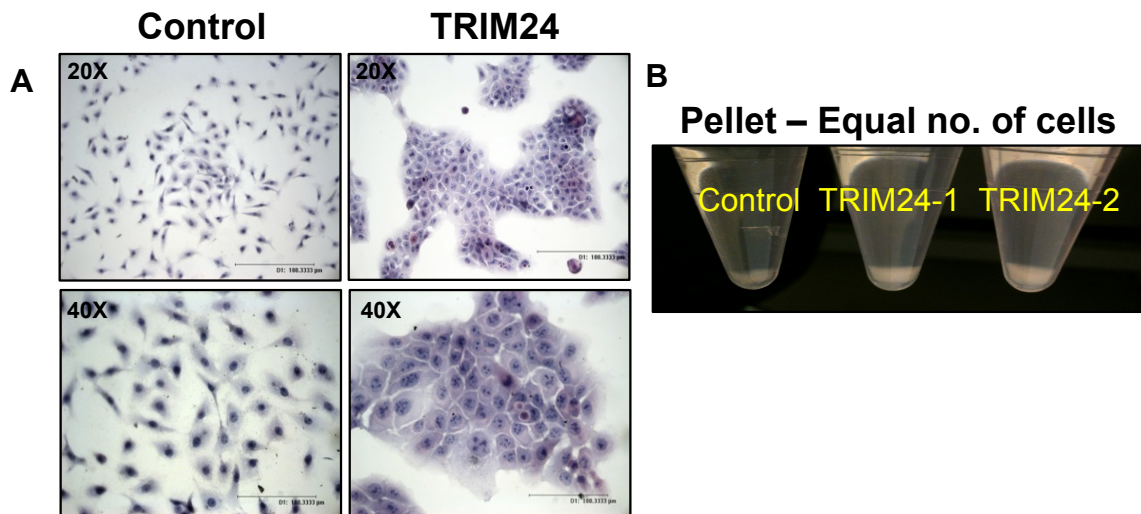


Figure 22: TRIM24-iHMECs are bigger in size

A. Bright light images of H&E staining of control and TRIM24-iHMECs. B. Bright light image of 15ml tube (bottom) after spinning down equal number (4 million) of control and TRIM24-iHMECs.

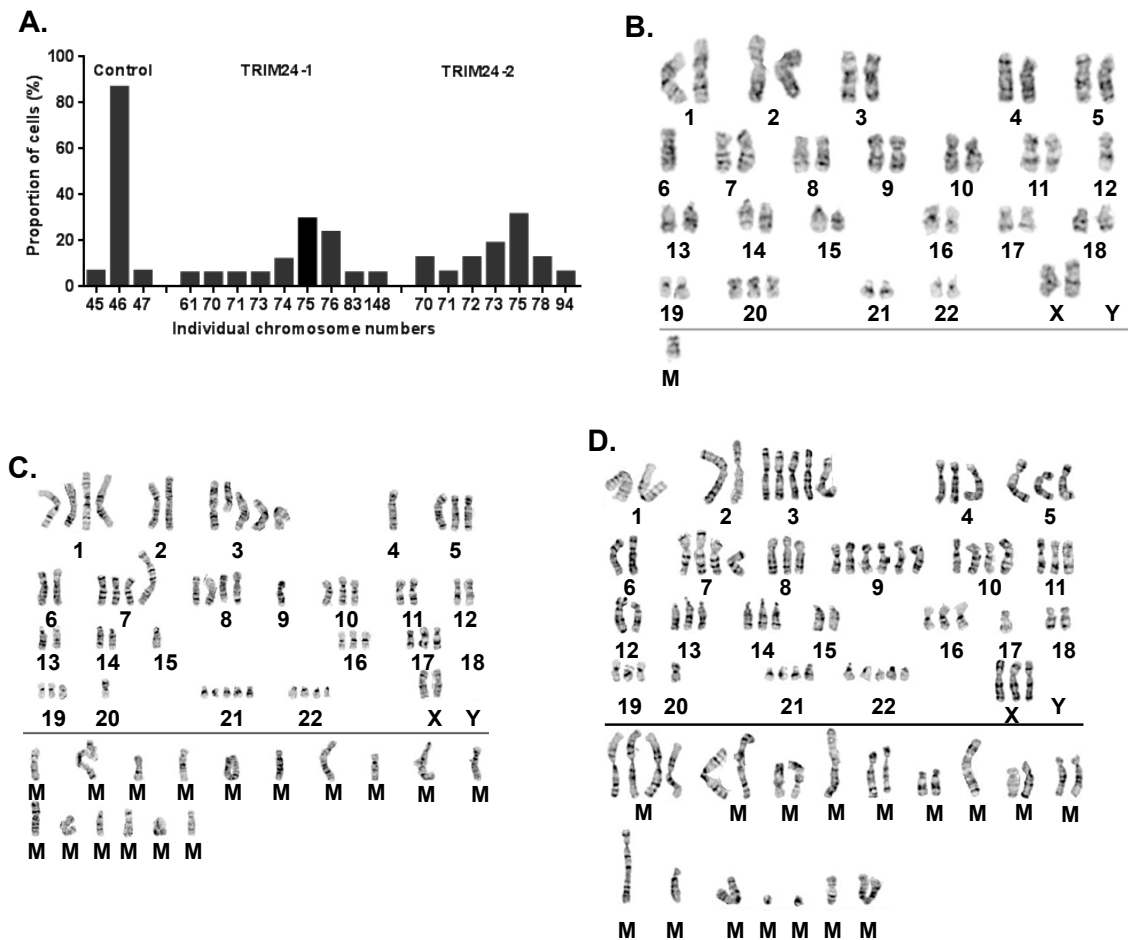


Figure 23: Over-expression of TRIM24 in HMEC stable cell lines causes aneuploidy.

A. Individual chromosome numbers from metaphase spreads of immortalized stable HMEC line in control or stably over-expressing TRIM24 (TRIM24-1, TRIM24-2) were analyzed (n=100). B. Representative images of G-band karyotype in control HMEC, showing the 46 chromosomes with haploid 6 and trisomy 20 that is similar to the previous published data. (C, D) Over-expression of TRIM24 in immortalized HMEC is sufficient to induce chromosome instability (CIN): Representative images of G-band karyotypes in TRIM24-1 and -2.

In order to determine if TRIM24-bromodomain function is critical for regulating glucose metabolism in breast cancer, MCF7 cells were treated with IACS-9571 for 48 h (1 μ M and 5 μ M). The mRNA expression of several metabolic genes was evaluated by RT-qPCR. Interestingly we found that expression of several genes associated with glucose metabolism was downregulated upon treatment with IACS-9571 (Fig. 24). These results suggest that interaction of TRIM24 with chromatin via its bromodomain may be critical for regulating glucose metabolism in breast cancer. However, the effect of IACS-9571 on glucose metabolism needs to be tested in additional breast cancer cell lines. In the future, TRIM24-overexpressing stable cells with a bromodomain chromatin-binding mutant of TRIM24 will be generated and tested for various hallmarks of transformation and changes in glucose metabolism to clearly define the role of the TRIM24 bromo-domain in regulating cancer metabolism.

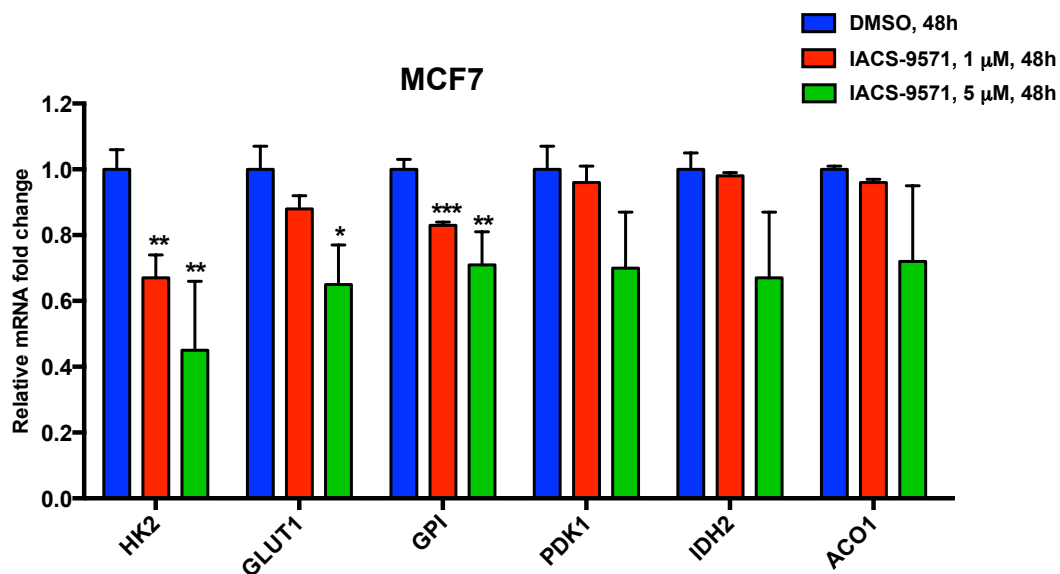


Figure 24: Effect of TRIM24-bromodomain inhibition on glucose metabolism in breast cancer

RT-qPCR data of various glucose metabolism genes after treatment of MCF7 cells with respective concentration of IACS-9571.

Chapter 4: TRIM24 acts as a novel regulator of Epithelial Mesenchymal Transition (EMT) in breast cancer

Chapter 4: TRIM24 acts as a novel regulator of EMT in breast cancer

4.1 Introduction and Rationale

TRIM24 is a multi-domain protein that i) negatively regulates p53 via its RING domain, ii) interacts with estrogen receptor (ER) via an LXXLL motif, and iii) recruits transcription factors to chromatin as a histone reader with PHD-Bromodomains that recognize H3K4me0/H3K23ac [96]. Recently, aberrant over-expression of TRIM24 in cancers of multiple origins, including prostate cancer [47], breast cancer [55, 56], non-small cell lung cancer [57], head and neck carcinoma [58], colorectal cancer [59], gastric cancer [60], glioblastoma [61] and human hepatocellular carcinoma [63] has been reported. My previous work showed that TRIM24 expression caused major shifts in metabolism and promoted transformation of immortalized human mammary epithelial cells [97]. However, a detailed analysis of how TRIM24 promotes tumorigenesis with progression to metastasis has not been elucidated.

Metastasis is multi-step process consisting of migration of cancer cells from the primary tumor, followed by local invasion, intravasation, extravasation and finally colonization of cancer cells at distant organs. Each step of metastasis involves a series of specific cell-to-cell interactions and activation of different signaling pathways [16]. Prior to metastatic progression of a tumor, cancer cells undergo a transition from an epithelial to a mesenchymal phenotype; the process is called the epithelial-mesenchymal transition (EMT). My preliminary analysis shows that TRIM24 is a driver of EMT. During the EMT process, epithelial cells lose cell polarity, disassemble their cell-cell adhesion machinery and acquire cell motility. The EMT process is marked by loss of E-cadherin and acquisition of various mesenchymal markers such as N-cadherin and vimentin, as a result of activation of transcription factors such as Snail,

Slug, Twist and ZEB1/ZEB2 [16]. I hypothesize that aberrant expression of TRIM24 impacts a network of transcription regulatory factors and signaling pathways to induce EMT and metastasis in breast cancer.

4.2 TRIM24 induces a mesenchymal-phenotype in breast epithelial cells

MCF10A cells exhibit multiple characteristics of normal breast epithelium such as an absence of anchorage-independent growth, hormone-dependent and growth factors for survival and proliferation and absence of tumorigenicity in xenograft studies in mice [68]. To further understand the mechanism of TRIM24-mediated oncogenesis, TRIM24 (FLAG-tagged, wild-type) was stably overexpressed in MCF10A. Overexpression of TRIM24 was confirmed by RNA (RT-qPCR) and protein (immunoblotting) analyses (Fig. 25A). Interestingly, TRIM24-expression substantially altered the morphology of MCF10A cells, resulting in decreased clustering, loss of cell-cell contacts and increased mesenchymal-like morphology (Fig. 25B).

In order to determine if TRIM24-overexpression induced EMT, RNA and protein expression analyses of known EMT markers were performed. Gene and protein expression analysis revealed an EMT signature, specific to TRIM24-overexpressing MCF10A cells versus vector only cells, marked by decreased E-cadherin (*CDH1*), and increased N-cadherin (*CDH2*), Fibronectin-1 (*FN1*, *mRNA*), Snail Family Zinc Finger 1 (*SNAI1*, protein), Snail Family Zinc Finger 2 (*SNAI2* aka *Slug*, *mRNA*) and Vimentin (*VIM*, protein) (Fig. 26C-D). Loss of E-cadherin (*CDH1*) in TRIM24-overexpressing MCF10A cells was further confirmed by immunofluorescence staining. As shown in Fig. 24E, TRIM24-OE MCF10A cells have no detectable expression of CDH1 (red), whereas vector MCF10A cells show very high expression of CDH1.

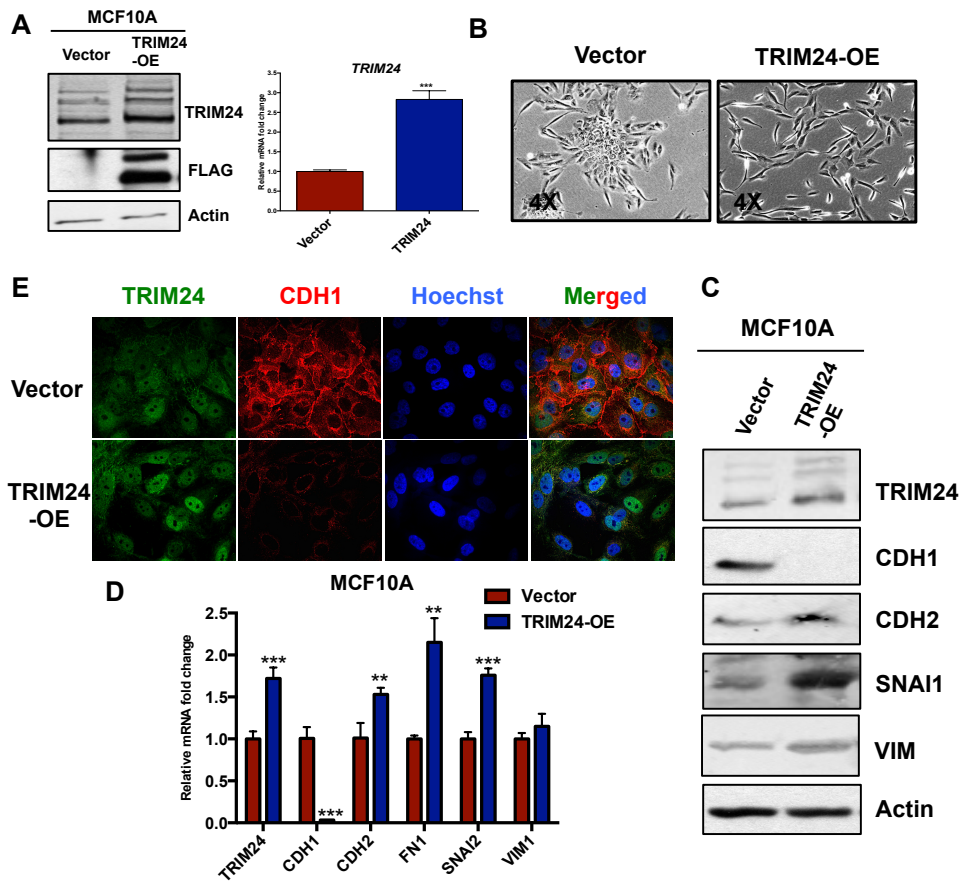


Figure 25: TRIM24 induces EMT signature in breast epithelial cells

A, qRT-PCR analysis of TRIM24 mRNA and Immunoblot of TRIM24 and actin in vector and TRIM24-OE MCF10A cells. B, Bright light images of vector and TRIM24-OE MCF10A cells. C & D, qRT-PCR and Immunoblot analysis of TRIM24, CDH1, CDH2, SNAI1, VIM and Actin in vector and TRIM24-OE MCF10A cells. E, Immunofluorescence analysis of TRIM24 (green) and CDH1 (red) in vector and TRIM24-OE MCF10A cells. The data are averages from two replicates \pm SD. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

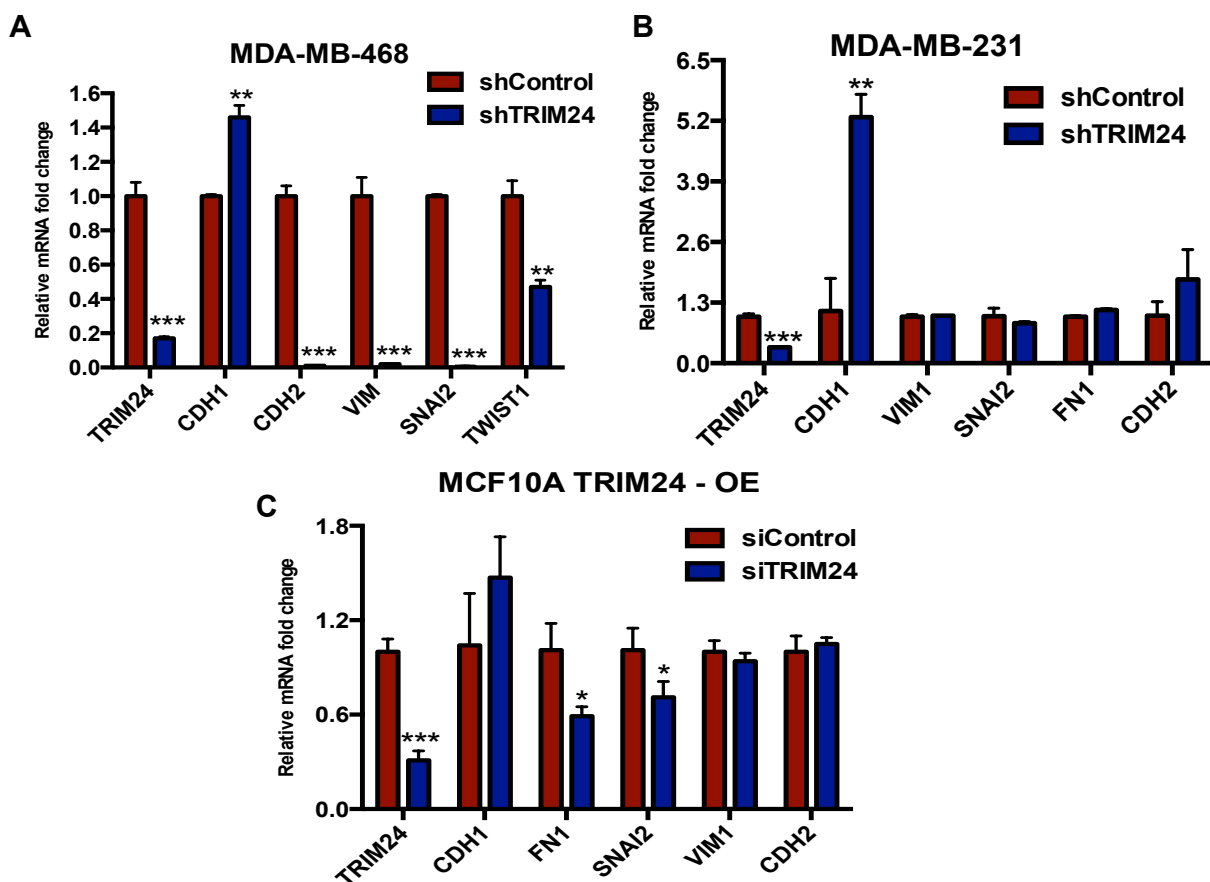


Figure 26: TRIM24 regulates EMT-target gene expression in breast epithelial cells

A, B & C, qRT-PCR analysis of TRIM24, CDH1, FN1, SNAI2, VIM1 and CDH2 mRNA in MDA-MB-468 (A) and MDA-MB-231 (B) – shControl and shTRIM24, and MCF10A TRIM24-OE (C) – transfected with siControl or siTRIM24, respectively. The data are averages from two replicates \pm SD. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

The effects of TRIM24-mediated regulation of EMT markers were also observed after stable depletion of TRIM24 (shTRIM24) in triple-negative breast cancer cells (MDA-MB-468 and MDA-MB-231) (Fig. 26A-B). Interestingly, in MDA-MB-231 there was significant upregulation of *CDH1* upon TRIM24 knockdown, however no significant changes in other EMT markers were observed. Notably, siRNA mediated knock down of TRIM24 in MCF10A cells over-expressing TRIM24 reversed this effect, as shown by reduced expression of *FN1*, *SNAI2* and slightly increased expression of *CDH1* (Fig. 26C).

In order to determine if TRIM24 expression functionally impacts epithelial cell migration, Boyden-chamber based migration assays were performed. As expected, TRIM24-OE MCF10A cells had significantly higher migration as compared to Control MCF10A cells and consistent with this there was significant reduction in migration of shTRIM24 MDA-MB-231 cells as compared to shControl MDA-MB-231 cells (Fig. 27A-B).

4.3 EMT-related pathways are significantly enriched upon TRIM24-overexpression in MCF10A cells

To understand the mechanisms of TRIM24-induced transformation of MCF10A, we analyzed the gene expression profiles of 2560 cancer-associated genes using the HTG EdgeSeq Oncology biomarker assay and identified 501 genes (20%) whose expression was significantly altered, compared to vector only cells. TRIM24 overexpression resulted in activation of 223 (44%) and repression of 278 (56%) genes (Fig. 28A). For a complete list of deregulated genes, please see Appendix 9. Molecular signature hallmark analysis (GSEA) revealed that TRIM24-induced expression of genes altered

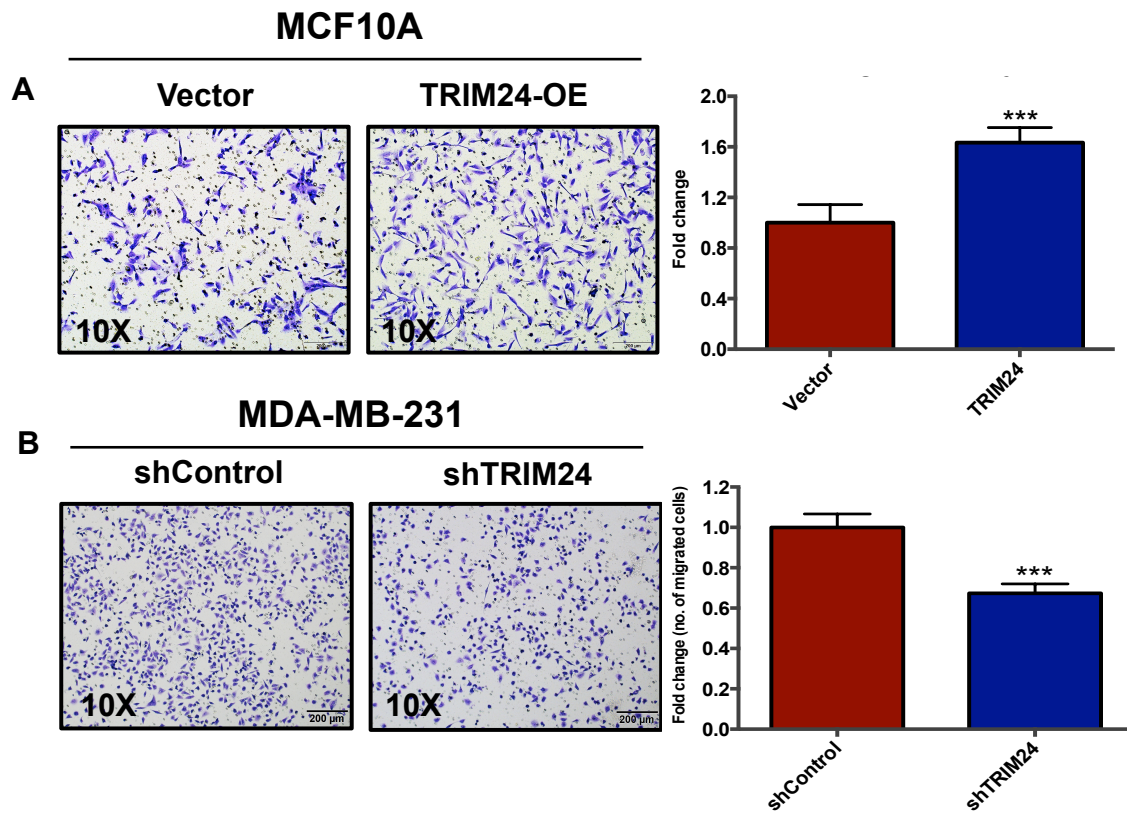


Figure 27: TRIM24 regulates migration of breast cancer cells

A & B, Boyden chamber-based migration assay of MCF10A (vector and TRIM24-OE, A) and MDA-MB-231 (shControl and shTRIM24, B). The data are averages from two replicates \pm SD. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

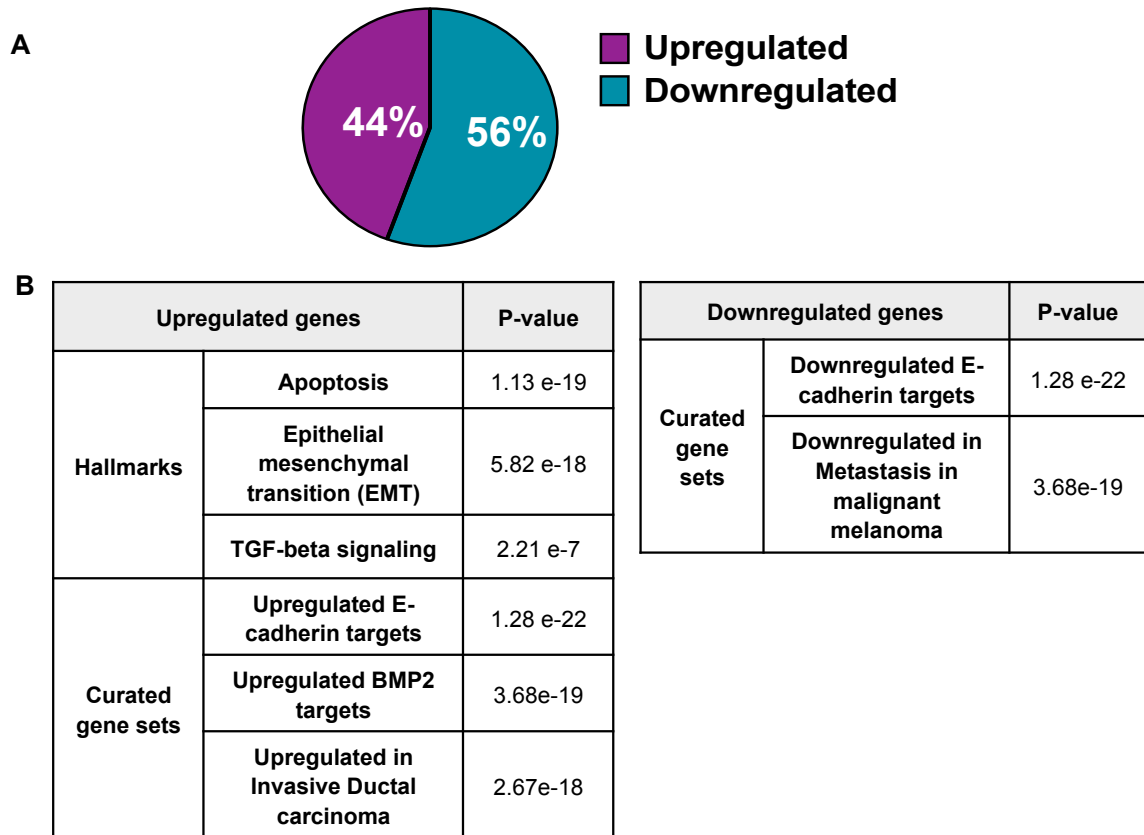


Figure 28: EMT-related pathways are significantly enriched in TRIM24-OE MCF10A cells

A, Pie-chart showing percentage of up-regulated and down-regulated genes upon TRIM24 over-expression as compared to Vector in MCF10A cells. B, Molecular Signature database-based pathway analysis of up-regulated and down-regulated genes upon TRIM24 over-expression as compared to Vector in MCF10A cells.

various hallmarks including apoptosis, epithelial mesenchymal transition (EMT) and TGF-beta signaling (Fig. 28B). Interestingly, TRIM24-activated genes include genes upregulated upon E-cadherin knockdown in HMLE (human mammary epithelial cells) cells, genes upregulated by BMP2 and genes upregulated in invasive ductal carcinoma (IDC) by using curative gene set search of molecular signature database. Similarly, TRIM24-repressed genes include genes downregulated upon E-cadherin knockdown in HMLE cells and genes downregulated in metastatic melanoma (Fig. 28B). For a complete list of enriched pathways, please see Appendix 10.

4.4 TRIM24 directly binds to several EMT-target genes

To elucidate critical targets of TRIM24-induced transformation, previously published TRIM24 ChIP-seq data (from MCF7 cells) was intersected with the transcriptional signature obtained after TRIM24-overexpression in MCF10A. Interestingly, several differentially regulated EMT genes were bound by TRIM24. Recruitment of TRIM24 at several of these EMT genes was confirmed by TRIM24-ChIP followed by qPCR in MDA-MB-468 cells. As shown in Fig. 29A, TRIM24 is recruited at several EMT genes such as *CDH1* and *MMP2*.

As previously mentioned TRIM24 via its C-terminal Bromo domain acts as a histone reader and binds with highest affinity to an H3K23ac signature but also has the capacity to bind H3K9me2 signature [55]. To confirm the presence of an H3K23ac histone signature at TRIM24 bound sites on EMT genes, ChIP analysis of H3K23ac was performed. As shown in Fig. 29B, an H3K23ac signature is enriched at several EMT genes such as *CDH1* and *MMP2* at TRIM24-bound sites. Hence, it is clear that

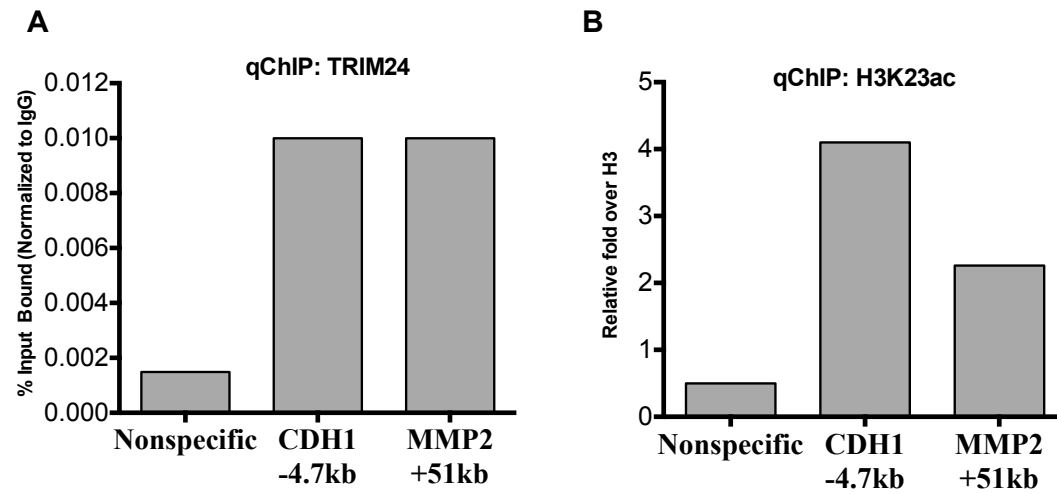


Figure 29: TRIM24 ChIP in MDA-MB-468 cells

A & B, TRIM24 and H3K23Ac ChIP confirms recruitment of TRIM24 at various EMT genes such as *CDH1* and *MMP2* along with previous assessment of gene expression to show direct impact of TRIM24 binding. (TRIM24 ChIP done by Roxsan Manshouri).

CDH1 and MMP2 are direct TRIM24 targets and bromo-domain dependent histone reader function may be important for recruitment of TRIM24 at these EMT genes.

4.5 TRIM24 over-expression inhibits acini formation

Primary mouse and human mammary epithelial cells, when grown on basement membrane gels, exhibit morphogenic changes to form polarized structures with a hollow lumen known as acini. Engelbreth-Holm-Swarm (EHS) tumor or matrigel (BD Biosciences) basement membrane is composed of matrix components such as laminin, collagen IV and entactin. MCF10A cells when cultured in matrigel-based 3D culture for about 10-12 days form acinar structures and retain several properties of glandular epithelial cells found *in vivo*. They exhibit extremely low levels of cell proliferation and are highly consistent with respect to size of acinar and cell number (8,9). Inhibition of cell proliferation and disruption of morphogenic events during acini formation is an important characteristic to investigate the effects of oncogenes and/or tumor suppressors on MCF10A morphogenesis.

Strikingly, when the TRIM24-OE MCF10A were subjected to 3D culture on growth-factor reduced matrigel they fail to form acini, whereas the vector cells formed distinct acini by Day 10 (Fig. 30A). By confocal microscopy analysis, it was further confirmed that TRIM24-OE MCF10A cells have very low expression of CDH1, whereas the vector cells have distinct CDH1 expression at cell-to-cell boundaries (Fig. 30B). As shown here, TRIM24-OE cells have very low expression of CDH1 and unlikely able to build proper cell-to-cell adhesion contacts, thus affecting MCF10A morphogenesis to form acinar structures. In future, CDH1 rescue studies will be performed to determine if CDH1 over-expression is sufficient to rescue the loss of acinar phenotype in TRIM24-

OE MCF10A cells or if a more complex network of TRIM24-dependent gene targets are involved.

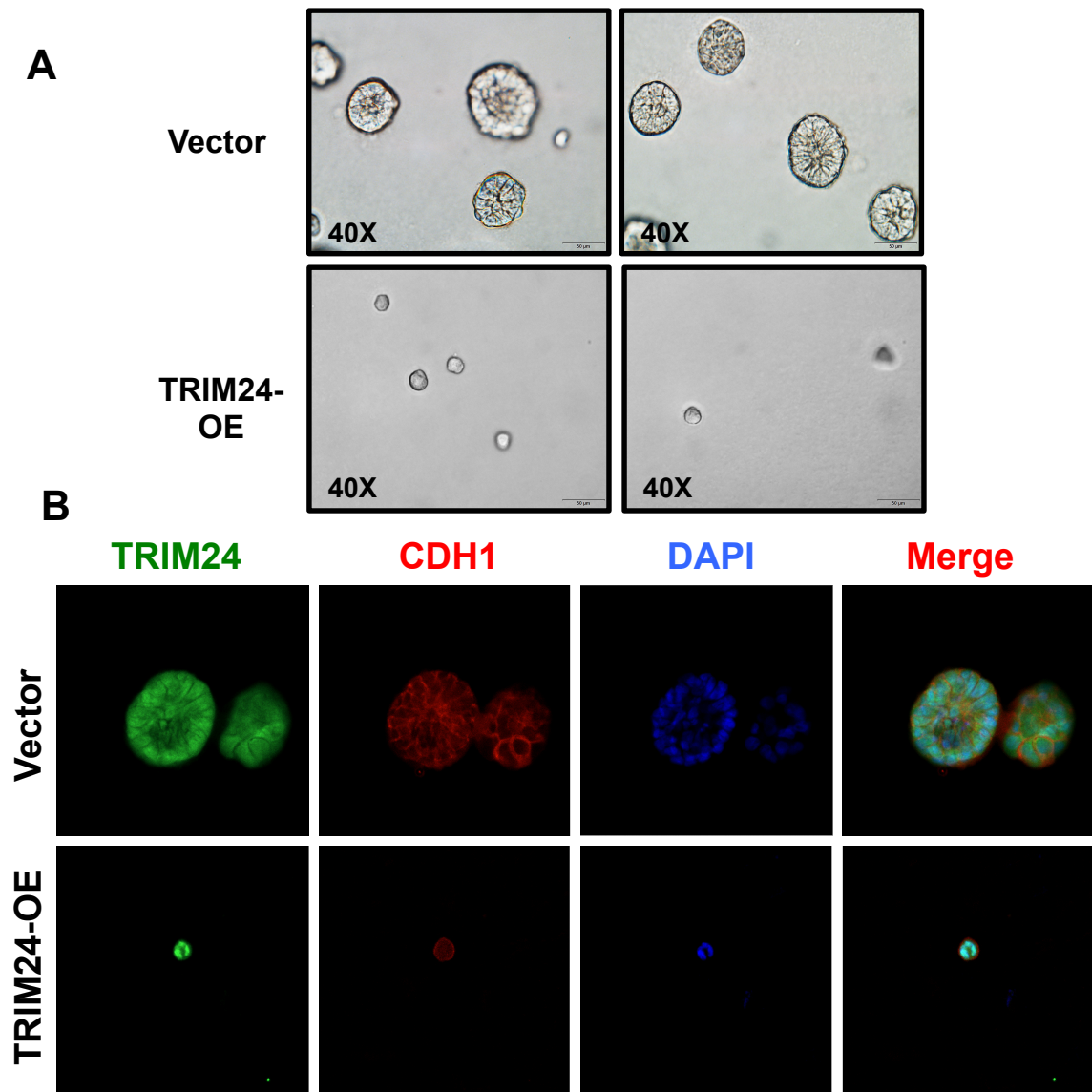


Figure 30: TRIM24 over-expression suppresses acini formation

A, Bright field images of acini formed by Vector and TRIM24-OE MCF10A cells.

B, Confocal images of Immunofluorescence for TRIM24 and CDH1 in acini formed by Vector and TRIM24-OE MCF10A cells (Confocal microscopy done by Erin Williams).

Chapter 5: Discussion and Future directions

Chapter 5: Discussion and Future directions

5.1 TRIM24 - a keystone to cancer metabolism

TRIM24 over-expression in immortalized human mammary epithelial cells induces oncogenic transformation coupled with an array of transcriptional and functional effects. In association with TRIM24-induced breast tumorigenesis, there is deregulation of multiple cancer related pathways including glucose metabolism. To the best of our knowledge, this is the first report of recognition of TRIM24 as one of the prime regulators of cancer metabolism. Over-expression of TRIM24 induces malignant transformation of iHMECs and when these cells are injecting sub-cutaneously into nude mice, a majority of tumors formed by TRIM24-iHMECs were either intermediate or high grade. Interestingly, tumors formed by TRIM24-iHMECs had about 40% epithelial cells with abnormally larger nuclei and this significantly correlated with TRIM24 expression.

In addition to glucose metabolism multiple growth-stimulating pathways such as the insulin [98], ErbB [99] and MAPK [100] that are linked with breast tumor development and progression were also significantly induced upon TRIM24-overexpression in iHMECs. The induction of both glycolysis and TCA cycle in TRIM24-iHMECs was confirmed by mRNA and protein analysis in addition to increased glucose uptake. Of note, Sea-horse analysis also showed that the Extracellular acidification rate (ECAR is rate of glycolysis) and Oxygen consumption rate (OCR is rate of TCA cycle) were higher in TRIM24-iHMECs. To determine whether a correlation between TRIM24 expression and various metabolic pathways existed, a Gene set enrichment analysis (GSEA) using gene expression datasets of invasive breast tumors (n =1008) was performed. The glucose transport pathway was identified as one of the top ten

pathways that correlated positively with TRIM24 expression in invasive breast carcinoma patients from the TCGA database.

One of the key emerging hallmarks of cancer is metabolic reprogramming [26]. Otto Warburg reported that cancer cells intake high amounts of glucose and produce higher lactate even in the presence of oxygen, the phenomenon was termed as 'aerobic glycolysis' or Warburg effect [27]. Although it is quite clear that epigenetic, oncogenic and tumor suppressor mechanisms play a crucial role in regulating metabolic reprogramming of cancer cells, the key molecules involved in regulation of cancer metabolism and their role in oncogenesis are not very well understood. As mentioned previously, TRIM24 overexpression in iHMECs caused induction of not only glycolysis but also the TCA cycle pathway. Although this is in disagreement with the conventional Warburg effect, similar to TRIM24 iHMECs there are several reports of certain cancer cells that preferentially use only OXPHOS or a mix of glycolysis and OXPHOS to produce energy [101]. Several recent reports provide ample genetic evidence that mitochondria are indeed essential for tumorigenesis [34]. It is hypothesized that tumors are metabolically heterogeneous. A majority of tumor cells, exhibit aerobic glycolysis (Warburg effect), however, a small population of slow-cycling tumorigenic cells are dependent on OXPHOS activity. These slow-cycling cells are proposed to be the cells that are resistant to various cytotoxic treatments [35].

The schematic in the Figure 31 depicts how we think TRIM24 might regulate metabolism in breast cancer. As shown previously, TRIM24 is over-expressed in breast cancer and TRIM24 targets p53 for proteasomal-mediated degradation. TRIM24 is recruited to promoters of several metabolic and cell proliferation related genes such as GLUT1, IDH1/2 and c-Myc and it induces their gene expression. Since p53 is degraded it can no longer suppress the expression of glucose transporters. Increased GLUT1

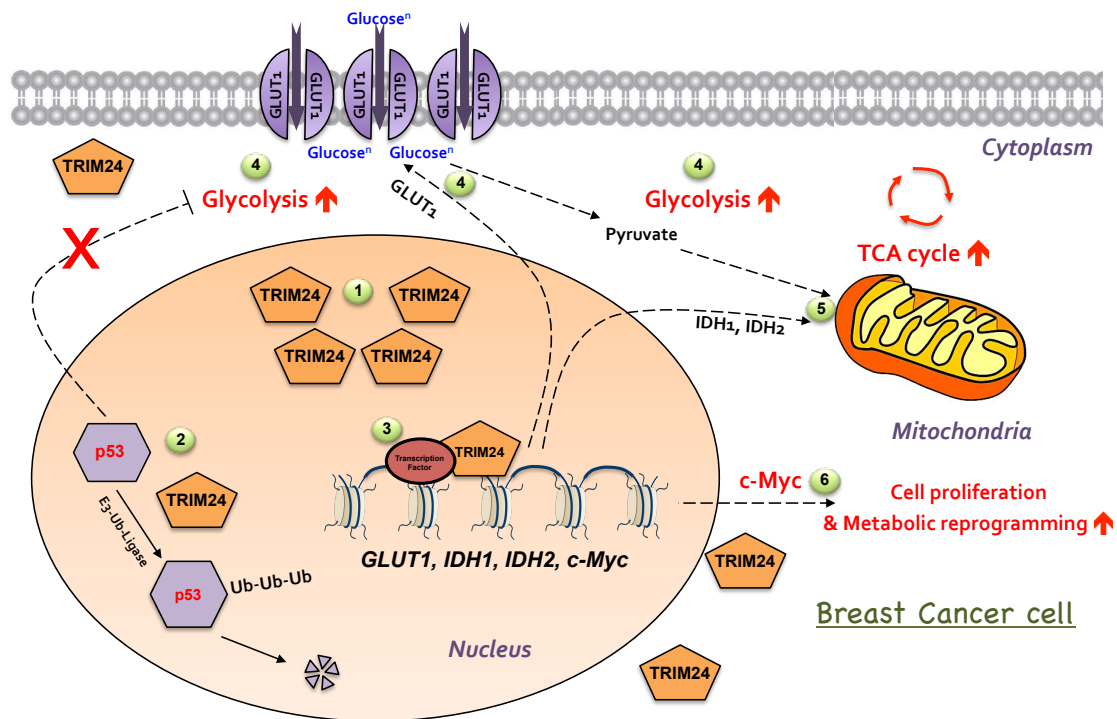


Figure 31: Schematic for role TRIM24 in regulating cancer metabolism in breast cancer

(1) TRIM24 is over-expressed in breast cancer, (2) TRIM24 targets p53 for degradation, (3) TRIM24 is recruited to promoters of metabolic and cell proliferation related genes (GLUT1, IDH1 & c-Myc) and it induces their expression, (4) p53 can no longer block Glycolysis & Increased GLUT1 expression causes increased glucose uptake and Glycolysis, (5) Increased IDH1/IDH2 causes increase in TCA cycle, (6) Increased c-Myc has multiple effects including increased glycolysis and TCA cycle and increased cell proliferation, ultimately leading to breast cancer.

expression causes increased glucose uptake and glycolysis. Increased IDH1/IDH2 causes increase in TCA cycle. Higher c-Myc levels have multiple effects including increased glycolysis and TCA cycle and increased cell proliferation, ultimately leading to breast cancer.

It is plausible that abnormal TRIM24 expression in cancer might affect cellular homeostasis by virtue of its epigenetic functions. Research has shown that cross talk between cellular metabolism and epigenetics might be very crucial in cancer progression [102]. The metabolic status of a cell is directly affected by epigenetic alterations that regulate gene expression of enzymes associated with metabolism, thus contributing to metabolic reprogramming in cancer. On the other hand, almost every known epigenetic enzyme relies on metabolites as cofactors or substrates, such as acetyl CoA. Thus, metabolic reprogramming in cancer can affect epigenetic status and ultimately alter the gene expression of oncogenes as well as tumor suppressors [102].

Of note, several metabolic enzymes are reportedly mutated in various cancers. One prominent example of cancer-associated mutation in metabolic enzymes is IDH1/IDH2 mutations found frequently in low-grade gliomas, that exhibit a gain of function by catalyzing the conversion of α -ketoglutarate (α -KG) to 2-hydroxy glutarate (2-HG), resulting in 100-fold higher intracellular concentration of 2-HG as compared to wild-type cells. 2-HG is a competitive inhibitor of α -KG dependent dioxygenases, including TET2 and the Jumonji C domain containing histone demethylases [103]. Further, various metabolic enzymes are mutated in several cancers for e.g. fumarate dehydrogenase (FH), succinate dehydrogenase (SDH) and phospho-glycerate dehydrogenase (PHGDH) [102].

A key question in the cancer metabolism field is whether an alteration in metabolism is the cause or consequence of oncogenic transformation. In the current

study, we show that TRIM24 induces oncogenic transformation of iHMECs in concert with metabolic adaptations and it is plausible that TRIM24-mediated metabolic reprogramming is more likely a driver event in breast tumorigenesis. Given the enhanced expression profile of TRIM24 not only in breast cancer but also in various other cancers, it is expected that loss of TRIM24 would oppose cancer development in mouse models. Surprisingly, it was found that complete or liver specific loss of TRIM24 in mouse models causes hepatocellular carcinoma. In the TRIM24 null mice, there was increased expression of lipase and inflammatory signaling genes, along with repression of de novo lipogenesis, steroid and lipid metabolism, and lipid transport. The TRIM24 null phenotype of increased inflammation, fibrosis and lipid accumulation in the liver was very similar to human non-alcoholic fatty liver disease (NAFLD) and non-obese non-alcoholic steatosis (NASH).

These contrasting phenotypes may be a result of differential interaction of TRIM24 with tissue-specific transcriptional complexes and target genes and/or tissue-specific response to deregulated p53 levels. In future, these questions may be answered by generating conditional TRIM24-over expression mouse models (breast or liver specific) along with mechanistic studies to determine key functional domains of TRIM24 and transcriptional complexes critical in order to orchestrate metabolic reprogramming by TRIM24.

5.2 Is the molecular interaction between TRIM24 and PPAR γ critical for metabolic reprogramming in breast cancer?

As discussed previously, upon performing transcription factor affinity prediction (TRAP) analysis of TRIM24-ChIP binding sites it was found that Peroxisome

Proliferator Activated Receptor- γ (PPAR γ) was among the top-three enriched transcription factors (Fig. 21). PPAR γ is among the key players that regulate adipogenesis and whole-body lipid metabolism [92]. It regulates glucose homeostasis by inducing GLUT4 and c-Cbl associated protein (CAP) expression [92]. Its role in breast cancer is highly controversial. Based on studies in various breast cancer cell lines and mouse models, certain reports propose that PPAR γ suppresses cell proliferation, induces differentiation and programmed cell death. In DMBA-induced breast cancer mouse models, PPAR γ agonists have been reported to block breast cancer [104]. In contrast, when PPAR γ transgenic mice (breast-specific) are crossed with MMTV-neu mouse model, they display severely accelerated rate of breast cancer propensity [105].

Intriguingly, in breast cancer cells ER- α physically associates with PPAR γ and functionally interferes with PPAR γ signaling [93]. As discussed previously, TRIM24 binds ER- α and chromatin (at specific histone modifications mentioned above) to activate estrogen-dependent genes associated with tumor development and cell proliferation [55]. Future studies will investigate the crosstalk between TRIM24, PPAR γ and ER- α by co-immunoprecipitation studies in breast cancer cells. TRIM24 and PPAR γ ChIP assays will be performed to confirm their co-recruitment and mechanism of action at various metabolic genes. Effects of PPAR γ knockdown and over-expression on TRIM24-induced metabolic reprogramming in breast cancer cells will be assessed. Ultimately, these experiments will address if PPAR γ and TRIM24 association or dissociation is critical to induce metabolic reprogramming in breast cancer.

5.3 TRIM24 a novel regulator of EMT in breast cancer

Our preliminary analysis shows that TRIM24 is a driver of EMT in breast epithelial cells. Upon TRIM24 over-expression the MCF10A cells, exhibit mesenchymal-like morphology with decreased clustering and loss of cell-to-cell contacts. The TRIM24 over-expressing MCF10A cells exhibited various markers consistent with EMT program, i. e. loss of CDH1 and increased expression of CDH2, VIM1, FN1, SNAI1 and SNAI2. Similarly, upon TRIM24 knockdown in several triple-negative breast epithelial cells a suppression of EMT program consistent with decrease in migration is observed.

Gene expression profiling of 2560 cancer-associated genes in TRIM24-OE MCF10A cells revealed that various hallmarks of cancer including apoptosis, epithelial mesenchymal transition and TGF-beta signaling were significantly altered. Molecular signature pathway analysis of TRIM24-activated genes revealed that these genes were upregulated in invasive ductal carcinoma; whereas TRIM24-repressed genes were among the genes that were downregulated in metastatic melanoma. In order to uncover the epigenetic mechanism of EMT regulation, we determined if TRIM24 was recruited to EMT genes. We found that TRIM24 is directly recruited to several EMT genes such as CDH1 and MMP2 and the TRIM24-bound sites have H3K23ac signature at these sites by ChIP analyses.

A major contributor of EMT is E-cadherin promoter methylation, resulting in suppression of E-cadherin expression, a key EMT signature. Amongst other mechanisms, methylation of histone H3 on lysine 9 (H3K9me2) by G9a is essential for DNA methylation of E-cadherin promoter in multiple breast cancer models [106]. Intriguingly, our laboratory has shown that the PHD and Bromo domains of TRIM24 can bind to H3K9me2 modification by performing *in vitro* GST-pull down assays [55].

Additionally, based on TRIM24-pulldown and mass-spectrometry analysis in MCF7 cells, TRIM24 interacts with G9a (EHMT2) (unpublished data). In the future, I will determine whether G9a and H3K9me2 histone signature are present at sites of TRIM24 enrichment at the *CDH1* promoter to uncover an epigenetic mechanism by which TRIM24 regulates E-cadherin expression. ChIP-PCR will be used to assess corecruitment of G9a (Transcription factor Y, Figure 32) and TRIM24 near *CDH1* promoter. In addition, effects of G9a knockdown of TRIM24 recruitment near *CDH1* promoter and vice-versa in MDA-MB-436 cells will be determined.

Future studies will examine the role of TRIM24 in invasion/metastasis by *in vivo* modeling. To determine the role of TRIM24 in regulation of metastasis, RFP/Luciferase labeled MDA-MB-231 cells will be generated with/without TRIM24 knockdown (shControl versus shTRIM24). Approximately, 0.5×10^6 cells will be injected via the tail vein of SCID mice and the presence of lung metastasis will be monitored every week using bioluminescent imaging. After an appropriate time, tumors will be harvested and tumor tissue and number of lung metastatic nodules will be determined. Metastatic tumors will be characterized by H&E staining and IHC for RFP and for expression of various metastatic markers. I expect that mice harboring TRIM24-depleted MDA-MB-231 cells (shTRIM24) will show reduced tumor invasion/metastasis as compared to control cells (shControl).

5.4 TRIM24-domain functional characterization for regulation of EMT:

Previous work from our laboratory has showed that histone reader TRIM24 not only negatively regulates p53 as an E3-ubiquitin ligase but also interacts with and recruits transcription factors, such as estrogen receptor, to chromatin via a tandem

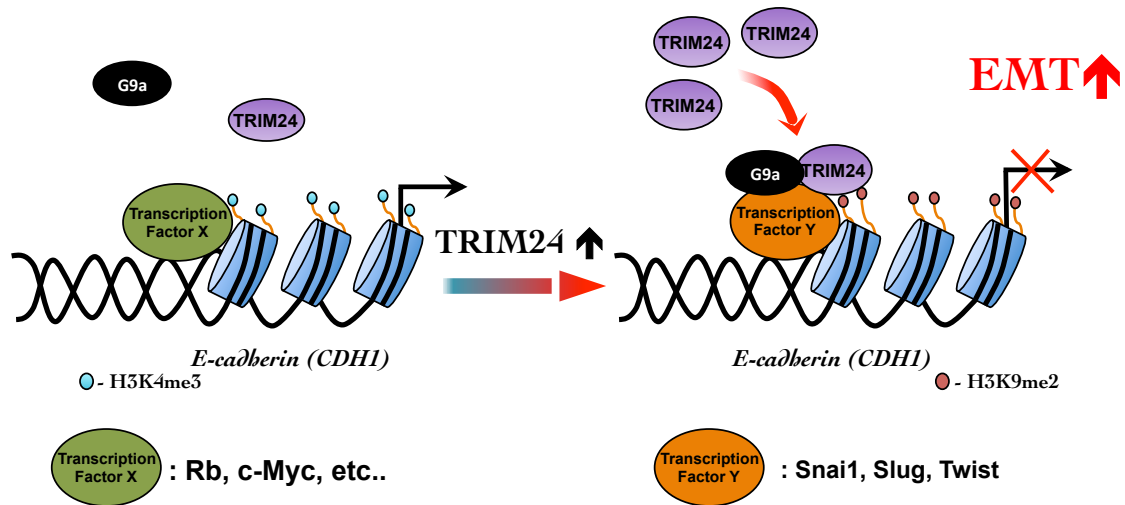


Figure 32: Proposed model for TRIM24 as a novel regulator of EMT in breast cancer

PHD-Bromo domains that binds unmethylated lysine 4 and acetylated lysine 23 of histone H3 (H3K4me0/H3K23ac). The question remains, which of the aforementioned functions is/are critical in regulation of TRIM24-mediated induction of EMT.

To test whether TRIM24-mediated EMT requires its histone reader, p53 regulation, ER-interaction or any subset of these functions, MCF10A-stable cell lines that over express TRIM24 with site-specific mutations in PHD and/or Bromo, p53-regulating RING domains, or LXXLL motifs will be generated. Stable cells will be characterized for various hallmarks of EMT such as change in morphology, gene expression changes and migration properties. Additionally, as part of a drug-discovery program we also have a TRIM24-bromodomain specific small molecule inhibitor (IACS9571). A titration of the IACS9571 inhibitor, guided by known IC₅₀ level, will be used to assess how TRIM24-overexpressing cell lines respond at the level of EMT markers, proliferation and migration/invasion. Its effects will be compared to the phenotype of the TRIM24-Bromodomain point mutant rescue line. These studies may support use of combinations of epigenetic inhibitors that oppose EMT in breast cancers.

Chapter 6: Conclusions

Chapter 6: Conclusions

Since our laboratory's initial report showing TRIM24 over expression correlates with poor survival of breast cancer patients [55], several studies reported roles of TRIM24 in multiple cancers such as in prostate cancer [47], breast cancer [55, 56], non-small cell lung cancer [57], head and neck carcinoma [58], colorectal cancer [59], gastric cancer [60], glioblastoma [61] and human hepatocellular carcinoma [63]. However, except for the glioblastoma study, none of the other studies uncovered the mechanism of TRIM24-mediated oncogenesis. It was found that TRIM24 is up regulated in early stages of breast cancer progression not only in clinical human breast cancer tissues but also in an isogenic human mammary epithelial cell (iHMEC) model system consisting of cells from various stages of breast cancer. Exogenous over-expression of TRIM24 promoted oncogenic transformation of immortalized human mammary epithelial cells (TRIM24-iHMECs) and efficient growth of high-grade tumors in nude mice upon performing xenograft studies.

Gene expression analysis of TRIM24-iHMECs uncovered that aerobic glycolysis and tricarboxylic acid cycle was activated in addition increased glucose uptake was confirmed. In TRIM24-iHMECs, increased c-Myc expression and a decrease in p53 protein levels were observed. c-Myc and p53 play key roles in regulating cellular metabolism, hence, we proposed that these could be the possible mediators of TRIM24-induced metabolic reprogramming. Additionally, TRIM24 is recruited to the promoters of several metabolic genes such as *GLUT1*, *IDH1* and *IDH2*. Further, upon treatment of MCF7 breast cancer cells with a TRIM24-bromodomain inhibitor I found that several glucose metabolism genes were down regulated. Hence, it can be concluded that TRIM24 induces oncogenic transformation coupled with metabolic

reprogramming in breast cancer and TRIM24-bromodomain function may be critical for malignant transformation.

As mentioned before, TRIM24 is reported to be over-expressed in several cancers; however, a detailed analysis of how TRIM24 promotes tumorigenesis with progression to metastasis has not been elucidated. Our preliminary analysis shows that TRIM24 is a driver of epithelial to mesenchymal transition (EMT) in breast cancer. It was found that upon TRIM24 over-expression in MCF10A cells, there is mesenchymal-like change in the cellular morphology. As observed in a classical EMT program, TRIM24-MCF10A cells exhibit marked loss of E-cadherin and acquisition of various mesenchymal markers such as N-cadherin, fibronectin and vimentin. Further, TRIM24-knockdown in MDA-MB-231 and MDA-MB-468, there is reversal of EMT gene signature. HTG Oncology gene expression analysis of TRIM24-OE MCF10A cells further confirmed the EMT was among the top pathways that were enriched. TRIM24 may be involved in transcriptional regulation of EMT genes as confirmed by TRIM24-ChIP analysis and the presence of H3K23ac signature at several EMT genes in MDA-MB-468 cells. Our hypothesis is that aberrant expression of TRIM24 impacts a network of transcription regulatory factors and signaling pathways to induce EMT and metastasis in breast cancer.

Approximately 90% of breast cancer-related deaths are caused by metastasis not by primary tumor [13]. Hence, for effective reduction of cancer mortality, it is very critical to intervene in the critical transition of non-invasive ductal carcinoma to extremely lethal invasive breast cancer. The proposed research holds great promise to identify a unique role of TRIM24 in metastasis and mechanisms involving EMT. These studies will provide a better understanding of molecular mechanisms that drive EMT

and will pave the way for developing therapeutic opportunities for metastatic tumors by targeting the epigenetic regulator TRIM24.

List of Appendices

Appendix 1: List of primers for cloning and qPCR

Cloning Primers			
TRIM24-F1		ATTTTACACGTGATGGAGGTGGCGGTGGAGAA	
TRIM24-R1		CGGGGCTCTAGATTATTTAAGCAACTGGCGTT	
qRT-PCR primers			
Gene	Species	Forward primer	Reverse primer
TRIM24	Human	GCGCCTACT TTTATTTCTTTACTG	TTTGTCAAGAAAGGGTGTAA CG
β-ACTIN	Human	GGACTTCGAGCAAGAGA TGG	AGCACTGTGTTGGCGTACAG
TP53	Human	AAAGAAGAAACCACTGGATG	ATTCAGCTCTCGGAACAT
GLUT1	Human	CCA GCT GCC ATT GCC GTT	GAC GTA GGG ACC ACA CAG TTGC
GLUT4	Human	GTCGGGCTTCCAACAGATAG	ACCCCAATGTTGTACCCAAA
HK2	Human	ATG AGG GGC GGA TGT GTA TCA	GGT TCA GTG AGC CCA TGT CAA
ENO1	Human	AAA GCT GGT GCC GTT GAG AAG	AGC ATG AGA ACC GCC ATT GAT
ALDO-A	Human	GGG TGT CAT CCT CTT CCA TGA	GAC CAC GCC CTT GTC TAC CTT
PGM1	Human	ACC CAC TCC CTT CAT ACA ATC	CTC CTC ACT GGT CAT GGC TTA
PDK1	Human	CCG CTC TCC ATG AAG CAG TT	TTG CCG CAG AAA CAT AAA TGA G
GPI	Human	GGT ACA CAG GCA AGA CCA TC	GTT TTG GCA ATG TGA GTT CC
PKM1	Human	CGAGCCTCAAGTCACTCCAC	GTGAGCAGACCTGCCAGACT
PKM2	Human	ATTATTTGAGGAACTCCGCC GCCT	ATTCCGGGTCACAGCAATGA TGG
LDHC	Human	CCTCTTGGGCTATTGGACTG	GCCTCCTCCTCAGAATTCAA
ACO1	Human	TTGGAGGATTCAAGATATGG	ACTCATCACAATTCCGAAT
DLAT	Human	GTTCTACAGGTGTCTTC	AAGGTAATAATGAGGTAT GGT
IDH1	Human	GAGATAACCTACACACCA AGT	AACACCACCACCTTCTTC
IDH2	Human	GATGGATGGTGATGAGAT	AAATACTTTAGCTGGATGTC

MDH2	Human	TGTGATGTGGTAGTTATTCC	ATTGGTGTTGAACAGGTC
OGDH	Human	AATGTCATCAGGAAGGAG	GGTGGTACTTCACATCTC
PDHA1	Human	ATGTATGCCAAGAACTTC	ATTATACTTACAGGCTAGAG
PDHB	Human	AGGAGTTGAATGTGAGGTGA TAA	ATGACACTGGCTTCTATGGT
SDHA	Human	ATGCCATCCACTACATGA	ATAAATCTTCCCATCTTCA GTT
SDHB	Human	GACACCAACCTCAATAAG	GATTCATCCTTCTTCTTCAA
SDHD	Human	ACCGACCTATCCCAGAATG	CAGCCTTGGAGCCAGAAT
FH	Human	TTAAGATTGGAGGTGTGA	CATAATCCTGGTTTACTTCA
TWIST1	Human	GCCGGAGACCTAGATGTCAT T	CCCACGCCCTGTTTCTTTGA
TGFB	Human	CTTTGGATGCGGCCTATTGC	TAAGCTCAGGACCCTGCTGT
CDH1	Human	TGCCCAGAAAATGAAAAAGG	GTGTATGTGGCAATGCGTTC
CDH2	Human	ACAATCCTCCAGAGTTAC	CTTATCGGTCAGAGTTAGA
VIM	Human	AAGTGGAATGCCCTTAAAG	ATAGTGTCTTGGTAGTTAGC
SNAI2	Human	TCCTGGTCAAAGAAGCATT	ATCTCTGGTTGTGGTATGA

Appendix 2: List of antibodies used for Immunoblotting

Antibody details (Catalog)	Species	Company	Application
TRIM24 (NB100-2597)	Rabbit	Novus Biologicals	Western
TRIM24 (14208-1-AP)	Rabbit	Protein tech	IHC
p53(DO1)-HRP (sc-126)	Mouse	Santa Cruz	Western
b-Actin- HRP (5125)	Rabbit	Cell Signaling	Western
Hexokinase II (2687)	Rabbit	Cell Signaling	Western
Aldolase (3188)	Rabbit	Cell Signaling	Western
Enolase -1 (3810)	Rabbit	Cell Signaling	Western
PDHK1 (3820)	Rabbit	Cell Signaling	Western
LDHA/ LDHC (3538)	Rabbit	Cell Signaling	Western
PGM1 (ab55616)	Mouse	Abcam	Western

Appendix 3: All red score data for TRIM24 staining in Human breast tissue array

Status	Levels	TRIM24 (Number of patients)			Total	Chi-square	P-value
		Low	Medium	High			
ER status	Low	15	22	6	43		
	High	3	12	14	29	11.87	0.0026
	Total	18	34	20	72		
		TRIM24			Total		
		Low	Medium	High			
PR status	Low	14	24	10	48		
	High	4	10	10	24	3.74	0.1541
	Total	18	34	20	72		
		TRIM24			Total		
		Low	Medium	High			
HER2 status	Low	14	8	0	22		
	High	4	26	20	50	28.51	<.0001
	Total	18	34	20	72		
		TRIM24			Total		
		Low	Medium	High			
P53	Low	12	16	12	40		
	High	6	18	8	32	2.05	0.3588
	Total	18	34	20	72		
		TRIM24			Total		
		Low	Medium	High			
Ki67	Low	8	7	7	22		
	High	10	27	13	50	3.41	0.1818
	Total	18	34	20	72		
Key to All Red score							
TRIM24		ER/PR			HER2		
Low (0-2)		Low (0-3)			Low (0-1)		
Medium (3-5)		High (3-8)			High (2-3)		
High (6-8)							

Appendix 4: List of deregulated genes in TRIM24-iHMECs by nanostring analysis

Upregulated genes								
	Control			TRIM24				
Gene	A	B	C	A	B	C	t-test	Fold change
TRIM39	0.00	167.49	132.51	605.68	572.34	591.15	0.00	5.9
PDE4DIP	0.00	219.12	80.88	833.22	376.66	408.42	0.05	5.4
GLS2	236.96	35.30	27.74	518.79	374.78	469.60	0.01	4.5
PRKCI	91.91	92.75	115.34	397.29	440.82	450.99	0.00	4.3
PRKCH	75.47	97.49	127.04	369.65	389.17	473.85	0.00	4.1
CAMKK2	117.28	87.50	95.22	462.87	391.28	374.84	0.00	4.1
SgK494	0.00	202.37	97.63	525.91	415.23	285.19	0.03	4.1
TESK2	0.00	84.47	215.53	370.71	330.23	496.93	0.02	4.0
MYC	84.91	106.26	108.84	336.89	370.51	392.51	0.00	3.7
MAPKAPK5	70.14	94.23	135.63	331.82	317.40	397.15	0.00	3.5
STRADA	21.47	168.55	109.99	285.93	384.41	290.46	0.01	3.2
ME3	17.33	172.80	109.87	310.73	288.43	353.48	0.01	3.2
FGFR2	100.45	114.59	84.96	273.19	277.81	358.25	0.00	3.0
TTL	84.61	111.39	104.00	293.80	305.09	309.23	0.00	3.0
DDX5	90.50	124.69	84.81	357.23	258.22	290.89	0.00	3.0
TRIM26	44.07	142.46	113.47	260.31	272.90	327.49	0.01	2.9
CEBPA	90.19	111.15	98.66	266.75	298.82	274.99	0.00	2.8
TRIM37	85.26	113.30	101.44	292.72	278.46	264.67	0.00	2.8
TRIM39	84.33	116.55	99.12	296.83	235.94	284.93	0.00	2.7
MTOR	57.47	135.06	107.47	218.24	306.41	280.85	0.01	2.7
PIK3CA	63.52	93.07	143.41	254.09	241.62	309.69	0.01	2.7
BIRC3	98.26	102.09	99.65	288.82	256.61	243.06	0.00	2.6
PTPN11	80.24	120.97	98.80	238.57	259.67	286.49	0.00	2.6
PFKFB2	57.89	122.72	119.40	216.86	265.92	299.32	0.01	2.6
EPHA1	86.47	88.62	124.92	274.97	203.83	302.21	0.01	2.6
TTK	103.65	88.67	107.68	238.36	225.32	295.02	0.00	2.5
YES1	80.08	109.47	110.45	240.24	247.23	272.17	0.00	2.5
NEK6	99.98	87.87	112.15	256.38	207.41	255.11	0.00	2.4
NADK	80.59	132.86	86.55	193.31	240.18	272.71	0.01	2.4
STK38L	108.16	97.62	94.22	229.55	233.62	228.09	0.00	2.3
CDKN2A	189.02	74.63	36.35	217.37	229.33	240.38	0.05	2.3
ERN1	78.27	133.34	88.39	265.59	213.98	203.27	0.01	2.3
SDHD	96.31	83.77	119.92	210.35	205.87	264.55	0.00	2.3
TNK2	88.53	97.96	113.52	201.61	234.60	243.05	0.00	2.3
LGR5	162.81	64.29	72.90	176.33	251.31	206.28	0.04	2.1
COX6C	96.44	102.71	100.85	207.39	198.92	224.80	0.00	2.1

TSC1	80.94	107.44	111.62	220.08	174.18	231.16	0.01	2.1
STK38	71.10	105.05	123.85	198.54	203.43	215.53	0.00	2.1
TRIM59	73.62	115.41	110.97	207.78	203.48	203.30	0.00	2.1
PIP4K2A	80.18	116.42	103.40	211.78	196.78	182.02	0.00	2.0
BRCA1	61.92	123.09	115.00	190.10	195.23	195.77	0.01	1.9
SIX4	117.49	84.35	98.17	166.19	214.52	202.52	0.01	1.9
SIK3	97.85	116.82	85.33	189.45	199.08	193.78	0.00	1.9
MASTL	63.29	121.37	115.34	197.64	184.52	198.17	0.01	1.9
ADRBK1	98.21	95.03	106.76	176.92	204.95	197.93	0.00	1.9
MAP3K5	101.84	103.09	95.07	208.56	139.72	227.76	0.03	1.9
SUFU	112.01	99.86	88.12	168.24	211.29	196.43	0.00	1.9
RPIA	104.92	102.65	92.43	152.23	192.15	221.93	0.01	1.9
EZH2	78.28	98.23	123.49	211.74	164.89	187.48	0.01	1.9
BRD3	66.22	104.59	129.19	194.66	160.63	189.77	0.02	1.8
PFKFB3	100.47	114.40	85.13	186.71	186.88	170.40	0.00	1.8
TRIM8	110.66	97.48	91.86	160.42	184.29	189.82	0.00	1.8
GNAQ	114.66	96.08	89.26	192.33	166.22	174.53	0.00	1.8
DLAT	88.02	116.26	95.71	157.99	208.15	163.98	0.01	1.8
SUZ12	96.47	92.31	111.22	162.27	156.41	208.20	0.01	1.8
RPS6KB1	109.51	99.07	91.42	167.09	187.02	165.46	0.00	1.7
TUBB	97.55	95.46	106.99	175.70	162.12	179.90	0.00	1.7
CSK	74.68	117.18	108.14	149.28	207.10	160.90	0.03	1.7
AURKB	86.51	111.06	102.43	161.71	188.84	164.13	0.00	1.7
HPRT1	103.22	87.37	109.41	171.22	164.29	179.41	0.00	1.7
ACLY	88.10	95.11	116.79	171.95	156.01	185.87	0.00	1.7
VEGFA	96.71	95.72	107.56	161.44	162.53	182.52	0.00	1.7
SDHB	88.47	96.21	115.32	159.71	157.81	183.03	0.00	1.7
SNX16	110.81	94.84	94.35	175.13	151.39	174.28	0.00	1.7
PIK3R4	76.17	113.25	110.58	159.56	168.70	163.27	0.01	1.6
EXOSC10	103.90	92.24	103.86	182.68	151.99	153.98	0.00	1.6
DDR1	117.47	98.22	84.31	163.14	146.66	177.28	0.01	1.6
TIAM1	139.77	86.31	73.93	163.60	150.38	173.17	0.04	1.6
CSNK1D	75.44	119.51	105.05	149.96	171.24	160.75	0.01	1.6
DNMT3A	119.72	82.23	98.05	151.03	154.57	177.99	0.01	1.6
NRAS	83.60	121.47	94.93	163.56	167.72	152.35	0.01	1.6
MELK	87.94	104.29	107.77	137.44	163.73	174.99	0.01	1.6
GSG2	88.51	106.43	105.06	160.65	153.49	164.00	0.00	1.6
KRAS	72.02	113.59	114.40	163.79	159.94	150.47	0.02	1.6
PAK2	87.05	101.44	111.51	143.84	161.38	161.67	0.00	1.6
PANK4	85.52	100.27	114.21	162.11	159.11	147.79	0.00	1.6
GMPS	88.50	103.49	108.01	157.02	147.45	161.72	0.00	1.6
CHEK1	95.68	101.35	102.97	166.56	147.35	150.39	0.00	1.6
PDP1	104.21	84.35	111.44	147.34	148.89	165.52	0.01	1.5

SGK1	111.61	97.68	90.71	170.02	146.22	142.16	0.01	1.5
MAPK8	106.53	101.84	91.63	144.52	149.29	163.98	0.00	1.5
RIOK1	100.31	106.32	93.37	142.94	154.84	156.08	0.00	1.5
BUB1B	103.13	90.63	106.24	167.40	138.12	140.39	0.01	1.5
HIPK1	106.47	97.06	96.47	151.93	148.97	146.76	0.00	1.5
DYRK1A	95.03	110.40	94.57	126.29	173.39	144.49	0.03	1.5
SDHA	115.02	87.55	97.43	159.51	137.58	146.76	0.01	1.5
HK1	69.60	108.27	122.13	139.79	145.81	156.05	0.05	1.5
LGR4	92.55	101.35	106.10	174.12	134.23	128.84	0.04	1.5
PRKAR2A	108.79	96.03	95.18	138.54	153.30	143.89	0.00	1.5
SRPK1	77.14	124.73	98.13	134.12	147.25	151.52	0.04	1.4
MARK2	112.57	85.94	101.49	128.35	137.62	167.43	0.03	1.4
BRD8	95.25	113.48	91.27	129.06	142.60	159.09	0.02	1.4
INPP4A	123.68	92.94	83.37	162.34	131.32	135.13	0.05	1.4
GPR172A	110.01	92.75	97.24	150.82	140.09	134.87	0.00	1.4
PAFAH1B2	97.27	92.88	109.85	131.99	140.01	151.57	0.01	1.4
NLK	114.80	83.74	101.46	146.56	140.69	133.29	0.01	1.4
MAP3K7	111.10	103.76	85.14	139.29	139.75	138.50	0.01	1.4
CS	119.50	92.02	88.48	157.98	128.22	130.72	0.05	1.4
P2RY2	90.52	92.84	116.64	137.82	134.08	145.78	0.01	1.4
KIAA1524	106.11	94.53	99.36	146.37	145.96	124.19	0.01	1.4
AK3	88.50	112.84	98.65	133.79	134.47	146.39	0.01	1.4
BRD2	92.84	105.03	102.13	138.39	148.03	127.62	0.01	1.4
ABL1	100.00	104.92	95.08	116.72	158.40	134.39	0.04	1.4
SH3BP5	85.21	107.66	107.13	137.75	138.42	132.58	0.01	1.4
BRD7	88.56	108.28	103.16	116.82	138.78	148.61	0.04	1.4
C17orf75	102.12	94.30	103.58	120.93	145.56	131.93	0.01	1.3
PI4K2B	81.40	114.94	103.65	137.73	127.26	131.01	0.04	1.3
MDH2	106.64	99.58	93.78	132.61	136.44	125.99	0.00	1.3
SMARCB1	89.38	105.81	104.81	115.12	141.11	135.71	0.03	1.3
BRD9	109.05	100.42	90.53	144.25	128.72	120.62	0.02	1.3
MAPK14	89.74	110.37	99.89	133.47	127.90	127.64	0.01	1.3
VHL	111.08	98.47	90.45	136.91	135.91	112.84	0.04	1.3
CSNK2B	87.32	101.10	111.58	120.88	135.08	131.34	0.02	1.3
MAPK9	93.30	103.48	103.23	116.71	134.68	136.63	0.02	1.3
FH	88.01	103.30	108.69	116.95	122.93	134.12	0.04	1.3
RIPK2	107.86	93.78	98.36	128.03	118.96	127.05	0.01	1.3
RB1	100.30	99.01	100.69	127.90	127.04	119.56	0.00	1.2
MAPK6	89.31	101.11	109.58	112.56	134.50	123.49	0.05	1.2
MAP2K4	106.21	87.45	106.34	113.59	124.80	128.78	0.04	1.2
TGFBR1	106.30	96.83	96.87	123.82	125.83	116.76	0.01	1.2
MET	100.36	97.04	102.60	117.81	120.36	123.81	0.00	1.2
ACO1	91.31	106.02	102.67	116.76	121.04	122.48	0.01	1.2

GPI	98.78	108.12	93.10	125.55	118.94	111.49	0.04	1.2
RAF1	100.61	102.72	96.68	114.38	109.57	131.47	0.05	1.2
CDK4	101.71	103.23	95.06	113.55	113.40	125.26	0.02	1.2
CDK1	94.53	104.79	100.68	113.34	112.18	125.61	0.03	1.2
PTEN	104.69	102.38	92.93	118.46	113.63	114.01	0.02	1.2
Down-Regulated Genes								
CANT1	102.26	102.13	95.61	74.26	77.76	72.82	0.00	0.7
RPS6KA4	89.61	112.76	97.63	67.81	80.92	74.18	0.03	0.7
ATIC	92.36	103.75	103.90	69.51	72.44	77.62	0.00	0.7
RPS6KC1	97.65	110.53	91.82	63.45	84.16	71.48	0.03	0.7
PRKAR1A	99.04	100.29	100.67	68.68	72.59	76.65	0.00	0.7
RIPK1	100.25	95.01	104.74	63.70	69.64	83.27	0.01	0.7
BRCA2	94.16	106.10	99.74	68.15	71.34	77.11	0.00	0.7
PDXK	95.39	105.72	98.89	73.37	69.33	73.59	0.00	0.7
LATS2	109.68	106.83	83.49	76.02	66.31	71.03	0.03	0.7
AURKA	94.64	113.24	92.13	71.74	65.29	75.09	0.02	0.7
SPTAN1	94.90	102.35	102.74	68.42	69.69	72.68	0.00	0.7
CAMK2D	85.56	115.08	99.36	69.50	65.52	74.53	0.03	0.7
BMPR1A	109.26	103.28	87.46	65.26	71.55	69.21	0.01	0.7
TKT	94.42	106.27	99.31	65.99	67.99	70.86	0.00	0.7
SGK196	86.59	112.34	101.08	72.34	66.83	65.30	0.01	0.7
LIMK2	101.43	93.63	104.94	69.47	68.76	64.83	0.00	0.7
FBXW7	106.32	104.96	88.72	72.23	65.27	65.31	0.01	0.7
SRPK2	94.96	101.78	103.26	71.26	63.48	65.17	0.00	0.7
SMAD4	106.16	95.81	98.03	58.31	63.28	74.85	0.00	0.7
PIP5K1A	97.04	106.67	96.29	65.90	68.32	62.03	0.00	0.7
CARS	109.98	104.43	85.59	63.52	60.63	65.80	0.01	0.6
BRD1	120.83	90.76	88.41	65.75	63.12	60.99	0.03	0.6
PDK1	103.94	109.44	86.61	73.79	60.17	53.65	0.01	0.6
H6PD	124.34	78.77	96.89	54.75	70.57	60.73	0.05	0.6
RUNX1	110.51	88.95	100.54	60.45	62.33	62.23	0.00	0.6
PIKFYVE	105.65	89.69	104.66	65.22	53.00	66.37	0.00	0.6
PRKD2	114.02	96.97	89.01	59.53	57.54	66.06	0.01	0.6
NF2	104.52	97.14	98.34	60.72	64.47	57.76	0.00	0.6
JUN	110.79	105.36	83.84	60.50	58.71	62.61	0.01	0.6
BMPR2	102.34	97.17	100.49	61.37	54.70	63.72	0.00	0.6
CCND1	107.49	97.78	94.73	58.79	58.02	61.48	0.00	0.6
CDH1	93.28	106.31	100.41	58.32	61.00	58.15	0.00	0.6
TRIB1	114.51	88.92	96.56	58.70	54.87	63.68	0.01	0.6
ABL2	110.03	101.83	88.14	54.74	62.36	58.45	0.00	0.6
NPM1	96.75	104.96	98.29	54.00	54.00	56.62	0.00	0.5
ETNK2	95.16	87.67	117.17	44.66	52.02	66.98	0.01	0.5
JAK1	70.16	104.17	125.66	55.93	54.15	51.56	0.05	0.5

PLXNB2	107.81	99.81	92.38	53.14	49.49	57.30	0.00	0.5
NEK7	106.05	94.44	99.51	50.83	55.51	53.60	0.00	0.5
HERC4	101.72	105.21	93.07	53.40	50.35	53.23	0.00	0.5
RIOK3	114.66	102.70	82.64	52.09	41.01	62.60	0.01	0.5
ATF2	93.99	107.22	98.78	38.01	56.30	58.19	0.00	0.5
APC	93.07	133.46	73.47	54.11	50.09	48.61	0.05	0.5
PIK3R1	71.49	110.08	118.43	55.74	41.38	52.83	0.03	0.5
TNIK	92.42	118.61	88.97	48.05	40.12	59.52	0.01	0.5
CXCR7	96.52	94.33	109.15	52.16	52.47	38.81	0.00	0.5
GNAS	99.34	98.90	101.76	48.50	47.80	48.36	0.00	0.5
PKN1	104.60	88.72	106.68	45.31	46.34	47.45	0.00	0.5
LPAR1	93.72	116.63	89.65	36.29	49.87	48.58	0.00	0.4
PIK3C2B	101.12	103.81	95.08	56.94	39.02	38.43	0.00	0.4
IRAK2	85.32	108.56	106.12	42.10	56.71	35.03	0.00	0.4
TGFB2	87.37	113.03	99.60	37.35	40.70	43.57	0.00	0.4
CALM3	88.83	102.81	108.35	36.49	43.15	36.87	0.00	0.4
HRAS	69.46	116.39	114.15	38.16	35.63	41.85	0.02	0.4
IDH1	106.85	109.69	83.46	37.03	29.78	47.63	0.00	0.4
MAGI2	133.29	92.11	74.60	43.31	45.01	21.11	0.03	0.4
FLT3	122.76	113.11	64.13	26.59	47.37	34.56	0.03	0.4
PRKCQ	101.45	100.14	98.41	32.96	35.88	39.27	0.00	0.4
ILK	96.77	106.34	96.88	29.07	34.35	42.80	0.00	0.4
INPP4B	92.55	107.86	99.59	36.86	29.67	37.83	0.00	0.3
PLK2	97.54	92.11	110.35	30.90	32.64	37.50	0.00	0.3
AURKC	122.94	116.51	60.55	21.30	37.95	41.54	0.03	0.3
FLNB	105.10	95.84	99.05	32.87	31.05	28.04	0.00	0.3
EPHB6	121.89	96.25	81.86	26.40	39.20	25.74	0.01	0.3
NTRK1	119.13	109.76	71.12	25.80	22.99	41.93	0.01	0.3
TGFB3	149.12	84.11	66.77	36.91	32.88	17.99	0.05	0.3
TRIM5	119.21	101.99	78.80	22.75	26.84	35.36	0.00	0.3
HKDC1	98.24	135.77	65.98	26.60	33.17	20.75	0.02	0.3
TBX22	130.75	120.46	48.79	28.32	33.64	18.41	0.05	0.3
HOXB7	118.67	96.82	84.51	24.45	31.31	19.46	0.00	0.3
MST1R	104.36	103.37	92.27	22.22	24.91	26.15	0.00	0.2
MYLK4	69.48	137.18	93.33	30.10	26.81	14.67	0.02	0.2
MRGPRX3	118.30	93.42	88.28	25.62	22.83	16.65	0.00	0.2
ALK	104.07	102.73	93.20	22.54	20.08	21.98	0.00	0.2
INSR	99.84	92.06	108.09	13.64	17.95	29.07	0.00	0.2
LRP1B	119.97	126.32	53.71	0.00	15.43	33.78	0.03	0.2
EPHA4	128.80	106.80	64.40	20.46	14.91	12.69	0.01	0.2
ERBB4	98.24	135.77	65.98	10.64	18.96	10.37	0.01	0.1
ROBO1	103.32	95.99	100.69	14.48	12.90	11.55	0.00	0.1
STK32A	84.10	121.76	94.14	10.12	16.23	7.89	0.00	0.1

MAP2K6	98.31	109.99	91.70	10.65	15.81	6.92	0.00	0.1
RPS6KA6	143.73	83.17	73.10	14.01	10.05	7.98	0.02	0.1
ANGPTL4	120.81	95.40	83.79	9.35	12.90	7.06	0.00	0.1
FYN	82.16	118.31	99.54	8.37	8.39	9.18	0.00	0.1
PRKD1	74.75	114.38	110.87	6.07	9.01	9.87	0.00	0.1
SMO	120.56	92.82	86.62	0.00	15.95	0.00	0.00	0.1
AKT3	88.61	105.97	105.42	3.84	4.88	5.35	0.00	0.0
TP53	110.82	99.07	90.11	4.99	3.59	2.62	0.00	0.0
CDK14	118.73	90.53	90.74	3.55	5.53	1.73	0.00	0.0
ROS1	92.87	100.01	107.11	2.13	1.90	3.86	0.00	0.0
HCK	100.78	126.35	72.87	0.00	0.00	0.00	0.00	0.0

Appendix 5: GO pathway analysis of deregulated genes in TRIM24-iHMECs

Upregulated Genes			
Term	Count	%	P-Value
GO:0006468~protein amino acid phosphorylation	52	41.3	4.83E-37
GO:0016310~phosphorylation	55	43.7	1.13E-36
GO:0006793~phosphorus metabolic process	57	45.2	1.61E-34
GO:0006796~phosphate metabolic process	57	45.2	1.61E-34
GO:0007242~intracellular signaling cascade	38	30.2	2.28E-12
GO:0043549~regulation of kinase activity	19	15.1	7.45E-10
GO:0042325~regulation of phosphorylation	21	16.7	1.28E-09
GO:0051338~regulation of transferase activity	19	15.1	1.44E-09
GO:0019220~regulation of phosphate metabolic process	21	16.7	2.55E-09
GO:0051174~regulation of phosphorus metabolic process	21	16.7	2.55E-09
GO:0045859~regulation of protein kinase activity	18	14.3	3.28E-09
GO:0006084~acetyl-CoA metabolic process	8	6.3	4.92E-09
GO:0007243~protein kinase cascade	18	14.3	9.35E-09
GO:0009060~aerobic respiration	8	6.3	1.23E-08
GO:0046356~acetyl-CoA catabolic process	7	5.6	2.53E-08
GO:0006099~tricarboxylic acid cycle	7	5.6	2.53E-08
GO:0006732~coenzyme metabolic process	12	9.5	5.32E-08
GO:0009109~coenzyme catabolic process	7	5.6	5.66E-08
GO:0051187~cofactor catabolic process	7	5.6	1.75E-07
GO:0042127~regulation of cell proliferation	23	18.3	4.02E-07
GO:0046777~protein amino acid autophosphorylation	9	7.1	4.86E-07
GO:0042981~regulation of apoptosis	23	18.3	5.78E-07
GO:0051186~cofactor metabolic process	12	9.5	6.22E-07
GO:0043067~regulation of programmed cell death	23	18.3	6.83E-07
GO:0010941~regulation of cell death	23	18.3	7.27E-07
GO:0000075~cell cycle checkpoint	9	7.1	8.24E-07
GO:0044093~positive regulation of molecular function	19	15.1	1.41E-06
GO:0043085~positive regulation of catalytic activity	17	13.5	5.66E-06
GO:0043066~negative regulation of apoptosis	14	11.1	7.11E-06
GO:0043648~dicarboxylic acid metabolic process	6	4.8	8.17E-06
GO:0043069~negative regulation of programmed cell death	14	11.1	8.27E-06
GO:0060548~negative regulation of cell death	14	11.1	8.52E-06
GO:0006091~generation of precursor metabolites and energy	13	10.3	1.06E-05
GO:0018107~peptidyl-threonine phosphorylation	5	4.0	1.25E-05
GO:0006917~induction of apoptosis	13	10.3	1.33E-05
GO:0012502~induction of programmed cell death	13	10.3	1.37E-05
GO:0045333~cellular respiration	8	6.3	1.54E-05

GO:0051726~regulation of cell cycle	13	10.3	1.86E-05
GO:0018210~peptidyl-threonine modification	5	4.0	2.39E-05
GO:0010033~response to organic substance	19	15.1	2.46E-05
GO:0019318~hexose metabolic process	10	7.9	3.02E-05
GO:0006915~apoptosis	17	13.5	3.47E-05
GO:0006006~glucose metabolic process	9	7.1	3.92E-05
GO:0012501~programmed cell death	17	13.5	4.14E-05
GO:0042770~DNA damage response, signal transduction	7	5.6	5.14E-05
GO:0043065~positive regulation of apoptosis	14	11.1	5.49E-05
GO:0043068~positive regulation of programmed cell death	14	11.1	5.90E-05
GO:0010942~positive regulation of cell death	14	11.1	6.18E-05
GO:0008219~cell death	18	14.3	8.34E-05
GO:0016265~death	18	14.3	9.08E-05
GO:0005996~monosaccharide metabolic process	10	7.9	9.33E-05
GO:0006007~glucose catabolic process	6	4.8	1.15E-04
GO:0007167~enzyme linked receptor protein signaling pathway	12	9.5	1.24E-04
GO:0033674~positive regulation of kinase activity	10	7.9	1.26E-04
GO:0000165~MAPKKK cascade	9	7.1	1.43E-04
Down-regulated Genes			
Term	Count	%	PValue
GO:0016310~phosphorylation	50	54.9	5.39E-37
GO:0006468~protein amino acid phosphorylation	47	51.6	1.07E-36
GO:0006793~phosphorus metabolic process	50	54.9	5.99E-33
GO:0006796~phosphate metabolic process	50	54.9	5.99E-33
GO:0007242~intracellular signaling cascade	31	34.1	1.56E-10
GO:0007167~enzyme linked receptor protein signaling pathway	17	18.7	6.50E-10
GO:0042325~regulation of phosphorylation	19	20.9	1.07E-09
GO:0051174~regulation of phosphorus metabolic process	19	20.9	2.01E-09
GO:0019220~regulation of phosphate metabolic process	19	20.9	2.01E-09
GO:0007243~protein kinase cascade	17	18.7	2.03E-09
GO:0042127~regulation of cell proliferation	22	24.2	2.66E-08
GO:0051094~positive regulation of developmental process	14	15.4	3.07E-08
GO:0044093~positive regulation of molecular function	18	19.8	2.19E-07
GO:0045597~positive regulation of cell differentiation	12	13.2	2.95E-07
GO:0045859~regulation of protein kinase activity	14	15.4	3.77E-07
GO:0043549~regulation of kinase activity	14	15.4	5.57E-07
GO:0010033~response to organic substance	19	20.9	8.33E-07
GO:0051338~regulation of transferase activity	14	15.4	8.86E-07
GO:0001932~regulation of protein amino acid phosphorylation	10	11.0	1.96E-06

GO:0007169~transmembrane receptor protein tyrosine kinase signaling pathway	11	12.1	2.06E-06
GO:0051130~positive regulation of cellular component organization	10	11.0	2.85E-06
GO:0008284~positive regulation of cell proliferation	14	15.4	2.91E-06
GO:0009725~response to hormone stimulus	13	14.3	4.81E-06
GO:0051098~regulation of binding	9	9.9	7.25E-06
GO:0007166~cell surface receptor linked signal transduction	29	31.9	1.27E-05
GO:0009719~response to endogenous stimulus	13	14.3	1.30E-05
GO:0051101~regulation of DNA binding	8	8.8	1.45E-05
GO:0045860~positive regulation of protein kinase activity	10	11.0	1.55E-05
GO:0033674~positive regulation of kinase activity	10	11.0	2.05E-05
GO:0008285~negative regulation of cell proliferation	12	13.2	2.38E-05
GO:0030097~hemopoiesis	10	11.0	2.43E-05
GO:0051347~positive regulation of transferase activity	10	11.0	2.77E-05
GO:0043085~positive regulation of catalytic activity	14	15.4	3.33E-05
GO:0032388~positive regulation of intracellular transport	5	5.5	4.21E-05
GO:0048534~hemopoietic or lymphoid organ development	10	11.0	5.18E-05
GO:0006275~regulation of DNA replication	6	6.6	5.30E-05
GO:0007569~cell aging	5	5.5	6.19E-05
GO:0044092~negative regulation of molecular function	11	12.1	6.64E-05
GO:0043408~regulation of MAPKKK cascade	7	7.7	8.05E-05
GO:0002520~immune system development	10	11.0	8.21E-05
GO:0007568~aging	7	7.7	8.47E-05
GO:0042493~response to drug	9	9.9	8.64E-05
GO:0007369~gastrulation	6	6.6	9.51E-05
GO:0043388~positive regulation of DNA binding	6	6.6	9.51E-05
GO:0032386~regulation of intracellular transport	6	6.6	1.02E-04
GO:0051050~positive regulation of transport	9	9.9	1.08E-04
GO:0030334~regulation of cell migration	8	8.8	1.22E-04
GO:0031399~regulation of protein modification process	10	11.0	1.36E-04
GO:0051259~protein oligomerization	8	8.8	1.47E-04
GO:0051092~positive regulation of NF-kappaB transcription factor activity	5	5.5	1.47E-04
GO:0051099~positive regulation of binding	6	6.6	1.59E-04
GO:0042306~regulation of protein import into nucleus	5	5.5	1.94E-04
GO:0010627~regulation of protein kinase cascade	9	9.9	2.30E-04
GO:0018108~peptidyl-tyrosine phosphorylation	5	5.5	2.31E-04
GO:0031346~positive regulation of cell projection organization	5	5.5	2.52E-04
GO:0040012~regulation of locomotion	8	8.8	2.68E-04
GO:0018212~peptidyl-tyrosine modification	5	5.5	2.73E-04
GO:0051270~regulation of cell motion	8	8.8	2.77E-04

GO:0032880~regulation of protein localization	7	7.7	2.93E-04
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Appendix 6: KEGG pathway analysis of deregulated genes in TRIM24-iHMECs

Up-regulated Genes				
Term	Count	%	P-Value	Benjamini
Citrate cycle (TCA cycle)	9	7.1	2E-08	2E-06
Pathways in cancer	22	17.5	3E-08	2E-06
ErbB signaling pathway	12	9.5	1E-07	4E-06
Chronic myeloid leukemia	11	8.7	3E-07	6E-06
Renal cell carcinoma	10	7.9	2E-06	3E-05
Pancreatic cancer	10	7.9	2E-06	3E-05
Bladder cancer	8	6.3	4E-06	6E-05
Neurotrophin signaling pathway	12	9.5	4E-06	5E-05
Glioma	9	7.1	7E-06	7E-05
Insulin signaling pathway	12	9.5	1E-05	9E-05
Melanoma	9	7.1	2E-05	1E-04
Acute myeloid leukemia	8	6.3	4E-05	3E-04
MAPK signaling pathway	15	11.9	9E-05	6E-04
Non-small cell lung cancer	7	5.6	2E-04	2E-03
Fc epsilon RI signaling pathway	8	6.3	3E-04	2E-03
T cell receptor signaling pathway	9	7.1	3E-04	2E-03
Colorectal cancer	8	6.3	4E-04	2E-03
Prostate cancer	8	6.3	6E-04	3E-03
Epithelial cell signaling in Helicobacter pylori infection	7	5.6	8E-04	4E-03
Cell cycle	9	7.1	9E-04	4E-03
GnRH signaling pathway	8	6.3	1E-03	5E-03
mTOR signaling pathway	6	4.8	2E-03	6E-03
Endometrial cancer	6	4.8	2E-03	6E-03
Progesterone-mediated oocyte maturation	7	5.6	3E-03	1E-02
Gap junction	7	5.6	3E-03	1E-02
NOD-like receptor signaling pathway	6	4.8	3E-03	1E-02
Down-regulated Genes				
Term	Count	%	P-Value	Benjamini
Neurotrophin signaling pathway	12	13.19	1.4E-07	1.2E-05
Endometrial cancer	8	8.79	2.0E-06	8.4E-05
Pathways in cancer	16	17.58	3.8E-06	1.1E-04
Pancreatic cancer	8	8.79	1.8E-05	3.8E-04
Chronic myeloid leukemia	8	8.79	2.4E-05	4.0E-04
Colorectal cancer	8	8.79	5.0E-05	7.0E-04
Glioma	7	7.69	8.4E-05	1.0E-03
Thyroid cancer	5	5.49	3.3E-04	3.5E-03
Apoptosis	7	7.69	5.0E-04	4.7E-03

ErbB signaling pathway	7	7.69	5.0E-04	4.7E-03
Acute myeloid leukemia	6	6.59	5.5E-04	4.6E-03
MAPK signaling pathway	11	12.09	9.8E-04	7.6E-03
Melanoma	6	6.59	1.4E-03	9.8E-03
T cell receptor signaling pathway	7	7.69	1.6E-03	1.0E-02
Phosphatidylinositol signaling system	6	6.59	1.7E-03	1.2E-02
Fc epsilon RI signaling pathway	6	6.59	2.1E-03	1.9E-02
Non-small cell lung cancer	5	5.49	3.6E-03	1.9E-02
Axon guidance	7	7.69	3.9E-03	2.3E-02
Fc gamma R-mediated phagocytosis	6	6.59	5.0E-03	2.5E-02
GnRH signaling pathway	6	6.59	5.7E-03	3.7E-02
Focal adhesion	8	8.79	8.9E-03	5.0E-02
Adherens junction	5	5.49	1.2E-02	4.9E-02
Bladder cancer	4	4.40	1.3E-02	7.3E-02
Prostate cancer	5	5.49	2.0E-02	7.2E-02
Insulin signaling pathway	6	6.59	2.1E-02	8.3E-02
Inositol phosphate metabolism	4	4.40	2.5E-02	9.7E-02
Toll-like receptor signaling pathway	5	5.49	3.1E-02	9.7E-02
Wnt signaling pathway	6	6.59	3.2E-02	1.3E-01
Long-term potentiation	4	4.40	4.6E-02	1.4E-01

Appendix 7: GSEA report for pathways positively correlating with TRIM24 expression in invasive carcinoma patients

Pathways postively enriched with TRIM24				
NAME	FDR q-val	ES	NES	NOM p-val
KEGG_OOCYTE_MEIOSIS	0.06408	0.52151	1.87444	0.00000
REACTOME_DEADENYLATION_DEPENDENT_MRNA_DECAY	0.06434	0.62592	1.87805	0.00382
REACTOME_KINESINS	0.06713	0.83314	1.87817	0.00192
PID_AURORA_B_PATHWAY	0.06795	0.71556	1.85762	0.00763
REACTOME_TRANSPORT_OF_MATURE_MRNA_DERIVED_FROM_AN_INTRONLESS_TRANSCRIPT	0.06845	0.71711	1.88203	0.00389
REACTOME_ANTIGEN_PROCESSING_UBIQUITINATION_PROTEASOME_DEGRADATION	0.07023	0.49987	1.85804	0.00186
REACTOME_GLUCOSE_TRANSPORT	0.07066	0.63171	1.88390	0.00186
REACTOME_TRANSCRIPTION	0.07192	0.55355	1.91430	0.00546
REACTOME_MITOTIC_G2_G2_M_PHASES	0.07214	0.56097	1.84754	0.00546
REACTOME_NEP_NS2_INTERACTS_WITH_THE_CELLULAR_EXPORT_MACHINERY	0.07263	0.75149	1.85856	0.00195
REACTOME_ANTIVIRAL_MECHANISM_BY_IFN_STIMULATED_GENES	0.07322	0.61327	1.88518	0.00000
KEGG_RNA_DEGRADATION	0.07602	0.59834	1.91539	0.00000
REACTOME_REGULATION_OF_GLUCOKINASE_BY_GLUCOKINASE_REGULATORY_PROTEIN	0.07690	0.71516	1.88557	0.00190
REACTOME_TRANSPORT_OF_RIBONUCLEOPROTEINS_INTO_THE_HOST_NUCLEUS	0.07828	0.75012	1.88901	0.00000
REACTOME_LATE_PHASE_OF_HIV_LIFE_CYCLE	0.07982	0.55328	1.83218	0.00570
REACTOME_TRANSPORT_OF_MATURE_TRANSCRIPT_TO_CYTOSOL	0.08048	0.68610	1.89741	0.00388
REACTOME_MEIOSIS	0.08146	0.59509	1.91615	0.00000
BIOCARTA_ATRBRCA_PATHWAY	0.08189	0.76268	1.89002	0.00000
REACTOME_MEIOTIC_RECOMBINATION	0.08225	0.64034	1.82548	0.00188
REACTOME_INTERACTIONS_OF_VPR_WITH_HOST_CELLULAR_PROTEINS	0.08325	0.72806	1.92217	0.00195
REACTOME_MRNA_3_END_PROCESSING	0.08413	0.69132	1.81575	0.00586
REACTOME_RNA_POL_II_TRANSCRIPTION	0.08626	0.55333	1.80294	0.01095
REACTOME_METABOLISM_OF_NON_CODING_RNA	0.08628	0.66598	1.81632	0.00576
REACTOME_TELOMERE_MAINTENANCE	0.08643	0.68094	1.80537	0.01328
REACTOME_HIV_LIFE_CYCLE	0.08865	0.54442	1.80587	0.00949
REACTOME_MEIOTIC_SYNAPSIS	0.08981	0.62712	1.92364	0.00000
PID_PLK1_PATHWAY	0.09299	0.70542	1.93973	0.00366
PID_BARD1PATHWAY	0.09308	0.70553	1.79135	0.00593
REACTOME_MITOTIC_M_M_G1_PHASES	0.09489	0.67837	1.92756	0.00372
REACTOME_CLEAVAGE_OF_GROWING_TRANSCRIPT_IN_THE_TERMINATION_REGION	0.09509	0.67971	1.78624	0.00568
PID_E2F_PATHWAY	0.09586	0.53455	1.78174	0.00727
REACTOME_G2_M_CHECKPOINTS	0.09661	0.74742	1.77472	0.01326
KEGG_CELL_CYCLE	0.09687	0.55862	1.77733	0.01296
REACTOME_LOSS_OF_NLP_FROM_MITOTIC_CENTROSOMES	0.09713	0.54059	1.76872	0.00749
REACTOME_CELL_CYCLE_MITOTIC	0.09823	0.63418	2.01532	0.00366

PID_ATM_PATHWAY	0.09938	0.62878	1.76889	0.00559
REACTOME_G0_AND_EARLY_G1	0.09998	0.74488	1.76329	0.00765
REACTOME_MRNA_PROCESSING	0.10062	0.55231	1.75046	0.01527
PID_P53REGULATIONPATHWAY	0.10128	0.51186	1.75457	0.00384
REACTOME_CELL_CYCLE_CHECKPOINTS	0.10152	0.63477	1.75934	0.01498
REACTOME_DEADENYLATION_OF_MRNA	0.10189	0.67875	1.75146	0.00411
REACTOME_DEPOSITION_OF_NEW_CENPA_CONTAINING_NUCL EOSOMES_AT_THE_CENTROMERE	0.10204	0.71104	1.73884	0.01333
REACTOME_MITOTIC_G1_G1_S_PHASES	0.10254	0.59616	1.74061	0.01679
REACTOME_ACTIVATION_OF_ATR_IN_RESPONSE_TO_REPLICA TION_STRESS	0.10291	0.76476	1.75521	0.01331
REACTOME_FACTORS_INVOLVED_IN_MEGAKARYOCYTE_DEVE LOPMENT_AND_PLATELET_PRODUCTION	0.10315	0.44949	1.74232	0.00514
REACTOME_SIGNALING_BY_TGF_BETA_RECEPTOR_COMPLEX	0.10343	0.49065	1.74428	0.00179
PID_ATR_PATHWAY	0.10404	0.73865	1.94007	0.00185
REACTOME_G1_S_TRANSITION	0.10676	0.61844	1.73105	0.02052
REACTOME_E2F_MEDIATED_REGULATION_OF_DNA_REPLICATI ON	0.10906	0.67603	1.72596	0.02068
REACTOME_REGULATION_OF_MITOTIC_CELL_CYCLE	0.11120	0.63876	1.72137	0.02385
PID_FANCONI_PATHWAY	0.11789	0.73211	1.94110	0.00188
REACTOME_PACKAGING_OF_TELOMERE_ENDS	0.11850	0.68107	1.69037	0.02457
REACTOME_TRANSCRIPTIONAL_ACTIVITY_OF_SMAD2_SMAD3_ SMAD4_HETEROTRIMER	0.11936	0.51551	1.69138	0.00549
REACTOME_PROCESSING_OF_CAPPED_INTRON_CONTAINING_ PRE_MRNA	0.11967	0.55976	1.71013	0.02682
REACTOME_DNA_STRAND_ELONGATION	0.12033	0.76700	1.68599	0.01905
KEGG_VIBRIO_CHOLERAЕ_INFECTION	0.12037	0.51593	1.69642	0.01289
REACTOME_DNA_REPAIR	0.12056	0.51428	1.69222	0.02617
REACTOME_DOUBLE_STRAND_BREAK_REPAIR	0.12056	0.65993	1.69440	0.02574
REACTOME_RNA_POL_I_RNA_POL_III_AND_MITOCHONDRIAL_T RANSSCRIPTION	0.12074	0.50434	1.68373	0.01842
REACTOME_SYNTHESIS_OF_DNA	0.12095	0.64069	1.69964	0.02578
REACTOME_RECRUITMENT_OF_MITOTIC_CENTROSOME_PROT EINS_AND_COMPLEXES	0.12103	0.50539	1.70437	0.00914
REACTOME_S_PHASE	0.12119	0.61692	1.69748	0.03142
REACTOME_CHROMOSOME_MAINTENANCE	0.12125	0.67446	2.02219	0.00000
REACTOME_CLASS_I_MHC_MEDIATED_ANTIGEN_PROCESSING PRESENTATION	0.12180	0.44952	1.70082	0.00550
REACTOME_DOWNREGULATION_OF_SMAD2_3_SMAD4_TRANS CRIPTIONAL_ACTIVITY	0.12293	0.60638	1.70455	0.00787
REACTOME_HIV_INFECTION	0.12411	0.50023	1.67602	0.02421
REACTOME_MITOTIC_PROMETAPHASE	0.12438	0.70370	1.95132	0.00189
PID_FOXOPATHWAY	0.12481	0.48528	1.67349	0.00890
REACTOME_PROCESSING_OF_CAPPED_INTRONLESS_PRE_MR NA	0.12514	0.69404	1.67675	0.01934
REACTOME_DNA_REPLICATION	0.13022	0.68481	1.96305	0.00368
REACTOME_ACTIVATION_OF_THE_PRE_REPLICATIVE_COMPLE X	0.13127	0.77417	1.66250	0.01934
KEGG_DNA_REPLICATION	0.13129	0.71138	1.66065	0.03781
PID_MYC_ACTIVPATHWAY	0.13238	0.51319	1.66345	0.03825
REACTOME_RNA_POL_I_TRANSCRIPTION	0.13568	0.54073	1.65408	0.03097
BIOCARTA_MPR_PATHWAY	0.13574	0.53412	1.65234	0.01835
PID_NFAT_3PATHWAY	0.13881	0.45721	1.64747	0.01622

REACTOME_APC_CDC20_MEDIATED_DEGRADATION_OF_NEK2_A	0.14737	0.65242	1.63723	0.02985
REACTOME_EXTENSION_OF_TELOMERES	0.14908	0.71081	1.63372	0.03738
KEGG_BASAL_TRANSCRIPTION_FACTORS	0.15401	0.54087	1.62712	0.02124
KEGG_PROGESTERONE_MEDIATED_OOCYTE_MATURATION	0.16323	0.44447	1.61206	0.01068
BIOCARTA_G2_PATHWAY	0.16461	0.60109	1.61253	0.03137
REACTOME_CELL_CYCLE	0.16472	0.61694	2.03270	0.00361
REACTOME_RNA_POL_II_PRE_TRANSCRIPTION_EVENTS	0.16534	0.49104	1.60485	0.03683
KEGG_MISMATCH_REPAIR	0.16577	0.64843	1.61320	0.03512
KEGG_NUCLEOTIDE_EXCISION_REPAIR	0.16581	0.51854	1.60270	0.03795
REACTOME_M_G1_TRANSITION	0.16619	0.63197	1.60577	0.05639
PID_CDC42_PATHWAY	0.16620	0.44910	1.61468	0.01299
PID_FOXM1PATHWAY	0.16721	0.55348	1.60655	0.05128
REACTOME_HOST_INTERACTIONS_OF_HIV_FACTORS	0.16792	0.51664	1.59923	0.04135
REACTOME_HOMOLOGOUS_RECOMBINATION_REPAIR_OF_REPLICATION_INDEPENDENT_DOUBLE_STRAND_BREAKS	0.16930	0.65792	1.59629	0.03512
REACTOME_LAGGING_STRAND_SYNTHESIS	0.17241	0.71544	1.59174	0.04461
REACTOME_APC_C_CDC20_MEDIATED_DEGRADATION_OF_MITOTIC_PROTEINS	0.18120	0.60192	1.58256	0.07063
REACTOME_APC_C_CDH1_MEDIATED_DEGRADATION_OF_CDC20_AND_OTHER_APC_C_CDH1_TARGETED_PROTEINS_IN_LATE_MITOSIS_EARLY_G1	0.19813	0.59770	1.56708	0.07421
KEGG_SPLICEOSOME	0.20514	0.51304	1.55945	0.06434
KEGG_HOMOLOGOUS_RECOMBINATION	0.20534	0.60543	1.55755	0.06180
REACTOME_CYCLIN_E_ASSOCIATED_EVENTS_DURING_G1_S_TRANSITION	0.21934	0.56082	1.54500	0.08736
REACTOME_RNA_POL_I_PROMOTER_OPENING	0.22835	0.64999	1.53680	0.06857
PID_PI3KCIKTPATHWAY	0.23155	0.45818	1.53275	0.03018
REACTOME_ORC1_REMOVAL_FROM_CHROMATIN	0.23915	0.58463	1.52366	0.09888
REACTOME_REGULATION_OF_MRNA_STABILITY_BY_PROTEINS_THAT_BIND_AU_RICH_ELEMENTS	0.24074	0.51165	1.52429	0.07934
BIOCARTA_PROTEASOME_PATHWAY	0.24789	0.65029	1.51581	0.09478
PID_TELOMERASEPATHWAY	0.26377	0.41596	1.50312	0.02974
PID_INSULIN_GLUCOSE_PATHWAY	0.26954	0.48411	1.49755	0.06579
REACTOME_SCFSKP2_MEDIATED_DEGRADATION_OF_P27_P21	0.27322	0.57794	1.48846	0.11359
REACTOME_SIGNALING_BY_WNT	0.27351	0.51564	1.49312	0.08679
BIOCARTA_ACTINY_PATHWAY	0.27415	0.59575	1.48946	0.06903
REACTOME_ASSEMBLY_OF_THE_PRE_REPLICATIVE_COMPLEX	0.27479	0.59730	1.49072	0.10841
REACTOME_MHC_CLASS_II_ANTIGEN_PRESENTATION	0.29102	0.41678	1.47185	0.07196
SA_G1_AND_S_PHASES	0.29140	0.59326	1.47326	0.08812
PID_ECADHERIN_NASCENTAJ_PATHWAY	0.29210	0.47020	1.47468	0.05321
KEGG_UBIQUITIN_MEDIATED_PROTEOLYSIS	0.29307	0.54255	2.04425	0.00188
KEGG_PYRIMIDINE_METABOLISM	0.29385	0.45504	1.46808	0.07547
REACTOME_GENERIC_TRANSCRIPTION_PATHWAY	0.29591	0.40078	1.46340	0.09464
REACTOME_TRANSCRIPTION_COUPLED_NER_TC_NER	0.29715	0.50489	1.46426	0.08969
REACTOME_RNA_POL_I_TRANSCRIPTION_INITIATION	0.30228	0.52275	1.45781	0.08364
REACTOME_AUTODEGRADATION_OF_CDH1_BY_CDH1_APC_C	0.30504	0.55141	1.45445	0.13197
REACTOME_INSULIN_RECEPTOR_RECYCLING	0.31323	0.55970	1.44766	0.11132

PID_AURORA_A_PATHWAY	0.31582	0.48821	1.44424	0.08672
REACTOME_ACTIVATION_OF_NF_KAPPAB_IN_B_CELLS	0.32016	0.51224	1.43992	0.12037
REACTOME_P53_DEPENDENT_G1_DNA_DAMAGE_RESPONSE	0.32274	0.54136	1.43336	0.13458
REACTOME_SCF_BETA_TRCP_MEDIATED_DEGRADATION_OF_EMI1	0.32282	0.57213	1.43491	0.13645
REACTOME_NUCLEOTIDE_EXCISION_REPAIR	0.32432	0.46860	1.43563	0.09830
REACTOME_MRNA_SPLICING	0.33122	0.48743	1.42301	0.11686
REACTOME_RNA_POL_II_TRANSCRIPTION_PRE_INITIATION_AND_PROMOTER_OPENING	0.33273	0.47377	1.42540	0.09641
KEGG_PROTEASOME	0.33354	0.59200	1.42326	0.15789
KEGG_PROTEIN_EXPORT	0.33894	0.58179	1.41684	0.12398
BIOCARTA_G1_PATHWAY	0.34162	0.46818	1.41175	0.09009
PID_AJDISS_2PATHWAY	0.34311	0.40989	1.41251	0.06944
PID_MYC_PATHWAY	0.34561	0.45365	1.40774	0.08663
REACTOME_DOWNREGULATION_OF_TGF_BETA_RECEPTOR_SIGNALING	0.34578	0.49901	1.40418	0.10638

Appendix 8: Histopathological report of tumors obtained after injection of Control or TRIM24-HMECs

Group		CONTROL							Summary
Animal ID		2506	2508	2509	2510	2549	2550		
Species		Mouse	Mouse	Mouse	Mouse	Mouse	Mouse		
Genotype/Strain		C57/129	C57/129	C57/129	C57/129	C57/129	C57/129		
Organ		Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Incidence or Average Score of lesions	
Morphologic Diagnosis									
Xenograft tissue sample on the slide		P	P	P	P	P	P	6	
Dx: Fibrous tissue		P	P			P		3	
Dx: Epithelial cells intermixed with fibrous tissue				P	P		P	3	
Epithelial cells (percent of epithelial cells from the total tumor mass)		0	0	1	1	0	1	0.5	
Fibrous tissue (percent fibrous tissue and collagen deposition from the total tumor mass)		100	100	99	99	100	99	99.5	
Cellular atypia of neoplastic epithelial cells (grade 0-4)				0	0		0	0	
Anysokaryosis (grade 0-4)				0	0		0	0	
Mitotic rate (total number of mitotic epithelial cells within the xenograft tumor)				0	0		0	0	
Vascularization (grade 1-4)		1	1	1	1	1	1	1	
Grade of malignancy (grade 0-3)		0	0	0	0	0	0	0	
Inflammation		P	P	P	P	P		5	
Infiltration of Neutrophils and/or Macrophages		1	1	1	1	2		1.2	
Infiltration of Lymphocytes		0	0	0	0	0		0	

Group		TRIM24-1												Summary
Animal ID	Specie	2513	2515	2516	2517	2518	2519	2520	2531	2532	2533	2534	2535	
Genotype/Strain		Mouse CS7/129	Mouse CS7/129	Mouse CS7/129	Mouse CS7/129	Mouse CS7/129	Mouse CS7/129	Mouse CS7/129	Mouse CS7/129	Mouse CS7/129	Mouse CS7/129	Mouse CS7/129	Mouse CS7/129	
Organ Morphologic Diagnosis		Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Incidence or Average Score of lesions 12
Xenograft tissue sample on the slide Dx: Fibrous tissue		P	P	P	P	P	P	P	P	P	P	P	P	
Dx: Epithelial cells intermixed with fibrous tissue		P	P	P	P	P	P	P	P	P	P	P	P	12
Epithelial cells (percent of epithelial cells from the total tumor mass)		5	25	35	30	40	40	30	35	20	35	1	2	24.8
Fibrous tissue (percent fibrous tissue and collagen deposition from the total tumor mass)		95	75	65	70	60	60	70	65	80	65	99	98	75.2
Cellular atypia of neoplastic epithelial cells (grade 0-4)		2	1	4	2	3	4	2	4	2	3	0	1	2.3
Anysokaryosis (grade 0-4)		1	1	3	2	3	3	2	3	3	3	0	2	2.2
Mitotic rate (total number of mitotic epithelial cells within the xenograft tumor)		1	0	3	0	2	2	0	0	0	1	0	1	0.8
Vascularization (grade 1-4)		1	2	2	1	2	2	2	2	2	2	2	2	1.8
Grade of malignancy (grade 0-3)		1	1	3	2	2	3	2	3	2	2	0	1	1.8
Inflammation		P	P	P	P	P	P	P	P	P	P	P	P	12
Infiltration of Neutrophils and/or Macrophages		3	2	3	3	2	3	2	2	4	4	3	2	3
Infiltration of Lymphocytes		1	1	2	1	2	2	1	3	3	1	0	0	1

Group		TRIM24-2														Summary
Animal ID	2528	2521	2522	2525	2526	2527	2529	2530	2536	2537	2540	2541	2542			
Species	Mouse	Mouse	Mouse	Mouse	Mouse	Mouse	Mouse	Mouse	Mouse	Mouse	Mouse	Mouse	Mouse			
Genotype/Strain	C57/129	C57/129	C57/129	C57/129	C57/129	C57/129	C57/129	C57/129	C57/129	C57/129	C57/129	C57/129	C57/129			
Organ	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade			
Morphologic Diagnosis	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade	Lesion Grade			Incidence or Average Score of lesions
Xenograft tissue sample on the slide		P	P	P	P	P	P	P	P	P	P	P	P			12
Dx: Fibrous tissue																0
Dx: Epithelial cells intermixed with fibrous tissue		P	P	P	P	P	P	P	P	P	P	P	P			12
Epithelial cells (percent of epithelial cells from the total tumor mass)		50	40	35	20	35	1	35	20	25	35	60	25			31.8
Fibrous tissue (percent fibrous tissue and collagen deposition from the total tumor mass)		50	60	65	80	65	99	65	80	75	65	40	75			68.3
Cellular atypia of neoplastic epithelial cells (grade 0-4)		4	4	4	2	4	1	4	2	3	4	4	3			3.3
Anysokaryosis (grade 0-4)		3	3	3	3	3	1	4	2	3	4	4	3			3.0
Mitotic rate (total number of mitotic epithelial cells within the xenograft tumor)		0	2	2	1	0	0	0	0	2	0	0	0			0.6
Vascularization (grade 1-4)		2	2	2	1	2	1	2	1	1	2	2	2			1.7
Grade of malignancy (grade 0-3)		3	3	3	1	3	0	3	1	2	3	3	2			2.3
Inflammation		P	P	P	P	P	P	P	P	P	P	P	P			12
Infiltration of Neutrophils and/or Macrophages		2	2	1	1	2	1	2	1	3	3	3	2			2
Infiltration of Lymphocytes		3	2	2	1	0	0	0	0	1	3	0	2			1

Appendix 9: List of deregulated genes in TRIM24-OE MCF10A cells by HTG

Edgeseq analysis

Upregulated genes							
		Vector	Normalized	FC	WT	Normalized	FC
1	FYN	18	0.001	1	101	0.005	6.19
2	MMP2	197	0.010	1	978	0.053	5.47
3	IGFBP2	90	0.004	1	423	0.023	5.18
4	CITED2	1211	0.060	1	4958	0.270	4.51
5	ITGAM	3	0.000	1	12	0.001	4.41
6	CCL17	11	0.001	1	43	0.002	4.31
7	F2R	117	0.006	1	448	0.024	4.22
8	NOV	5	0.000	1	19	0.001	4.19
9	TIMP1	5089	0.251	1	18943	1.031	4.10
10	STC1	19	0.001	1	62	0.003	3.60
11	NEIL1	12	0.001	1	38	0.002	3.49
12	BMP4	82	0.004	1	259	0.014	3.48
13	DKK1	2435	0.120	1	7608	0.414	3.44
14	HSF4	15	0.001	1	46	0.003	3.38
15	ADRA2A	4	0.000	1	12	0.001	3.31
16	FCGR1A	7	0.000	1	21	0.001	3.31
17	LFNG	13	0.001	1	39	0.002	3.31
18	APBB1	12	0.001	1	34	0.002	3.12
19	ANGPT1	202	0.010	1	572	0.031	3.12
20	TXNRD1	1960	0.097	1	5528	0.301	3.11
21	EFNA2	4	0.000	1	11	0.001	3.03
22	FAT4	88	0.004	1	241	0.013	3.02
23	PDCD1LG2	7	0.000	1	19	0.001	2.99
24	IGFBP4	314	0.016	1	843	0.046	2.96
25	IL27	3	0.000	1	8	0.000	2.94
26	FBN1	309	0.015	1	797	0.043	2.84
27	FOXO1	195	0.010	1	484	0.026	2.74
28	GSTM1	27	0.001	1	66	0.004	2.69
29	PRMT8	10	0.000	1	24	0.001	2.65
30	HSP90AA1	15	0.001	1	36	0.002	2.65
31	PBX1	80	0.004	1	192	0.010	2.65
32	CARD11	33	0.002	1	77	0.004	2.57
33	CLDN3	6	0.000	1	14	0.001	2.57
34	TNFRSF8	12	0.001	1	28	0.002	2.57
35	SLC7A11	110	0.005	1	256	0.014	2.57
36	CD68	269	0.013	1	619	0.034	2.54
37	RELN	12	0.001	1	27	0.001	2.48
38	SULT2A1	13	0.001	1	29	0.002	2.46
39	HPSE	123	0.006	1	272	0.015	2.44
40	IRS1	134	0.007	1	295	0.016	2.43
41	TAT	12	0.001	1	26	0.001	2.39
42	S100A4	497	0.025	1	1061	0.058	2.35
43	DAPK1	65	0.003	1	137	0.007	2.32
44	PPAP2B	71	0.004	1	146	0.008	2.27
45	WNT5A	218	0.011	1	448	0.024	2.27
46	ID4	5	0.000	1	10	0.001	2.20
47	GNLY	21	0.001	1	42	0.002	2.20
48	BTC	26	0.001	1	52	0.003	2.20
49	TCL1A	30	0.001	1	60	0.003	2.20
50	PROK1	35	0.002	1	70	0.004	2.20
51	MAF	3	0.000	1	6	0.000	2.20
52	PDPN	49	0.002	1	98	0.005	2.20
53	SIX1	41	0.002	1	81	0.004	2.18
54	GADD45B	407	0.020	1	800	0.044	2.17

55	MAGEA4	26	0.001	1	51	0.003	2.16
56	TSC1	51	0.003	1	99	0.005	2.14
57	MYBL1	46	0.002	1	89	0.005	2.13
58	MAP1LC3A	85	0.004	1	164	0.009	2.13
59	AXL	348	0.017	1	669	0.036	2.12
60	LTF	25	0.001	1	48	0.003	2.12
61	COL18A1	64	0.003	1	122	0.007	2.10
62	GPB1	19	0.001	1	36	0.002	2.09
63	IFNW1	18	0.001	1	34	0.002	2.08
64	GAS6	637	0.031	1	1197	0.065	2.07
65	HOXB13	16	0.001	1	30	0.002	2.07
66	SYP	30	0.001	1	56	0.003	2.06
67	SCUBE1	7	0.000	1	13	0.001	2.05
68	IL11	33	0.002	1	60	0.003	2.00
69	TOP3B	21	0.001	1	38	0.002	1.99
70	GSN	107	0.005	1	193	0.011	1.99
71	CDKN2C	67	0.003	1	120	0.007	1.97
72	ABCC3	2222	0.110	1	3979	0.217	1.97
73	CST6	57	0.003	1	102	0.006	1.97
74	NQO1	6428	0.317	1	11484	0.625	1.97
75	FLRT1	14	0.001	1	25	0.001	1.97
76	CYR61	743	0.037	1	1316	0.072	1.95
77	CDKN2D	39	0.002	1	69	0.004	1.95
78	FGF13	17	0.001	1	30	0.002	1.95
79	IKBK1	17	0.001	1	30	0.002	1.95
80	KLF17	17	0.001	1	30	0.002	1.95
81	EPS8	515	0.025	1	908	0.049	1.94
82	BMF	83	0.004	1	146	0.008	1.94
83	SOX3	28	0.001	1	49	0.003	1.93
84	FZD2	183	0.009	1	320	0.017	1.93
85	DUSP10	132	0.007	1	230	0.013	1.92
86	ZEB1	160	0.008	1	278	0.015	1.92
87	S1PR1	34	0.002	1	59	0.003	1.91
88	PFKFB1	44	0.002	1	76	0.004	1.90
89	BNIP3L	310	0.015	1	535	0.029	1.90
90	MAOB	46	0.002	1	79	0.004	1.89
91	IGFBP6	653	0.032	1	1120	0.061	1.89
92	GFAP	28	0.001	1	48	0.003	1.89
93	PRDM14	28	0.001	1	48	0.003	1.89
94	CAMKK1	31	0.002	1	53	0.003	1.88
95	STAT2	344	0.017	1	586	0.032	1.88
96	DCK	10	0.000	1	17	0.001	1.87
97	DNAJC4	76	0.004	1	129	0.007	1.87
98	PDCD1	29	0.001	1	49	0.003	1.86
99	ACADS	57	0.003	1	96	0.005	1.86
100	CLIC3	119	0.006	1	199	0.011	1.84
101	ARNT2	9	0.000	1	15	0.001	1.84
102	FGF22	18	0.001	1	30	0.002	1.84
103	PDZK1	53	0.003	1	88	0.005	1.83
104	ZBTB16	449	0.022	1	745	0.041	1.83
105	IDH2	315	0.016	1	522	0.028	1.83
106	RPS6KB1	20	0.001	1	33	0.002	1.82
107	SMAD7	134	0.007	1	221	0.012	1.82
108	IL2RB	37	0.002	1	61	0.003	1.82
109	NUDT13	17	0.001	1	28	0.002	1.82
110	SLCO2B1	17	0.001	1	28	0.002	1.82
111	MAP3K1	198	0.010	1	326	0.018	1.82
112	CALML5	39	0.002	1	64	0.003	1.81
113	NRP1	204	0.010	1	334	0.018	1.80
114	HSPA6	11	0.001	1	18	0.001	1.80
115	DNAJB2	63	0.003	1	103	0.006	1.80
116	TGFB1	49	0.002	1	80	0.004	1.80

117	GZMH	27	0.001	1	44	0.002	1.80
118	RUNX3	27	0.001	1	44	0.002	1.80
119	DMC1	8	0.000	1	13	0.001	1.79
120	TIMP2	842	0.042	1	1368	0.074	1.79
121	EYA1	34	0.002	1	55	0.003	1.78
122	CTSD	3612	0.178	1	5810	0.316	1.77
123	CIDEA	33	0.002	1	53	0.003	1.77
124	BCL2L1	930	0.046	1	1489	0.081	1.76
125	CREB5	25	0.001	1	40	0.002	1.76
126	RAMP1	157	0.008	1	250	0.014	1.76
127	NKX2_1	22	0.001	1	35	0.002	1.75
128	AKR1C3	1864	0.092	1	2949	0.160	1.74
129	ATP6V1G2	45	0.002	1	71	0.004	1.74
130	TRADD	158	0.008	1	249	0.014	1.74
131	TMEFF1	14	0.001	1	22	0.001	1.73
132	UTF1	14	0.001	1	22	0.001	1.73
133	PPP2R1B	44	0.002	1	69	0.004	1.73
134	THBS1	12509	0.618	1	19505	1.061	1.72
135	CYP2U1	104	0.005	1	162	0.009	1.72
136	CNPY4	38	0.002	1	59	0.003	1.71
137	MLPH	154	0.008	1	239	0.013	1.71
138	EFNA5	956	0.047	1	1483	0.081	1.71
139	DNAJC22	60	0.003	1	93	0.005	1.71
140	ID1	2794	0.138	1	4329	0.236	1.71
141	PLAT	473	0.023	1	732	0.040	1.71
142	AHNAK2	145	0.007	1	224	0.012	1.70
143	SPINK1	13	0.001	1	20	0.001	1.70
144	FGFR1	160	0.008	1	246	0.013	1.69
145	NOL3	54	0.003	1	83	0.005	1.69
146	SQSTM1	3038	0.150	1	4652	0.253	1.69
147	CLCF1	121	0.006	1	185	0.010	1.69
148	COL1A2	19	0.001	1	29	0.002	1.68
149	NCOA1	78	0.004	1	119	0.006	1.68
150	CCL24	21	0.001	1	32	0.002	1.68
151	HNF1B	23	0.001	1	35	0.002	1.68
152	MAGED1	493	0.024	1	745	0.041	1.67
153	DAAM1	96	0.005	1	145	0.008	1.67
154	NGFRAP1	49	0.002	1	74	0.004	1.66
155	ZEB2	106	0.005	1	160	0.009	1.66
156	APH1B	95	0.005	1	143	0.008	1.66
157	ID3	320	0.016	1	481	0.026	1.66
158	LOX	348	0.017	1	523	0.028	1.66
159	CKMT2	40	0.002	1	60	0.003	1.65
160	KLF12	70	0.003	1	105	0.006	1.65
161	CCDC103	48	0.002	1	72	0.004	1.65
162	FBX1	180468	8.909	1	270144	14.702	1.65
163	ITGB2	168	0.008	1	251	0.014	1.65
164	FAS	61	0.003	1	91	0.005	1.64
165	GLI2	31	0.002	1	46	0.003	1.64
166	HECTD4	54	0.003	1	80	0.004	1.63
167	DHH	23	0.001	1	34	0.002	1.63
168	FRS3	38	0.002	1	56	0.003	1.62
169	IFNB1	17	0.001	1	25	0.001	1.62
170	ADORA2A	15	0.001	1	22	0.001	1.62
171	PLG	54	0.003	1	79	0.004	1.61
172	COL11A1	26	0.001	1	38	0.002	1.61
173	CD5	22	0.001	1	32	0.002	1.60
174	FLJ10474	275	0.014	1	400	0.022	1.60
175	PDGFC	416	0.021	1	605	0.033	1.60
176	CYP1B1	436	0.022	1	634	0.035	1.60
177	TMEM45B	20	0.001	1	29	0.002	1.60
178	NUMBL	55	0.003	1	79	0.004	1.58

179	PIK3R1	786	0.039	1	1128	0.061	1.58
180	CYCS	23	0.001	1	33	0.002	1.58
181	SLC10A1	26	0.001	1	37	0.002	1.57
182	HSPB8	19	0.001	1	27	0.001	1.57
183	PPP3R2	19	0.001	1	27	0.001	1.57
184	TNFRSF1B	38	0.002	1	54	0.003	1.57
185	FGFRL1	278	0.014	1	395	0.021	1.57
186	TMEM74B	50	0.002	1	71	0.004	1.57
187	HAVCR2	43	0.002	1	61	0.003	1.56
188	TLR4	86	0.004	1	122	0.007	1.56
189	CACNB4	12	0.001	1	17	0.001	1.56
190	PRKCA	169	0.008	1	239	0.013	1.56
191	LIN28B	29	0.001	1	41	0.002	1.56
192	SPP1	29	0.001	1	41	0.002	1.56
193	IL2RG	49	0.002	1	69	0.004	1.55
194	PCOLCE	27	0.001	1	38	0.002	1.55
195	MEF2C	32	0.002	1	45	0.002	1.55
196	EGR3	62	0.003	1	87	0.005	1.55
197	CD3D	40	0.002	1	56	0.003	1.54
198	CCS	183	0.009	1	255	0.014	1.54
199	CCL18	41	0.002	1	57	0.003	1.53
200	GAMT	124	0.006	1	171	0.009	1.52
201	PLK2	1449	0.072	1	1997	0.109	1.52
202	LYN	43	0.002	1	59	0.003	1.51
203	MEIS1	43	0.002	1	59	0.003	1.51
204	RPTOR	43	0.002	1	59	0.003	1.51
205	AQP4	35	0.002	1	48	0.003	1.51
206	S100B	35	0.002	1	48	0.003	1.51
207	NR5A2	27	0.001	1	37	0.002	1.51
208	LATS2	73	0.004	1	100	0.005	1.51
209	RHOA	141	0.007	1	193	0.011	1.51
210	INSR	57	0.003	1	78	0.004	1.51
211	GSR	949	0.047	1	1297	0.071	1.51
212	PDK2	123	0.006	1	168	0.009	1.51
213	CDH13	733	0.036	1	1000	0.054	1.50
214	NOTCH4	11	0.001	1	15	0.001	1.50
215	MKNK1	22	0.001	1	30	0.002	1.50
216	CCL26	44	0.002	1	60	0.003	1.50
217	BMP8B	39	0.002	1	53	0.003	1.50
218	ITM2A	39	0.002	1	53	0.003	1.50
219	ID2	67	0.003	1	91	0.005	1.50
220	WISP2	109	0.005	1	148	0.008	1.50
221	SLC38A2	4109	0.203	1	5577	0.304	1.50
222	SLC7A7	28	0.001	1	38	0.002	1.50
223	SIRT2	198	0.010	1	268	0.015	1.49
224	SULT1A1	34	0.002	1	46	0.003	1.49
225	ACSL5	20	0.001	1	27	0.001	1.49
226	HSPB2	23	0.001	1	31	0.002	1.49
227	NRG1	116	0.006	1	156	0.008	1.48
228	CTGF	644	0.032	1	866	0.047	1.48
229	CFTR	38	0.002	1	51	0.003	1.48
230	GRN	503	0.025	1	675	0.037	1.48
231	NFATC3	117	0.006	1	157	0.009	1.48
232	SHC3	44	0.002	1	59	0.003	1.48
233	FMO5	53	0.003	1	71	0.004	1.48
234	SERPINE1	3651	0.180	1	4874	0.265	1.47
235	IGF1	18	0.001	1	24	0.001	1.47
236	ABCB5	36	0.002	1	48	0.003	1.47
237	NR1H3	45	0.002	1	60	0.003	1.47
238	CMTM1	12	0.001	1	16	0.001	1.47
239	RAD52	21	0.001	1	28	0.002	1.47
240	SLCO1A2	27	0.001	1	36	0.002	1.47

241	ABCC11	43	0.002	1	57	0.003	1.46
242	ZNF189	86	0.004	1	114	0.006	1.46
243	VEGFA	160	0.008	1	212	0.012	1.46
244	CCL13	37	0.002	1	49	0.003	1.46
245	SERPINC1	37	0.002	1	49	0.003	1.46
246	NCOA2	145	0.007	1	192	0.010	1.46
247	ACVR2A	59	0.003	1	78	0.004	1.46
248	GATA2	56	0.003	1	74	0.004	1.46
249	KRT8	4165	0.206	1	5501	0.299	1.46
250	CEBPE	22	0.001	1	29	0.002	1.45
251	HDAC2	44	0.002	1	58	0.003	1.45
252	SF3A1	104	0.005	1	137	0.007	1.45
253	CREBBP	120	0.006	1	158	0.009	1.45
254	SALL4	98	0.005	1	129	0.007	1.45
255	CDH2	541	0.027	1	712	0.039	1.45
256	PMS2	362	0.018	1	476	0.026	1.45
257	DACH1	35	0.002	1	46	0.003	1.45
258	F8	83	0.004	1	109	0.006	1.45
259	MESP1	32	0.002	1	42	0.002	1.45
260	IGFBP7	224	0.011	1	294	0.016	1.45
261	OR10J3	71	0.004	1	93	0.005	1.44
262	CTSH	343	0.017	1	449	0.024	1.44
263	NRP2	230	0.011	1	301	0.016	1.44
264	BDNF	244	0.012	1	319	0.017	1.44
265	GATA4	36	0.002	1	47	0.003	1.44
266	CCT6B	56	0.003	1	73	0.004	1.44
267	FGF2	478	0.024	1	620	0.034	1.43
268	IRAK3	27	0.001	1	35	0.002	1.43
269	FEN1	115	0.006	1	149	0.008	1.43
270	HIF1A	2835	0.140	1	3673	0.200	1.43
271	C1orf86	85	0.004	1	110	0.006	1.43
272	BTG1	177	0.009	1	229	0.012	1.43
273	PRICKLE1	86	0.004	1	111	0.006	1.42
274	NPM1	52	0.003	1	67	0.004	1.42
275	MGST1	6186	0.305	1	7968	0.434	1.42
276	ANPEP	3038	0.150	1	3911	0.213	1.42
277	LAT	7	0.000	1	9	0.000	1.42
278	DDB2	70	0.003	1	90	0.005	1.42
279	LRRK2	116	0.006	1	149	0.008	1.42
280	TIPARP	991	0.049	1	1272	0.069	1.41
281	SUFU	46	0.002	1	59	0.003	1.41
282	TFDP2	235	0.012	1	301	0.016	1.41
283	MCM7	25	0.001	1	32	0.002	1.41
284	DMD	43	0.002	1	55	0.003	1.41
285	MT1A	18	0.001	1	23	0.001	1.41
286	NRG3	18	0.001	1	23	0.001	1.41
287	RUNX2	310	0.015	1	396	0.022	1.41
288	MLF1	44	0.002	1	56	0.003	1.40
289	SLC22A9	44	0.002	1	56	0.003	1.40
290	SNAI2	228	0.011	1	290	0.016	1.40
291	STK32A	37	0.002	1	47	0.003	1.40
292	GATA3	248	0.012	1	315	0.017	1.40
293	HMOX1	89	0.004	1	113	0.006	1.40
294	LETMD1	293	0.014	1	372	0.020	1.40
295	FLNC	79	0.004	1	100	0.005	1.40
296	CHI3L1	121	0.006	1	153	0.008	1.39
297	PRF1	38	0.002	1	48	0.003	1.39
298	LRP2	42	0.002	1	53	0.003	1.39
299	SLC10A2	65	0.003	1	82	0.004	1.39
300	PF4	50	0.002	1	63	0.003	1.39
301	HOXA11	27	0.001	1	34	0.002	1.39
302	NTRK1	27	0.001	1	34	0.002	1.39

303	LIN28A	35	0.002	1	44	0.002	1.39
304	ITGB8	494	0.024	1	620	0.034	1.38
305	CD40LG	59	0.003	1	74	0.004	1.38
306	ENDOD1	244	0.012	1	306	0.017	1.38
307	DLC1	185	0.009	1	232	0.013	1.38
308	NOS2	4	0.000	1	5	0.000	1.38
309	COL9A3	16	0.001	1	20	0.001	1.38
310	TOLLIP	20	0.001	1	25	0.001	1.38
311	KLK2	36	0.002	1	45	0.002	1.38
312	DES	12	0.001	1	15	0.001	1.38
313	NPPB	28	0.001	1	35	0.002	1.38
314	PRKACG	28	0.001	1	35	0.002	1.38
315	DNAJC12	57	0.003	1	71	0.004	1.37
316	CDK5	322	0.016	1	401	0.022	1.37
317	XCL2	49	0.002	1	61	0.003	1.37
318	DNAJB12	239	0.012	1	297	0.016	1.37
319	ACKR3	33	0.002	1	41	0.002	1.37
320	PECAM1	58	0.003	1	72	0.004	1.37
321	CAPNS1	1055	0.052	1	1309	0.071	1.37
322	CCL7	25	0.001	1	31	0.002	1.37
323	CDH3	50	0.002	1	62	0.003	1.37
324	GHR	63	0.003	1	78	0.004	1.36
325	PARP3	21	0.001	1	26	0.001	1.36
326	LY96	38	0.002	1	47	0.003	1.36
327	NR0B1	17	0.001	1	21	0.001	1.36
328	CYBA	51	0.003	1	63	0.003	1.36
329	SLC22A2	47	0.002	1	58	0.003	1.36
330	PTGR1	270	0.013	1	333	0.018	1.36
331	PPP2R2B	52	0.003	1	64	0.003	1.36
332	NEIL3	139	0.007	1	171	0.009	1.36
333	ITGB4	2049	0.101	1	2518	0.137	1.35
334	OSM	44	0.002	1	54	0.003	1.35
335	ATF1	194	0.010	1	238	0.013	1.35
336	ETV1	75	0.004	1	92	0.005	1.35
337	CPT1C	40	0.002	1	49	0.003	1.35
338	TRAF3	80	0.004	1	98	0.005	1.35
339	ORM2	49	0.002	1	60	0.003	1.35
340	PIK3R2	640	0.032	1	783	0.043	1.35
341	DNAJB5	36	0.002	1	44	0.002	1.35
342	RPA3	36	0.002	1	44	0.002	1.35
343	CTSB	494	0.024	1	603	0.033	1.35
344	MCM4	23	0.001	1	28	0.002	1.34
345	IRF2	92	0.005	1	112	0.006	1.34
346	MAP2K6	37	0.002	1	45	0.002	1.34
347	SIRT4	42	0.002	1	51	0.003	1.34
348	UBE2Z	257	0.013	1	312	0.017	1.34
349	TWIST1	234	0.012	1	284	0.015	1.34
350	IFIT2	61	0.003	1	74	0.004	1.34
351	CYLD	90	0.004	1	109	0.006	1.34
352	LGALS3	488	0.024	1	591	0.032	1.34
353	EPOR	38	0.002	1	46	0.003	1.33
354	PRMT6	48	0.002	1	58	0.003	1.33
355	MAP3K8	87	0.004	1	105	0.006	1.33
356	IL18	1042	0.051	1	1257	0.068	1.33
357	PAG1	39	0.002	1	47	0.003	1.33
358	TEP1	39	0.002	1	47	0.003	1.33
359	CES2	205	0.010	1	247	0.013	1.33
360	PAF1	183	0.009	1	220	0.012	1.33
361	NFAT5	213	0.011	1	256	0.014	1.32
362	EXT1	763	0.038	1	917	0.050	1.32
363	GLUD1	774	0.038	1	930	0.051	1.32
364	FRMD6	973	0.048	1	1168	0.064	1.32

365	F2	5	0.000	1	6	0.000	1.32
366	TEK	5	0.000	1	6	0.000	1.32
367	UGT2B7	20	0.001	1	24	0.001	1.32
368	CAPN3	70	0.003	1	84	0.005	1.32
369	CD63	6672	0.329	1	7991	0.435	1.32
370	HHAT	51	0.003	1	61	0.003	1.32
371	WNT10A	46	0.002	1	55	0.003	1.32
372	DNAJB14	115	0.006	1	137	0.007	1.31
373	TTF1	63	0.003	1	75	0.004	1.31
374	CAPN5	116	0.006	1	138	0.008	1.31
375	ERG	27	0.001	1	32	0.002	1.31
376	ERCC6	254	0.013	1	301	0.016	1.31
377	FGF9	38	0.002	1	45	0.002	1.31
378	SOCS5	478	0.024	1	566	0.031	1.31
379	OTX2	49	0.002	1	58	0.003	1.30
380	COL4A5	60	0.003	1	71	0.004	1.30
381	MAP3K5	224	0.011	1	265	0.014	1.30
382	CKB	44	0.002	1	52	0.003	1.30
383	C1QA	50	0.002	1	59	0.003	1.30
384	TERF2IP	179	0.009	1	211	0.011	1.30
385	MAP1B	247	0.012	1	291	0.016	1.30
386	SH2B1	225	0.011	1	265	0.014	1.30
387	SELM	467	0.023	1	550	0.030	1.30
388	NOG	17	0.001	1	20	0.001	1.30
389	FN1	31349	1.548	1	36875	2.007	1.30

Down-regulated genes

		Vector	Normalized	FC	WT	Normalized	FC
1	OLIG2	37	0.002	1	27	0.001	0.80
2	SLC15A1	37	0.002	1	27	0.001	0.80
3	SLC25A4	85	0.004	1	62	0.003	0.80
4	NR1I2	48	0.002	1	35	0.002	0.80
5	PRKG1	48	0.002	1	35	0.002	0.80
6	RAD23B	3379	0.167	1	2462	0.134	0.80
7	RBX1	1001	0.049	1	729	0.040	0.80
8	POLD3	187	0.009	1	136	0.007	0.80
9	MIA	11	0.001	1	8	0.000	0.80
10	ERBB4	22	0.001	1	16	0.001	0.80
11	GDF3	33	0.002	1	24	0.001	0.80
12	PFKFB4	77	0.004	1	56	0.003	0.80
13	DNAJC10	1052	0.052	1	765	0.042	0.80
14	AIFM1	359	0.018	1	261	0.014	0.80
15	RAD51B	183	0.009	1	133	0.007	0.80
16	RAB25	84	0.004	1	61	0.003	0.80
17	IL12B	51	0.003	1	37	0.002	0.80
18	EMP1	3160	0.156	1	2291	0.125	0.80
19	CASP4	457	0.023	1	331	0.018	0.80
20	CX3CL1	29	0.001	1	21	0.001	0.80
21	XRCC3	174	0.009	1	126	0.007	0.80
22	WHSC1	756	0.037	1	547	0.030	0.80
23	AICDA	47	0.002	1	34	0.002	0.80
24	FOXA2	47	0.002	1	34	0.002	0.80
25	MPO	36	0.002	1	26	0.001	0.80
26	NCOA3	928	0.046	1	670	0.036	0.80
27	BAMBI	212	0.010	1	153	0.008	0.80
28	LTA4H	825	0.041	1	595	0.032	0.80
29	COX5A	1697	0.084	1	1223	0.067	0.79
30	POU5F1	243	0.012	1	175	0.010	0.79
31	IL20RB	443	0.022	1	319	0.017	0.79
32	FZD1	100	0.005	1	72	0.004	0.79
33	CSNK1A1	2277	0.112	1	1639	0.089	0.79
34	RAD17	339	0.017	1	244	0.013	0.79

35	CDKL5	57	0.003	1	41	0.002	0.79
36	SFRP2	57	0.003	1	41	0.002	0.79
37	KNG1	32	0.002	1	23	0.001	0.79
38	AIMP2	312	0.015	1	224	0.012	0.79
39	HSPA8	19076	0.942	1	13686	0.745	0.79
40	CRTC2	473	0.023	1	339	0.018	0.79
41	BCR	522	0.026	1	374	0.020	0.79
42	SLC22A7	74	0.004	1	53	0.003	0.79
43	EXOC3	155	0.008	1	111	0.006	0.79
44	SMAD5	859	0.042	1	614	0.033	0.79
45	TGFB2	1537	0.076	1	1098	0.060	0.79
46	IL12A	35	0.002	1	25	0.001	0.79
47	IL22RA1	42	0.002	1	30	0.002	0.79
48	PIK3CG	49	0.002	1	35	0.002	0.79
49	POLD1	63	0.003	1	45	0.002	0.79
50	ERCC8	289	0.014	1	206	0.011	0.79
51	CDK2	452	0.022	1	322	0.018	0.79
52	RBPMS2	66	0.003	1	47	0.003	0.79
53	ABCB4	59	0.003	1	42	0.002	0.78
54	DAPL1	45	0.002	1	32	0.002	0.78
55	WNT1	45	0.002	1	32	0.002	0.78
56	FLOT2	1306	0.064	1	928	0.051	0.78
57	AFF1	228	0.011	1	162	0.009	0.78
58	TP53	677	0.033	1	481	0.026	0.78
59	ITGA5	556	0.027	1	395	0.021	0.78
60	MIF	4457	0.220	1	3165	0.172	0.78
61	CARM1	959	0.047	1	681	0.037	0.78
62	ACSL3	438	0.022	1	311	0.017	0.78
63	FZD9	31	0.002	1	22	0.001	0.78
64	SIN3B	234	0.012	1	166	0.009	0.78
65	NTF3	55	0.003	1	39	0.002	0.78
66	HSPA12A	48	0.002	1	34	0.002	0.78
67	RASGRF1	48	0.002	1	34	0.002	0.78
68	BNIP2	1265	0.062	1	896	0.049	0.78
69	CXCL10	65	0.003	1	46	0.003	0.78
70	RTN1	65	0.003	1	46	0.003	0.78
71	TFPI2	65	0.003	1	46	0.003	0.78
72	SMC4	328	0.016	1	232	0.013	0.78
73	WNT6	58	0.003	1	41	0.002	0.78
74	LIFR	133	0.007	1	94	0.005	0.78
75	MECOM	92	0.005	1	65	0.004	0.78
76	RHOA	3524	0.174	1	2488	0.135	0.78
77	HAND1	85	0.004	1	60	0.003	0.78
78	CD74	102	0.005	1	72	0.004	0.78
79	PML	469	0.023	1	331	0.018	0.78
80	KEAP1	122	0.006	1	86	0.005	0.78
81	HMGB1	850	0.042	1	599	0.033	0.78
82	NRIP1	816	0.040	1	575	0.031	0.78
83	ACTR2	5221	0.258	1	3679	0.200	0.78
84	IL33	71	0.004	1	50	0.003	0.78
85	NRG2	27	0.001	1	19	0.001	0.78
86	AP2B1	1088	0.054	1	765	0.042	0.78
87	FZD3	138	0.007	1	97	0.005	0.77
88	CES1	84	0.004	1	59	0.003	0.77
89	DNAJC7	225	0.011	1	158	0.009	0.77
90	DLK1	47	0.002	1	33	0.002	0.77
91	ZIC1	47	0.002	1	33	0.002	0.77
92	ZNF420	104	0.005	1	73	0.004	0.77
93	PPARD	181	0.009	1	127	0.007	0.77
94	SPRY1	67	0.003	1	47	0.003	0.77
95	IRF3	757	0.037	1	531	0.029	0.77
96	MOB1B	442	0.022	1	310	0.017	0.77

97	ATG7	147	0.007	1	103	0.006	0.77
98	BRIP1	197	0.010	1	138	0.008	0.77
99	FUS	1865	0.092	1	1306	0.071	0.77
100	CTLA4	30	0.001	1	21	0.001	0.77
101	LRP1B	40	0.002	1	28	0.002	0.77
102	RB1CC1	50	0.002	1	35	0.002	0.77
103	PHB2	3448	0.170	1	2412	0.131	0.77
104	RAD23A	998	0.049	1	698	0.038	0.77
105	BIRC5	156	0.008	1	109	0.006	0.77
106	USMG5	3786	0.187	1	2643	0.144	0.77
107	CCNF	139	0.007	1	97	0.005	0.77
108	CNTFR	33	0.002	1	23	0.001	0.77
109	CYP2C19	56	0.003	1	39	0.002	0.77
110	PIAS3	658	0.032	1	458	0.025	0.77
111	COL4A2	1036	0.051	1	721	0.039	0.77
112	FES	23	0.001	1	16	0.001	0.77
113	SLC11A1	46	0.002	1	32	0.002	0.77
114	PDGFRB	59	0.003	1	41	0.002	0.77
115	CMKLR1	36	0.002	1	25	0.001	0.77
116	PHF6	436	0.022	1	302	0.016	0.76
117	PRL	52	0.003	1	36	0.002	0.76
118	SCN3A	52	0.003	1	36	0.002	0.76
119	SEC61G	1832	0.090	1	1268	0.069	0.76
120	CAD	341	0.017	1	236	0.013	0.76
121	BAG4	201	0.010	1	139	0.008	0.76
122	UBE2I	1147	0.057	1	793	0.043	0.76
123	BCAT1	68	0.003	1	47	0.003	0.76
124	NUP62	680	0.034	1	470	0.026	0.76
125	PIM2	97	0.005	1	67	0.004	0.76
126	FASLG	42	0.002	1	29	0.002	0.76
127	CEBPB	3662	0.181	1	2528	0.138	0.76
128	CDK7	358	0.018	1	247	0.013	0.76
129	NDC80	216	0.011	1	149	0.008	0.76
130	AQP7	303	0.015	1	209	0.011	0.76
131	C5AR1	58	0.003	1	40	0.002	0.76
132	SRSF1	1184	0.058	1	815	0.044	0.76
133	ADRA2B	93	0.005	1	64	0.003	0.76
134	DISP2	64	0.003	1	44	0.002	0.76
135	PRKAR1B	80	0.004	1	55	0.003	0.76
136	NR1H4	48	0.002	1	33	0.002	0.76
137	PIK3CA	466	0.023	1	320	0.017	0.76
138	GPI	3807	0.188	1	2611	0.142	0.76
139	SLCO2A1	108	0.005	1	74	0.004	0.76
140	ATP5A1	6316	0.312	1	4327	0.235	0.76
141	FOXRED1	200	0.010	1	137	0.007	0.76
142	FBP1	19	0.001	1	13	0.001	0.75
143	CACYBP	133	0.007	1	91	0.005	0.75
144	XAB2	550	0.027	1	376	0.020	0.75
145	CCNT1	395	0.020	1	270	0.015	0.75
146	IRAK1	771	0.038	1	527	0.029	0.75
147	CD80	60	0.003	1	41	0.002	0.75
148	DENND4A	230	0.011	1	157	0.009	0.75
149	EIF4E	107	0.005	1	73	0.004	0.75
150	ITPKB	343	0.017	1	234	0.013	0.75
151	PER1	686	0.034	1	468	0.025	0.75
152	EN2	22	0.001	1	15	0.001	0.75
153	CXCR6	66	0.003	1	45	0.002	0.75
154	THEM4	198	0.010	1	135	0.007	0.75
155	PSMD7	1049	0.052	1	715	0.039	0.75
156	CASP8AP2	226	0.011	1	154	0.008	0.75
157	PORCN	47	0.002	1	32	0.002	0.75
158	PPP2CB	772	0.038	1	524	0.029	0.75

159	DNAJA4	28	0.001	1	19	0.001	0.75
160	TLR5	56	0.003	1	38	0.002	0.75
161	DNAJB1	578	0.029	1	392	0.021	0.75
162	VAPA	2858	0.141	1	1938	0.105	0.75
163	SOCS2	31	0.002	1	21	0.001	0.75
164	POLD2	638	0.031	1	432	0.024	0.75
165	MAOA	65	0.003	1	44	0.002	0.75
166	EI24	1742	0.086	1	1179	0.064	0.75
167	ZNF552	68	0.003	1	46	0.003	0.75
168	PRC1	241	0.012	1	163	0.009	0.75
169	GYS2	37	0.002	1	25	0.001	0.74
170	ALDOC	151	0.007	1	102	0.006	0.74
171	NFE2L2	40	0.002	1	27	0.001	0.74
172	PAK1IP1	163	0.008	1	110	0.006	0.74
173	MAGEB3	43	0.002	1	29	0.002	0.74
174	STK3	162	0.008	1	109	0.006	0.74
175	CNOT3	605	0.030	1	407	0.022	0.74
176	RPS6KA6	55	0.003	1	37	0.002	0.74
177	NPC1	116	0.006	1	78	0.004	0.74
178	MAPK14	265	0.013	1	178	0.010	0.74
179	FUBP1	2343	0.116	1	1573	0.086	0.74
180	PAICS	3065	0.151	1	2057	0.112	0.74
181	LPCAT1	76	0.004	1	51	0.003	0.74
182	HSP90B1	8492	0.419	1	5693	0.310	0.74
183	APP	5840	0.288	1	3912	0.213	0.74
184	KIF14	118	0.006	1	79	0.004	0.74
185	JUNB	1520	0.075	1	1017	0.055	0.74
186	MYCN	18	0.001	1	12	0.001	0.73
187	CX3CR1	33	0.002	1	22	0.001	0.73
188	LRMP	39	0.002	1	26	0.001	0.73
189	PLA2G5	45	0.002	1	30	0.002	0.73
190	MRE11A	51	0.003	1	34	0.002	0.73
191	DNAJB9	69	0.003	1	46	0.003	0.73
192	IRF9	81	0.004	1	54	0.003	0.73
193	FLRT3	144	0.007	1	96	0.005	0.73
194	NCK2	156	0.008	1	104	0.006	0.73
195	CD276	306	0.015	1	204	0.011	0.73
196	PDIA4	462	0.023	1	308	0.017	0.73
197	PAK7	3	0.000	1	2	0.000	0.73
198	WVOX	24	0.001	1	16	0.001	0.73
199	KL	27	0.001	1	18	0.001	0.73
200	IL23A	75	0.004	1	50	0.003	0.73
201	SLIT2	174	0.009	1	116	0.006	0.73
202	SNCA	249	0.012	1	166	0.009	0.73
203	HSPA9	1807	0.089	1	1204	0.066	0.73
204	KRT18	8025	0.396	1	5344	0.291	0.73
205	STAG2	670	0.033	1	446	0.024	0.73
206	CHD1	603	0.030	1	401	0.022	0.73
207	DNAJC11	337	0.017	1	224	0.012	0.73
208	GPAM	146	0.007	1	97	0.005	0.73
209	SRSF2	2771	0.137	1	1839	0.100	0.73
210	PHB	1672	0.083	1	1109	0.060	0.73
211	ATR	487	0.024	1	323	0.018	0.73
212	CCT7	2444	0.121	1	1620	0.088	0.73
213	HJURP	240	0.012	1	159	0.009	0.73
214	SERPINF1	74	0.004	1	49	0.003	0.73
215	PSMD2	876	0.043	1	580	0.032	0.73
216	POLE2	133	0.007	1	88	0.005	0.73
217	FAF1	443	0.022	1	293	0.016	0.73
218	STAT5A	56	0.003	1	37	0.002	0.73
219	GPC4	53	0.003	1	35	0.002	0.73
220	CALM1	471	0.023	1	311	0.017	0.73

221	SERTAD1	250	0.012	1	165	0.009	0.73
222	EAF2	91	0.004	1	60	0.003	0.73
223	WWC1	91	0.004	1	60	0.003	0.73
224	CLIC1	4551	0.225	1	3000	0.163	0.73
225	MAPK10	88	0.004	1	58	0.003	0.73
226	CRYAA	82	0.004	1	54	0.003	0.73
227	TAP1	158	0.008	1	104	0.006	0.73
228	MAGEA1	38	0.002	1	25	0.001	0.73
229	OPCML	38	0.002	1	25	0.001	0.73
230	TNKS2	225	0.011	1	148	0.008	0.73
231	LTN1	594	0.029	1	390	0.021	0.72
232	LPL	32	0.002	1	21	0.001	0.72
233	DNAJA3	850	0.042	1	557	0.030	0.72
234	GRP	58	0.003	1	38	0.002	0.72
235	TNFSF8	84	0.004	1	55	0.003	0.72
236	CCL23	55	0.003	1	36	0.002	0.72
237	CXCL12	52	0.003	1	34	0.002	0.72
238	SHC2	46	0.002	1	30	0.002	0.72
239	CALML3	69	0.003	1	45	0.002	0.72
240	GPSM2	634	0.031	1	413	0.022	0.72
241	PTCH1	109	0.005	1	71	0.004	0.72
242	BCL2L2	478	0.024	1	311	0.017	0.72
243	MAFF	552	0.027	1	359	0.020	0.72
244	CCNH	40	0.002	1	26	0.001	0.72
245	EGF	60	0.003	1	39	0.002	0.72
246	MSI1	120	0.006	1	78	0.004	0.72
247	CXCL16	180	0.009	1	117	0.006	0.72
248	MYCL	37	0.002	1	24	0.001	0.72
249	TNXB	74	0.004	1	48	0.003	0.72
250	CIDEB	54	0.003	1	35	0.002	0.71
251	SORT1	695	0.034	1	450	0.024	0.71
252	RPS4X	3881	0.192	1	2511	0.137	0.71
253	ADM	1281	0.063	1	828	0.045	0.71
254	E2F1	82	0.004	1	53	0.003	0.71
255	ATP6V0C	814	0.040	1	526	0.029	0.71
256	HMGB2	1119	0.055	1	723	0.039	0.71
257	SYNCRIP	1355	0.067	1	875	0.048	0.71
258	CRYAB	79	0.004	1	51	0.003	0.71
259	ATG3	976	0.048	1	630	0.034	0.71
260	AXIN1	138	0.007	1	89	0.005	0.71
261	ADIPOR1	2580	0.127	1	1663	0.091	0.71
262	TP53BP2	244	0.012	1	157	0.009	0.71
263	SEC22C	157	0.008	1	101	0.005	0.71
264	PDGFRA	56	0.003	1	36	0.002	0.71
265	NOD2	42	0.002	1	27	0.001	0.71
266	SRM	1150	0.057	1	739	0.040	0.71
267	MAP3K4	494	0.024	1	317	0.017	0.71
268	KRAS	533	0.026	1	342	0.019	0.71
269	PYCARD	92	0.005	1	59	0.003	0.71
270	ASPM	1201	0.059	1	769	0.042	0.71
271	CSF2	25	0.001	1	16	0.001	0.71
272	GJA1	75	0.004	1	48	0.003	0.71
273	SMC3	789	0.039	1	504	0.027	0.70
274	CCL21	83	0.004	1	53	0.003	0.70
275	CBX5	1687	0.083	1	1077	0.059	0.70
276	FGF3	47	0.002	1	30	0.002	0.70
277	GTF2H3	290	0.014	1	185	0.010	0.70
278	HSPA4	2941	0.145	1	1875	0.102	0.70
279	GSK3B	1354	0.067	1	863	0.047	0.70
280	SACS	583	0.029	1	371	0.020	0.70
281	RIPK1	225	0.011	1	143	0.008	0.70
282	XRCC1	85	0.004	1	54	0.003	0.70

283	RASD1	52	0.003	1	33	0.002	0.70
284	FBXW11	1088	0.054	1	689	0.037	0.70
285	CXCL6	79	0.004	1	50	0.003	0.70
286	FANCE	117	0.006	1	74	0.004	0.70
287	TNFRSF10A	185	0.009	1	117	0.006	0.70
288	ZHX2	141	0.007	1	89	0.005	0.70
289	FZD5	65	0.003	1	41	0.002	0.70
290	PARP2	241	0.012	1	152	0.008	0.70
291	ETV5	268	0.013	1	169	0.009	0.70
292	NOLC1	522	0.026	1	329	0.018	0.69
293	DKC1	868	0.043	1	547	0.030	0.69
294	MTMR2	762	0.038	1	480	0.026	0.69
295	CUL2	578	0.029	1	364	0.020	0.69
296	NOD1	54	0.003	1	34	0.002	0.69
297	IRAK2	97	0.005	1	61	0.003	0.69
298	EIF5B	288	0.014	1	181	0.010	0.69
299	FOXP1	304	0.015	1	191	0.010	0.69
300	FLRT2	191	0.009	1	120	0.007	0.69
301	DUSP5	320	0.016	1	201	0.011	0.69
302	PPP2R4	486	0.024	1	305	0.017	0.69
303	ATP7B	51	0.003	1	32	0.002	0.69
304	FGF14	51	0.003	1	32	0.002	0.69
305	CAV2	2812	0.139	1	1764	0.096	0.69
306	TOP3A	142	0.007	1	89	0.005	0.69
307	S100A2	163	0.008	1	102	0.006	0.69
308	CD55	1228	0.061	1	768	0.042	0.69
309	MAG	8	0.000	1	5	0.000	0.69
310	CAMKK2	72	0.004	1	45	0.002	0.69
311	RELB	56	0.003	1	35	0.002	0.69
312	JUN	671	0.033	1	419	0.023	0.69
313	SERPINH1	1864	0.092	1	1163	0.063	0.69
314	CHEK2	170	0.008	1	106	0.006	0.69
315	TNFRSF10B	1561	0.077	1	973	0.053	0.69
316	RPL13	138	0.007	1	86	0.005	0.69
317	PRKAG3	53	0.003	1	33	0.002	0.69
318	CD209	151	0.007	1	94	0.005	0.69
319	CRLF2	45	0.002	1	28	0.002	0.69
320	CCL19	37	0.002	1	23	0.001	0.69
321	NUF2	459	0.023	1	285	0.016	0.68
322	PLA2G4C	58	0.003	1	36	0.002	0.68
323	PIM3	303	0.015	1	188	0.010	0.68
324	ECT2	740	0.037	1	459	0.025	0.68
325	GYS1	158	0.008	1	98	0.005	0.68
326	DNAJB11	752	0.037	1	466	0.025	0.68
327	NUPR1	744	0.037	1	461	0.025	0.68
328	HSPD1	6857	0.339	1	4248	0.231	0.68
329	KRT13	76	0.004	1	47	0.003	0.68
330	AK3	186	0.009	1	115	0.006	0.68
331	CENPF	1733	0.086	1	1071	0.058	0.68
332	ADIPOR2	709	0.035	1	438	0.024	0.68
333	FGF1	34	0.002	1	21	0.001	0.68
334	SLC16A1	1543	0.076	1	953	0.052	0.68
335	FZD10	47	0.002	1	29	0.002	0.68
336	PRKAA1	394	0.019	1	243	0.013	0.68
337	BAK1	621	0.031	1	383	0.021	0.68
338	NME1	1803	0.089	1	1111	0.060	0.68
339	HRAS	614	0.030	1	378	0.021	0.68
340	BMP1	554	0.027	1	341	0.019	0.68
341	RASSF1	26	0.001	1	16	0.001	0.68
342	HSPB3	39	0.002	1	24	0.001	0.68
343	PALM2_AKAP2	65	0.003	1	40	0.002	0.68
344	GNG4	122	0.006	1	75	0.004	0.68

345	CRY1	96	0.005	1	59	0.003	0.68
346	RUNDC1	272	0.013	1	167	0.009	0.68
347	ACKR4	44	0.002	1	27	0.001	0.68
348	DNAJC5G	44	0.002	1	27	0.001	0.68
349	YWHAZ	15370	0.759	1	9414	0.512	0.68
350	EVL	420	0.021	1	257	0.014	0.67
351	XIAP	170	0.008	1	104	0.006	0.67
352	BTG3	703	0.035	1	430	0.023	0.67
353	MAPKAPK5	36	0.002	1	22	0.001	0.67
354	FGF7	77	0.004	1	47	0.003	0.67
355	SPC25	136	0.007	1	83	0.005	0.67
356	TXNL4B	141	0.007	1	86	0.005	0.67
357	BCL2L10	123	0.006	1	75	0.004	0.67
358	NCOR1	361	0.018	1	220	0.012	0.67
359	ACKR2	64	0.003	1	39	0.002	0.67
360	NFKB1	471	0.023	1	287	0.016	0.67
361	GUCY1A3	69	0.003	1	42	0.002	0.67
362	C17orf53	194	0.010	1	118	0.006	0.67
363	SHC4	74	0.004	1	45	0.002	0.67
364	RIF1	630	0.031	1	383	0.021	0.67
365	TMEM123	51	0.003	1	31	0.002	0.67
366	ZFP42	51	0.003	1	31	0.002	0.67
367	NEIL2	79	0.004	1	48	0.003	0.67
368	MKI67	981	0.048	1	596	0.032	0.67
369	GADD45A	1908	0.094	1	1159	0.063	0.67
370	FAM117B	191	0.009	1	116	0.006	0.67
371	BOD1	84	0.004	1	51	0.003	0.67
372	DUSP8	117	0.006	1	71	0.004	0.67
373	CBX3	4206	0.208	1	2551	0.139	0.67
374	CPT1B	269	0.013	1	163	0.009	0.67
375	IDH1	2504	0.124	1	1516	0.083	0.67
376	GSTP1	10353	0.511	1	6267	0.341	0.67
377	CDC14A	119	0.006	1	72	0.004	0.67
378	KPNA2	2244	0.111	1	1357	0.074	0.67
379	MAGEE1	43	0.002	1	26	0.001	0.67
380	SMC1A	1084	0.054	1	655	0.036	0.67
381	JAG2	106	0.005	1	64	0.003	0.67
382	HHEX	121	0.006	1	73	0.004	0.67
383	RRM2	1951	0.096	1	1177	0.064	0.67
384	SNRPB	2786	0.138	1	1680	0.091	0.66
385	PRKAR2B	73	0.004	1	44	0.002	0.66
386	DNAJC17	479	0.024	1	288	0.016	0.66
387	MBL2	15	0.001	1	9	0.000	0.66
388	TERT	35	0.002	1	21	0.001	0.66
389	TNFSF15	40	0.002	1	24	0.001	0.66
390	HSPH1	45	0.002	1	27	0.001	0.66
391	ALDH4A1	50	0.002	1	30	0.002	0.66
392	TNFSF13B	60	0.003	1	36	0.002	0.66
393	CCT2	1535	0.076	1	920	0.050	0.66
394	USF2	112	0.006	1	67	0.004	0.66
395	BID	393	0.019	1	235	0.013	0.66
396	SLC25A13	393	0.019	1	235	0.013	0.66
397	CTPS1	455	0.022	1	272	0.015	0.66
398	MCM6	595	0.029	1	355	0.019	0.66
399	GNAQ	585	0.029	1	349	0.019	0.66
400	CTSS	99	0.005	1	59	0.003	0.66
401	TNKS	47	0.002	1	28	0.002	0.66
402	TAZ	185	0.009	1	110	0.006	0.66
403	PROK2	37	0.002	1	22	0.001	0.66
404	ATP5G1	1383	0.068	1	822	0.045	0.66
405	IHH	32	0.002	1	19	0.001	0.65
406	KDM5B	987	0.049	1	586	0.032	0.65

407	HK2	155	0.008	1	92	0.005	0.65
408	GNL3	1813	0.090	1	1076	0.059	0.65
409	PDGFA	258	0.013	1	153	0.008	0.65
410	RRM1	1153	0.057	1	683	0.037	0.65
411	AKAP1	884	0.044	1	523	0.028	0.65
412	AR	44	0.002	1	26	0.001	0.65
413	TLR7	44	0.002	1	26	0.001	0.65
414	XRCC5	83	0.004	1	49	0.003	0.65
415	RUNX1	432	0.021	1	255	0.014	0.65
416	CASP9	39	0.002	1	23	0.001	0.65
417	PCNA	2358	0.116	1	1390	0.076	0.65
418	FGF12	56	0.003	1	33	0.002	0.65
419	RASA1	503	0.025	1	296	0.016	0.65
420	HUS1	204	0.010	1	120	0.007	0.65
421	CCR8	34	0.002	1	20	0.001	0.65
422	VIPR1	34	0.002	1	20	0.001	0.65
423	SLC7A9	68	0.003	1	40	0.002	0.65
424	ARMC1	631	0.031	1	371	0.020	0.65
425	MDC1	150	0.007	1	88	0.005	0.65
426	TYMS	1982	0.098	1	1162	0.063	0.65
427	MYBL2	449	0.022	1	263	0.014	0.65
428	FGF5	41	0.002	1	24	0.001	0.65
429	COL5A1	82	0.004	1	48	0.003	0.65
430	FEM1B	598	0.030	1	350	0.019	0.65
431	IL15	118	0.006	1	69	0.004	0.64
432	CASP10	77	0.004	1	45	0.002	0.64
433	MAP7D1	125	0.006	1	73	0.004	0.64
434	DNAJC16	72	0.004	1	42	0.002	0.64
435	BMP2	12	0.001	1	7	0.000	0.64
436	LTB	24	0.001	1	14	0.001	0.64
437	MT1X	1247	0.062	1	727	0.040	0.64
438	AGER	115	0.006	1	67	0.004	0.64
439	TUBB3	184	0.009	1	107	0.006	0.64
440	SLC19A3	43	0.002	1	25	0.001	0.64
441	DHCR24	2919	0.144	1	1697	0.092	0.64
442	ALPL	93	0.005	1	54	0.003	0.64
443	CELSR2	100	0.005	1	58	0.003	0.64
444	PPP3CB	383	0.019	1	222	0.012	0.64
445	TPD52	1077	0.053	1	624	0.034	0.64
446	SPHK1	95	0.005	1	55	0.003	0.64
447	TMSB10	59206	2.923	1	34272	1.865	0.64
448	GREM1	64	0.003	1	37	0.002	0.64
449	SUV39H2	362	0.018	1	209	0.011	0.64
450	NUMB	291	0.014	1	168	0.009	0.64
451	PLA2G4A	279	0.014	1	161	0.009	0.64
452	EOMES	59	0.003	1	34	0.002	0.64
453	HMGXB3	177	0.009	1	102	0.006	0.64
454	TRIP13	750	0.037	1	432	0.024	0.63
455	FRZB	33	0.002	1	19	0.001	0.63
456	BAG1	233	0.012	1	134	0.007	0.63
457	ZNF589	127	0.006	1	73	0.004	0.63
458	ITGAV	674	0.033	1	387	0.021	0.63
459	OLR1	54	0.003	1	31	0.002	0.63
460	RIPK2	218	0.011	1	125	0.007	0.63
461	SLC38A5	82	0.004	1	47	0.003	0.63
462	KCNIP1	89	0.004	1	51	0.003	0.63
463	PLEK2	124	0.006	1	71	0.004	0.63
464	CCR10	49	0.002	1	28	0.002	0.63
465	CDH5	56	0.003	1	32	0.002	0.63
466	IL4	63	0.003	1	36	0.002	0.63
467	CCNE1	189	0.009	1	108	0.006	0.63
468	ALDH2	21	0.001	1	12	0.001	0.63

469	MMP7	42	0.002	1	24	0.001	0.63
470	TNNI3	42	0.002	1	24	0.001	0.63
471	MOS	84	0.004	1	48	0.003	0.63
472	HSF2	175	0.009	1	100	0.005	0.63
473	MELK	562	0.028	1	321	0.017	0.63
474	SORD	810	0.040	1	462	0.025	0.63
475	ANXA1	6621	0.327	1	3771	0.205	0.63
476	PLCG2	58	0.003	1	33	0.002	0.63
477	MAP3K3	109	0.005	1	62	0.003	0.63
478	PTPN11	814	0.040	1	463	0.025	0.63
479	DNAJC9	405	0.020	1	230	0.013	0.63
480	TINF2	127	0.006	1	72	0.004	0.62
481	SMS	2142	0.106	1	1213	0.066	0.62
482	E2F5	152	0.008	1	86	0.005	0.62
483	VDAC1	8163	0.403	1	4618	0.251	0.62
484	BUB1	1117	0.055	1	631	0.034	0.62
485	TRAF6	232	0.011	1	131	0.007	0.62
486	SUSD3	62	0.003	1	35	0.002	0.62
487	CXCL5	39	0.002	1	22	0.001	0.62
488	DUSP2	39	0.002	1	22	0.001	0.62
489	SLC31A1	314	0.016	1	177	0.010	0.62
490	TPX2	641	0.032	1	361	0.020	0.62
491	CDCA7	112	0.006	1	63	0.003	0.62
492	RAD51	217	0.011	1	122	0.007	0.62
493	CKS2	2946	0.145	1	1655	0.090	0.62
494	PLK1	1053	0.052	1	591	0.032	0.62
495	GMPS	1290	0.064	1	724	0.039	0.62
496	CXCL14	98	0.005	1	55	0.003	0.62
497	RUVL1	1645	0.081	1	923	0.050	0.62
498	PLK4	423	0.021	1	237	0.013	0.62
499	YBX3	981	0.048	1	549	0.030	0.62
500	MYC	1496	0.074	1	837	0.046	0.62
501	ERO1L	717	0.035	1	401	0.022	0.62
502	DBI	6117	0.302	1	3421	0.186	0.62
503	MAPK8	93	0.005	1	52	0.003	0.62
504	CKS1B	1720	0.085	1	960	0.052	0.62
505	TLR8	52	0.003	1	29	0.002	0.61
506	PDK1	296	0.015	1	165	0.009	0.61
507	CD22	79	0.004	1	44	0.002	0.61
508	SKP2	670	0.033	1	373	0.020	0.61
509	MAP2K3	291	0.014	1	162	0.009	0.61
510	NASP	812	0.040	1	452	0.025	0.61
511	DNMT1	922	0.046	1	513	0.028	0.61
512	PLCG1	283	0.014	1	157	0.009	0.61
513	PAK1	193	0.010	1	107	0.006	0.61
514	CENPN	74	0.004	1	41	0.002	0.61
515	TSHR	65	0.003	1	36	0.002	0.61
516	PSRC1	318	0.016	1	176	0.010	0.61
517	NOX1	47	0.002	1	26	0.001	0.61
518	NCL	17179	0.848	1	9503	0.517	0.61
519	CCNB1	2627	0.130	1	1453	0.079	0.61
520	PTTG2	2554	0.126	1	1412	0.077	0.61
521	FBXW7	123	0.006	1	68	0.004	0.61
522	MAPT	29	0.001	1	16	0.001	0.61
523	MNAT1	167	0.008	1	92	0.005	0.61
524	EFNB1	109	0.005	1	60	0.003	0.61
525	COL11A2	40	0.002	1	22	0.001	0.61
526	JPH3	82	0.004	1	45	0.002	0.60
527	TIFA	359	0.018	1	197	0.011	0.60
528	BCL10	966	0.048	1	530	0.029	0.60
529	MAP3K2	301	0.015	1	165	0.009	0.60
530	RAB27B	301	0.015	1	165	0.009	0.60

531	HSPA2	73	0.004	1	40	0.002	0.60
532	MSH6	482	0.024	1	264	0.014	0.60
533	CTNND2	42	0.002	1	23	0.001	0.60
534	CHSY1	170	0.008	1	93	0.005	0.60
535	DLGAP5	850	0.042	1	465	0.025	0.60
536	RAG1	64	0.003	1	35	0.002	0.60
537	XBP1	715	0.035	1	391	0.021	0.60
538	KRT16	236	0.012	1	129	0.007	0.60
539	CCT4	2101	0.104	1	1147	0.062	0.60
540	HHIP	44	0.002	1	24	0.001	0.60
541	BBC3	90	0.004	1	49	0.003	0.60
542	TTK	421	0.021	1	229	0.012	0.60
543	GBP2	114	0.006	1	62	0.003	0.60
544	ADRA1A	92	0.005	1	50	0.003	0.60
545	CCNO	116	0.006	1	63	0.003	0.60
546	PRKCZ	70	0.003	1	38	0.002	0.60
547	TAP2	144	0.007	1	78	0.004	0.60
548	NAT8L	24	0.001	1	13	0.001	0.60
549	RELA	48	0.002	1	26	0.001	0.60
550	WNT16	48	0.002	1	26	0.001	0.60
551	HMGA1	2257	0.111	1	1222	0.067	0.60
552	FUT8	352	0.017	1	190	0.010	0.60
553	WNT8A	63	0.003	1	34	0.002	0.59
554	NOTCH1	126	0.006	1	68	0.004	0.59
555	CACNA1G	39	0.002	1	21	0.001	0.59
556	PTCHD1	39	0.002	1	21	0.001	0.59
557	HNF1A	52	0.003	1	28	0.002	0.59
558	TNFRSF9	52	0.003	1	28	0.002	0.59
559	WNT9A	225	0.011	1	121	0.007	0.59
560	CCL2	80	0.004	1	43	0.002	0.59
561	MFNG	41	0.002	1	22	0.001	0.59
562	TGFBR1	427	0.021	1	229	0.012	0.59
563	MCM5	735	0.036	1	394	0.021	0.59
564	AKR1B1	6825	0.337	1	3658	0.199	0.59
565	MUC1	1394	0.069	1	747	0.041	0.59
566	DNAJC5B	43	0.002	1	23	0.001	0.59
567	RASGRF2	60	0.003	1	32	0.002	0.59
568	PHGDH	818	0.040	1	436	0.024	0.59
569	DPPA4	154	0.008	1	82	0.004	0.59
570	TMEM57	216	0.011	1	115	0.006	0.59
571	SIAH1	299	0.015	1	159	0.009	0.59
572	IRF1	79	0.004	1	42	0.002	0.59
573	TNFRSF17	49	0.002	1	26	0.001	0.58
574	FOXC1	17	0.001	1	9	0.000	0.58
575	HOXA10	17	0.001	1	9	0.000	0.58
576	POLD4	51	0.003	1	27	0.001	0.58
577	SPI1	68	0.003	1	36	0.002	0.58
578	NLRP3	85	0.004	1	45	0.002	0.58
579	TNFAIP2	85	0.004	1	45	0.002	0.58
580	ACLY	2291	0.113	1	1212	0.066	0.58
581	MCM2	465	0.023	1	245	0.013	0.58
582	MMS19	380	0.019	1	200	0.011	0.58
583	LIG1	333	0.016	1	175	0.010	0.58
584	PMAIP1	198	0.010	1	104	0.006	0.58
585	GPR160	40	0.002	1	21	0.001	0.58
586	CNPY3	128	0.006	1	67	0.004	0.58
587	CCT5	1811	0.089	1	947	0.052	0.58
588	CYP1A2	44	0.002	1	23	0.001	0.58
589	FOXD3	23	0.001	1	12	0.001	0.58
590	PLA2G4E	69	0.003	1	36	0.002	0.58
591	CDC20	2422	0.120	1	1262	0.069	0.57
592	PRLR	48	0.002	1	25	0.001	0.57

593	RORC	73	0.004	1	38	0.002	0.57
594	PTP4A1	892	0.044	1	464	0.025	0.57
595	TMPRSS2	25	0.001	1	13	0.001	0.57
596	MMP14	777	0.038	1	404	0.022	0.57
597	ABCA4	77	0.004	1	40	0.002	0.57
598	SLC45A3	231	0.011	1	120	0.007	0.57
599	BTG2	617	0.030	1	320	0.017	0.57
600	TIAM1	108	0.005	1	56	0.003	0.57
601	IL7R	195	0.010	1	101	0.005	0.57
602	MADCAM1	56	0.003	1	29	0.002	0.57
603	TNFRSF25	114	0.006	1	59	0.003	0.57
604	EPGN	892	0.044	1	461	0.025	0.57
605	KLRG1	60	0.003	1	31	0.002	0.57
606	CCNA2	1026	0.051	1	530	0.029	0.57
607	MAD2L1	775	0.038	1	400	0.022	0.57
608	SMO	64	0.003	1	33	0.002	0.57
609	BIRC7	97	0.005	1	50	0.003	0.57
610	DDIT3	33	0.002	1	17	0.001	0.57
611	GNAZ	66	0.003	1	34	0.002	0.57
612	RFC4	439	0.022	1	226	0.012	0.57
613	BUB1B	212	0.010	1	109	0.006	0.57
614	TYMP	395	0.020	1	203	0.011	0.57
615	MRPL13	477	0.024	1	245	0.013	0.57
616	IGF2	37	0.002	1	19	0.001	0.57
617	FPR1	417	0.021	1	214	0.012	0.57
618	LIG3	154	0.008	1	79	0.004	0.57
619	DICER1	525	0.026	1	269	0.015	0.56
620	PA2G4	3074	0.152	1	1575	0.086	0.56
621	CHD7	256	0.013	1	131	0.007	0.56
622	MAPK13	86	0.004	1	44	0.002	0.56
623	RFC3	507	0.025	1	259	0.014	0.56
624	FANCB	186	0.009	1	95	0.005	0.56
625	PPARGC1B	237	0.012	1	121	0.007	0.56
626	HNRNPA2B1	2706	0.134	1	1379	0.075	0.56
627	GEMIN2	263	0.013	1	134	0.007	0.56
628	AURKA	1727	0.085	1	879	0.048	0.56
629	CD86	63	0.003	1	32	0.002	0.56
630	PPP2R2C	203	0.010	1	103	0.006	0.56
631	C3	2949	0.146	1	1494	0.081	0.56
632	ITLN2	79	0.004	1	40	0.002	0.56
633	MDM2	2824	0.139	1	1429	0.078	0.56
634	SLK	832	0.041	1	421	0.023	0.56
635	MAP2K4	319	0.016	1	161	0.009	0.56
636	CMC2	465	0.023	1	234	0.013	0.55
637	REL	211	0.010	1	106	0.006	0.55
638	GTSE1	237	0.012	1	119	0.006	0.55
639	SLC3A2	1103	0.054	1	553	0.030	0.55
640	HIST1H3H	3524	0.174	1	1764	0.096	0.55
641	HGF	8	0.000	1	4	0.000	0.55
642	COMP	26	0.001	1	13	0.001	0.55
643	TBX21	30	0.001	1	15	0.001	0.55
644	MAGEB2	42	0.002	1	21	0.001	0.55
645	PRKAG2	66	0.003	1	33	0.002	0.55
646	PTCHD2	72	0.004	1	36	0.002	0.55
647	NGFR	86	0.004	1	43	0.002	0.55
648	BCL2	94	0.005	1	47	0.003	0.55
649	BRCA2	202	0.010	1	101	0.005	0.55
650	CAPN1	586	0.029	1	293	0.016	0.55
651	XRCC6BP1	24	0.001	1	12	0.001	0.55
652	ATG13	50	0.002	1	25	0.001	0.55
653	BMP8A	74	0.004	1	37	0.002	0.55
654	IGF1R	200	0.010	1	100	0.005	0.55

655	RPS6KA1	417	0.021	1	208	0.011	0.55
656	CASP1	524	0.026	1	261	0.014	0.55
657	CTSC	2319	0.114	1	1154	0.063	0.55
658	ZAK	561	0.028	1	278	0.015	0.55
659	DFFA	271	0.013	1	134	0.007	0.55
660	DAB2IP	393	0.019	1	194	0.011	0.54
661	LEF1	67	0.003	1	33	0.002	0.54
662	LIPE	61	0.003	1	30	0.002	0.54
663	GRIA3	57	0.003	1	28	0.002	0.54
664	KCTD11	55	0.003	1	27	0.001	0.54
665	MAPRE2	108	0.005	1	53	0.003	0.54
666	LRG1	53	0.003	1	26	0.001	0.54
667	CACNG1	51	0.003	1	25	0.001	0.54
668	KDM5C	737	0.036	1	361	0.020	0.54
669	SNAI3	45	0.002	1	22	0.001	0.54
670	HSPE1	7143	0.353	1	3492	0.190	0.54
671	SMPDL3B	119	0.006	1	58	0.003	0.54
672	TCP1	3441	0.170	1	1675	0.091	0.54
673	ADAM17	405	0.020	1	197	0.011	0.54
674	MAP2K1	999	0.049	1	485	0.026	0.54
675	CDX2	64	0.003	1	31	0.002	0.53
676	DLL3	60	0.003	1	29	0.002	0.53
677	LMO2	60	0.003	1	29	0.002	0.53
678	DNAJB13	58	0.003	1	28	0.002	0.53
679	NKX3 1	114	0.006	1	55	0.003	0.53
680	NOS3	27	0.001	1	13	0.001	0.53
681	PABPC1	81	0.004	1	39	0.002	0.53
682	CDC7	192	0.009	1	92	0.005	0.53
683	TNFRSF14	46	0.002	1	22	0.001	0.53
684	PROM1	67	0.003	1	32	0.002	0.53
685	NT5E	289	0.014	1	138	0.008	0.53
686	TNFRSF21	267	0.013	1	127	0.007	0.52
687	CCR3	61	0.003	1	29	0.002	0.52
688	ANAPC2	179	0.009	1	85	0.005	0.52
689	CEACAM7	59	0.003	1	28	0.002	0.52
690	BLM	133	0.007	1	63	0.003	0.52
691	ORC6	169	0.008	1	80	0.004	0.52
692	BAG3	656	0.032	1	310	0.017	0.52
693	TRIP4	91	0.004	1	43	0.002	0.52
694	HSPA4L	377	0.019	1	178	0.010	0.52
695	GM2A	303	0.015	1	143	0.008	0.52
696	NKD1	34	0.002	1	16	0.001	0.52
697	FAM162A	253	0.012	1	119	0.006	0.52
698	FGFR2	47	0.002	1	22	0.001	0.52
699	TDG	691	0.034	1	323	0.018	0.52
700	FOS	2531	0.125	1	1181	0.064	0.51
701	MCM3	533	0.026	1	248	0.013	0.51
702	PPID	570	0.028	1	265	0.014	0.51
703	EZR	994	0.049	1	462	0.025	0.51
704	VAMP8	1070	0.053	1	494	0.027	0.51
705	TIE1	52	0.003	1	24	0.001	0.51
706	ORM1	91	0.004	1	42	0.002	0.51
707	MCL1	4472	0.221	1	2064	0.112	0.51
708	NRAS	404	0.020	1	186	0.010	0.51
709	ANXA8	1838	0.091	1	846	0.046	0.51
710	SREBF1	1489	0.074	1	685	0.037	0.51
711	ACTR3B	50	0.002	1	23	0.001	0.51
712	BOC	37	0.002	1	17	0.001	0.51
713	THPO	85	0.004	1	39	0.002	0.51
714	NODAL	83	0.004	1	38	0.002	0.50
715	TSPAN14	996	0.049	1	456	0.025	0.50
716	NR4A1	142	0.007	1	65	0.004	0.50

717	LYZ	59	0.003	1	27	0.001	0.50
718	MLLT4	306	0.015	1	140	0.008	0.50
719	RALA	460	0.023	1	210	0.011	0.50
720	CASP14	57	0.003	1	26	0.001	0.50
721	LAG3	112	0.006	1	51	0.003	0.50
722	ITGB7	66	0.003	1	30	0.002	0.50
723	ETV4	55	0.003	1	25	0.001	0.50
724	PFKFB2	405	0.020	1	183	0.010	0.50
725	CCL28	186	0.009	1	84	0.005	0.50
726	CDC25A	182	0.009	1	82	0.004	0.50
727	RAC3	188	0.009	1	84	0.005	0.49
728	AQP1	18	0.001	1	8	0.000	0.49
729	MUTYH	54	0.003	1	24	0.001	0.49
730	PKMYT1	302	0.015	1	134	0.007	0.49
731	MTFP1	230	0.011	1	102	0.006	0.49
732	EGFR	954	0.047	1	423	0.023	0.49
733	SLC2A1	1039	0.051	1	460	0.025	0.49
734	SLC7A5	615	0.030	1	271	0.015	0.49
735	PF4V1	84	0.004	1	37	0.002	0.49
736	CEP55	50	0.002	1	22	0.001	0.49
737	TAGLN	1082	0.053	1	476	0.026	0.48
738	IL2RA	41	0.002	1	18	0.001	0.48
739	ACACA	647	0.032	1	284	0.015	0.48
740	STAT6	498	0.025	1	218	0.012	0.48
741	VCAM1	64	0.003	1	28	0.002	0.48
742	SLCO3A1	96	0.005	1	42	0.002	0.48
743	AURKB	605	0.030	1	264	0.014	0.48
744	TNFSF14	94	0.005	1	41	0.002	0.48
745	RAD9A	101	0.005	1	44	0.002	0.48
746	ESR1	74	0.004	1	32	0.002	0.48
747	XRCC4	74	0.004	1	32	0.002	0.48
748	HOXD1	58	0.003	1	25	0.001	0.48
749	GABBR2	72	0.004	1	31	0.002	0.47
750	TNC	144	0.007	1	62	0.003	0.47
751	ANGPT2	7	0.000	1	3	0.000	0.47
752	RIPK3	7	0.000	1	3	0.000	0.47
753	TREX1	49	0.002	1	21	0.001	0.47
754	MYOD1	42	0.002	1	18	0.001	0.47
755	DSC2	173	0.009	1	74	0.004	0.47
756	PELP1	131	0.006	1	56	0.003	0.47
757	CSNK2A1	797	0.039	1	340	0.019	0.47
758	RPS6KB2	455	0.022	1	194	0.011	0.47
759	PFKFB3	176	0.009	1	75	0.004	0.47
760	SPEF1	47	0.002	1	20	0.001	0.47
761	XCR1	47	0.002	1	20	0.001	0.47
762	TCF7L1	26	0.001	1	11	0.001	0.47
763	DDX58	346	0.017	1	146	0.008	0.47
764	NDRG1	524	0.026	1	221	0.012	0.46
765	EPO	57	0.003	1	24	0.001	0.46
766	PFDN2	3443	0.170	1	1449	0.079	0.46
767	CCR6	50	0.002	1	21	0.001	0.46
768	PRKX	360	0.018	1	151	0.008	0.46
769	SLCO4A1	31	0.002	1	13	0.001	0.46
770	HES1	117	0.006	1	49	0.003	0.46
771	SLC7A8	79	0.004	1	33	0.002	0.46
772	CCNB2	84	0.004	1	35	0.002	0.46
773	RAD51C	192	0.009	1	80	0.004	0.46
774	FST	118	0.006	1	49	0.003	0.46
775	CCL5	94	0.005	1	39	0.002	0.46
776	TLR2	82	0.004	1	34	0.002	0.46
777	TRAF2	70	0.003	1	29	0.002	0.46
778	CD70	298	0.015	1	123	0.007	0.46

779	STEAP4	80	0.004	1	33	0.002	0.45
780	CCT6A	3304	0.163	1	1361	0.074	0.45
781	ESM1	17	0.001	1	7	0.000	0.45
782	HLA_B	1665	0.082	1	685	0.037	0.45
783	ADORA2B	825	0.041	1	339	0.018	0.45
784	GPD2	677	0.033	1	277	0.015	0.45
785	KSR1	27	0.001	1	11	0.001	0.45
786	CCL22	59	0.003	1	24	0.001	0.45
787	HSPB7	32	0.002	1	13	0.001	0.45
788	NR4A3	47	0.002	1	19	0.001	0.45
789	SYK	124	0.006	1	50	0.003	0.44
790	CDK8	362	0.018	1	145	0.008	0.44
791	MAPK8IP2	5	0.000	1	2	0.000	0.44
792	LECT1	40	0.002	1	16	0.001	0.44
793	CHGA	60	0.003	1	24	0.001	0.44
794	SOCS3	65	0.003	1	26	0.001	0.44
795	XRCC2	501	0.025	1	200	0.011	0.44
796	HMGCR	540	0.027	1	215	0.012	0.44
797	CMTM2	73	0.004	1	29	0.002	0.44
798	CAPN6	43	0.002	1	17	0.001	0.44
799	PRKCB	43	0.002	1	17	0.001	0.44
800	FANCA	86	0.004	1	34	0.002	0.44
801	CCL3	81	0.004	1	32	0.002	0.44
802	ABCA3	119	0.006	1	47	0.003	0.44
803	SLC2A3	193	0.010	1	76	0.004	0.43
804	EIF4EBP1	1104	0.055	1	434	0.024	0.43
805	PTCHD3	56	0.003	1	22	0.001	0.43
806	EIF4A1	13513	0.667	1	5285	0.288	0.43
807	HSPA14	77	0.004	1	30	0.002	0.43
808	CCNA1	95	0.005	1	37	0.002	0.43
809	DKK2	67	0.003	1	26	0.001	0.43
810	WASL	49	0.002	1	19	0.001	0.43
811	CD8A	80	0.004	1	31	0.002	0.43
812	NMU	411	0.020	1	159	0.009	0.43
813	VCAN	539	0.027	1	208	0.011	0.43
814	FHIT	127	0.006	1	49	0.003	0.43
815	IL10RA	70	0.003	1	27	0.001	0.43
816	ICAM1	172	0.008	1	66	0.004	0.42
817	LTA	34	0.002	1	13	0.001	0.42
818	PAX8	68	0.003	1	26	0.001	0.42
819	DEFB1	97	0.005	1	37	0.002	0.42
820	EBF4	21	0.001	1	8	0.000	0.42
821	PTPN5	42	0.002	1	16	0.001	0.42
822	ANGPTL4	651	0.032	1	248	0.013	0.42
823	RBBP8	541	0.027	1	206	0.011	0.42
824	PIDD1	305	0.015	1	116	0.006	0.42
825	CDC6	500	0.025	1	190	0.010	0.42
826	EDN1	129	0.006	1	49	0.003	0.42
827	PCSK6	140	0.007	1	53	0.003	0.42
828	SMUG1	107	0.005	1	40	0.002	0.41
829	CXCR2	27	0.001	1	10	0.001	0.41
830	TLR9	46	0.002	1	17	0.001	0.41
831	SLC29A1	112	0.006	1	41	0.002	0.40
832	SLC29A2	389	0.019	1	142	0.008	0.40
833	CDC25B	1251	0.062	1	456	0.025	0.40
834	MT2A	45572	2.250	1	16510	0.899	0.40
835	FOXE1	58	0.003	1	21	0.001	0.40
836	ATXN1	61	0.003	1	22	0.001	0.40
837	TIMP3	272	0.013	1	98	0.005	0.40
838	BBS4	25	0.001	1	9	0.000	0.40
839	DUSP6	304	0.015	1	109	0.006	0.40
840	UBE2C	112	0.006	1	40	0.002	0.39

841	EXO1	266	0.013	1	95	0.005	0.39
842	ETV7	79	0.004	1	28	0.002	0.39
843	NFKB2	173	0.009	1	61	0.003	0.39
844	KIF2C	323	0.016	1	112	0.006	0.38
845	SERPINB5	1068	0.053	1	370	0.020	0.38
846	CD83	137	0.007	1	47	0.003	0.38
847	VEGFC	248	0.012	1	85	0.005	0.38
848	MTSS1	187	0.009	1	64	0.003	0.38
849	PLAUR	702	0.035	1	235	0.013	0.37
850	DLL4	9	0.000	1	3	0.000	0.37
851	AQP2	18	0.001	1	6	0.000	0.37
852	BCL2L11	66	0.003	1	22	0.001	0.37
853	PTHLH	78	0.004	1	26	0.001	0.37
854	IFI27	138	0.007	1	46	0.003	0.37
855	PGC	3	0.000	1	1	0.000	0.37
856	PTCRA	3	0.000	1	1	0.000	0.37
857	IFNA2	24	0.001	1	8	0.000	0.37
858	TWIST2	334	0.016	1	111	0.006	0.37
859	KLF4	112	0.006	1	37	0.002	0.36
860	HMGB3	185	0.009	1	61	0.003	0.36
861	ANXA3	1693	0.084	1	558	0.030	0.36
862	ODC1	405	0.020	1	132	0.007	0.36
863	IL1RAP	384	0.019	1	125	0.007	0.36
864	CCR1	40	0.002	1	13	0.001	0.36
865	FOXA1	253	0.012	1	82	0.004	0.36
866	RSP01	71	0.004	1	23	0.001	0.36
867	F11R	466	0.023	1	150	0.008	0.35
868	CSF3	1298	0.064	1	417	0.023	0.35
869	MST1R	131	0.006	1	42	0.002	0.35
870	PTTG1	163	0.008	1	52	0.003	0.35
871	IL4R	895	0.044	1	283	0.015	0.35
872	CTNNBIP1	54	0.003	1	17	0.001	0.35
873	TNF	54	0.003	1	17	0.001	0.35
874	SOX9	732	0.036	1	229	0.012	0.34
875	PPP1R15A	252	0.012	1	78	0.004	0.34
876	SPATA2	88	0.004	1	27	0.001	0.34
877	MACC1	170	0.008	1	52	0.003	0.34
878	PRDM1	247	0.012	1	75	0.004	0.33
879	EFNA1	449	0.022	1	136	0.007	0.33
880	DSP	1596	0.079	1	482	0.026	0.33
881	KRT5	9363	0.462	1	2827	0.154	0.33
882	FASN	2096	0.103	1	632	0.034	0.33
883	HEY1	10	0.000	1	3	0.000	0.33
884	NFKBIA	433	0.021	1	129	0.007	0.33
885	NFKBIE	195	0.010	1	58	0.003	0.33
886	SOX7	350	0.017	1	104	0.006	0.33
887	TSPAN13	288	0.014	1	85	0.005	0.33
888	SGK2	17	0.001	1	5	0.000	0.32
889	FAM213A	252	0.012	1	74	0.004	0.32
890	FGF17	75	0.004	1	22	0.001	0.32
891	VDR	550	0.027	1	161	0.009	0.32
892	CCNC	225	0.011	1	65	0.004	0.32
893	EPCAM	111	0.005	1	32	0.002	0.32
894	IER3	1443	0.071	1	413	0.022	0.32
895	CRTAC1	56	0.003	1	16	0.001	0.31
896	BHLHE40	1330	0.066	1	380	0.021	0.31
897	PLAU	704	0.035	1	200	0.011	0.31
898	C1orf106	554	0.027	1	157	0.009	0.31
899	GADD45G	50	0.002	1	14	0.001	0.31
900	PIK3R5	43	0.002	1	12	0.001	0.31
901	BMP6	18	0.001	1	5	0.000	0.31
902	MMP3	36	0.002	1	10	0.001	0.31

903	HMGA2	500	0.025	1	138	0.008	0.30
904	CTSV	240	0.012	1	66	0.004	0.30
905	PLK3	603	0.030	1	165	0.009	0.30
906	RAMP3	59	0.003	1	16	0.001	0.30
907	CDK6	451	0.022	1	122	0.007	0.30
908	NFKBIZ	832	0.041	1	224	0.012	0.30
909	ASNS	399	0.020	1	106	0.006	0.29
910	RAD54L	35	0.002	1	9	0.000	0.28
911	PRR15L	176	0.009	1	45	0.002	0.28
912	FGFR3	157	0.008	1	40	0.002	0.28
913	GALNT3	838	0.041	1	213	0.012	0.28
914	CKMT1A	489	0.024	1	124	0.007	0.28
915	VSNL1	214	0.011	1	54	0.003	0.28
916	KRT14	350	0.017	1	88	0.005	0.28
917	KCND2	4	0.000	1	1	0.000	0.28
918	PAX3	20	0.001	1	5	0.000	0.28
919	CCR7	28	0.001	1	7	0.000	0.28
920	GPR126	145	0.007	1	36	0.002	0.27
921	TTC9	57	0.003	1	14	0.001	0.27
922	TLR1	114	0.006	1	28	0.002	0.27
923	DDR1	961	0.047	1	235	0.013	0.27
924	CAMK2B	174	0.009	1	42	0.002	0.27
925	FGF19	46	0.002	1	11	0.001	0.26
926	GDF6	26	0.001	1	6	0.000	0.25
927	FZD8	57	0.003	1	13	0.001	0.25
928	EXOSC8	53	0.003	1	12	0.001	0.25
929	TICAM1	55	0.003	1	12	0.001	0.24
930	CXXC4	33	0.002	1	7	0.000	0.23
931	IDO1	24	0.001	1	5	0.000	0.23
932	ITGA2	882	0.044	1	181	0.010	0.23
933	KIT	5	0.000	1	1	0.000	0.22
934	ALB	20	0.001	1	4	0.000	0.22
935	AREG	9999	0.494	1	1984	0.108	0.22
936	BIRC3	1138	0.056	1	218	0.012	0.21
937	IL20RA	63	0.003	1	12	0.001	0.21
938	PTGS2	280	0.014	1	53	0.003	0.21
939	RBL1	228	0.011	1	43	0.002	0.21
940	CD24	869	0.043	1	162	0.009	0.21
941	FCER2	11	0.001	1	2	0.000	0.20
942	LIF	890	0.044	1	154	0.008	0.19
943	SOD2	7007	0.346	1	1174	0.064	0.18
944	GNGT2	12	0.001	1	2	0.000	0.18
945	OAS1	12	0.001	1	2	0.000	0.18
946	ABCC2	157	0.008	1	26	0.001	0.18
947	TMEM132A	222	0.011	1	36	0.002	0.18
948	KCNK5	32	0.002	1	5	0.000	0.17
949	TNNI2	270	0.013	1	42	0.002	0.17
950	KRT6A	262	0.013	1	40	0.002	0.17
951	JAG1	606	0.030	1	92	0.005	0.17
952	FAM83B	291	0.014	1	43	0.002	0.16
953	CXCL3	901	0.044	1	133	0.007	0.16
954	LAMB3	1577	0.078	1	229	0.012	0.16
955	BECN1	69	0.003	1	10	0.001	0.16
956	TGFA	410	0.020	1	58	0.003	0.16
957	BIK	310	0.015	1	43	0.002	0.15
958	CCND2	544	0.027	1	72	0.004	0.15
959	ALDH1A3	4918	0.243	1	642	0.035	0.14
960	INPP5J	23	0.001	1	3	0.000	0.14
961	CLCA2	529	0.026	1	66	0.004	0.14
962	ITGA6	1676	0.083	1	209	0.011	0.14
963	TP63	303	0.015	1	37	0.002	0.13
964	KRT17	2669	0.132	1	314	0.017	0.13

965	COL1A1	69	0.003	1	8	0.000	0.13
966	CXCL2	1589	0.078	1	178	0.010	0.12
967	CCL15	18	0.001	1	2	0.000	0.12
968	ELMO1	27	0.001	1	3	0.000	0.12
969	TNFAIP3	614	0.030	1	68	0.004	0.12
970	INHBA	222	0.011	1	22	0.001	0.11
971	SOX15	298	0.015	1	29	0.002	0.11
972	IL1B	377	0.019	1	36	0.002	0.11
973	SERPINB2	233	0.012	1	22	0.001	0.10
974	CXCL1	640	0.032	1	59	0.003	0.10
975	CXCL8	653	0.032	1	58	0.003	0.10
976	DLL1	24	0.001	1	2	0.000	0.09
977	SFRP1	648	0.032	1	51	0.003	0.09
978	SFN	6460	0.319	1	507	0.028	0.09
979	CBLC	188	0.009	1	13	0.001	0.08
980	CDH1	1637	0.081	1	113	0.006	0.08
981	HBEGF	75	0.004	1	5	0.000	0.07
982	LAMA3	1951	0.096	1	126	0.007	0.07
983	LAMC2	4464	0.220	1	277	0.015	0.07
984	KLK5	311	0.015	1	19	0.001	0.07
985	ACTA2	17	0.001	1	1	0.000	0.06
986	ARHGEF16	55	0.003	1	3	0.000	0.06
987	IL1A	1331	0.066	1	57	0.003	0.05
988	FGFBP1	2307	0.114	1	81	0.004	0.04
989	DSG3	978	0.048	1	28	0.002	0.03
990	ITGB6	415	0.020	1	3	0.000	0.01
991	COL17A1	517	0.026	1	3	0.000	0.01

Appendix 10: List of deregulated pathways in TRIM24-OE MCF10A cells by HTG









Edgeseq analysis














Hallmarks analysis for Upregulated genes

Gene Set Name [# Genes (K)]	Description	# Genes in Overlap (k)	k/K	p-value	FDR q-value
HALLMARK_APOPTOSIS [161]	Genes mediating programmed cell death (apoptosis) by activation of caspases.	18		1.13 e ⁻¹⁹	5.65 e ⁻¹⁸
HALLMARK_EPITHELIAL_MESENCHYMAL_TRANSITION [200]	Genes defining epithelial-mesenchymal transition, as in wound healing, fibrosis and metastasis.	18		5.82 e ⁻¹⁸	1.46 e ⁻¹⁶
HALLMARK_UV_RESPONSE_DN [144]	Genes down-regulated in response to ultraviolet (UV) radiation.	12		5.43 e ⁻¹²	9.04 e ⁻¹¹
HALLMARK_COMPLEMENT [200]	Genes encoding components of the complement system, which is part of the innate immune system.	12		2.54 e ⁻¹⁰	2.54 e ⁻⁹
HALLMARK_KRAS_SIGNALING_UP [200]	Genes up-regulated by KRAS activation.	12		2.54 e ⁻¹⁰	2.54 e ⁻⁹
HALLMARK_ALLOGRAFT_REJECTION [200]	Genes up-regulated during transplant rejection.	11		3.59 e ⁻⁹	2.99 e ⁻⁸
HALLMARK_ANGIOGENESIS [36]	Genes up-regulated during formation of blood vessels (angiogenesis).	6		1.79 e ⁻⁸	1.28 e ⁻⁷
HALLMARK_ESTROGEN_RESPONSE_LATE [200]	Genes defining late response to estrogen.	10		4.61 e ⁻⁸	2.56 e ⁻⁷
HALLMARK_P53_PATHWAY [200]	Genes involved in p53 pathways and networks.	10		4.61 e ⁻⁸	2.56 e ⁻⁷
HALLMARK_TGF_BETA_SIGNALING [54]	Genes up-regulated in response to TGFβ1 [GeneID=7040].	6		2.21 e ⁻⁷	1.11 e ⁻⁶

Pathway analysis for Upregulated genes

Gene Set Name [# Genes (K)]	Description	# Genes in Overlap (k)	k/K	p-value 	FDR q-value 
ONDER_CDH1_TARGETS_2_UP [256]	Genes up-regulated in HMLE cells (immortalized nontransformed mammary epithelium) after E-cadherin (CDH1) [GeneID=999] knockdown by RNAi.	23		1.28 e ⁻²²	4.33 e ⁻¹⁹
SENESE_HDAC1_TARGETS_DN [260]	Genes down-regulated in U2OS cells (osteosarcoma) upon knockdown of HDAC1 [GeneID=3065] by RNAi.	21		8.83 e ⁻²⁰	1.5 e ⁻¹⁶
VART_KSHV_INFECTION_ANGIOGENIC_MARKERS_UP [165]	Angiogenic markers up-regulated in lymph endothelial cells upon infection with KSHV (Kaposi's sarcoma herpes virus).	18		1.77 e ⁻¹⁹	2.01 e ⁻¹⁶
LEE_BMP2_TARGETS_UP [745]	Genes up-regulated in uterus upon knockout of BMP2 [GeneID=650].	30		3.68 e ⁻¹⁹	3.12 e ⁻¹⁶
KOKKINAKIS_METHIONINE_DEPRIVATION_96HR_6HR_UP [117]	Genes up-regulated in MEWO cells (melanoma) after 96 h of methionine [PubChem=876] deprivation.	16		4.61 e ⁻¹⁹	3.13 e ⁻¹⁶
SCHUETZ_BREAST_CANCER_DUCTAL_INVASIVE_VE_UP [351]	Genes up-regulated in invasive ductal carcinoma (IDC) relative to ductal carcinoma in situ (DCIS, non-invasive).	22		2.67 e ⁻¹⁸	1.51 e ⁻¹⁵

Pathway analysis for Downregulated genes

Gene Set Name [# Genes (K)]	Description	# Genes in Overlap (k)	k/K	p-value 	FDR q-value 
ONDER_CDH1_TARGETS_2_DN [464]	Genes down-regulated in HMLE cells (immortalized nontransformed mammary epithelium) after E-cadherin (CDH1) [GeneID=999] knockdown by RNAi.	80		1.28 e ⁻⁹³	6.79 e ⁻⁹⁰
SMID_BREAST_CANCER_LUMINAL_B_DN [564]	Genes down-regulated in the luminal B subtype of breast cancer.	54		2.96 e ⁻⁴⁸	7.84 e ⁻⁴⁵
SMID_BREAST_CANCER_BASAL_UP [648]	Genes up-regulated in basal subtype of breast cancer samples.	55		2.67 e ⁻⁴⁶	4.71 e ⁻⁴³
CHARAFE_BREAST_CANCER_LUMINAL_VS_BASAL_DN [455]	Genes down-regulated in luminal-like breast cancer cell lines compared to the basal-like ones.	43		3.78 e ⁻³⁸	5.01 e ⁻³⁵
ZWANG_CLASS_3_TRANSIENTLY_INDUCED_BY_EY_EGF [222]	Class III of genes transiently induced by EGF [GeneID=1950] in 184A1 cells (mammary epithelium).	34		1.49 e ⁻³⁷	1.58 e ⁻³⁴
LEI_MYB_TARGETS [318]	Myb-regulated genes in MCF7 (breast cancer) and lung epithelial cell lines overexpressing MYBL2, MYBL1 or MYB [GeneID=4605;4603;4602].	36		7.55 e ⁻³⁵	6.67 e ⁻³²
DELYS_THYROID_CANCER_UP [443]	Genes up-regulated in papillary thyroid carcinoma (PTC) compared to normal tissue.	40		1.01 e ⁻³⁴	7.64 e ⁻³²
GSE9988_LOW_LPS_VS_CTRL_TREATED_MONOCYOCYTE_UP [200]	Genes up-regulated in comparison of monocytes treated with 1 ng/ml LPS (TLR4 agonist) versus monocytes treated with control IgG.	31		1.78 e ⁻³⁴	1.18 e ⁻³¹
JAEGER_METASTASIS_DN [258]	Genes down-regulated in metastases from malignant melanoma compared to the primary tumors.	33		8.66 e ⁻³⁴	5.11 e ⁻³¹

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Vita

Kaushik Thakkar was born in Pune, in the state of Maharashtra, India. He was born on 13th May 1986 to Mr. Narendra Thakkar and Mrs. Lata Thakkar. After completing his high school at S.V Union High School, Kaushik, continued his college education at A.S.D.B Dadawala College of Sciences, University of Pune, Pune from 2003-2006. Kaushik graduated from college with a First class and obtained his Bachelor's in Science (B. Sc) majoring in Microbiology. Following his undergraduate training, Kaushik continued to pursue his Master's of Science majoring in Microbiology at Abasaheb Garware college, Pune from 2006-2008. Kaushik completed his Master's dissertation project titled "Synthesis, characterization and antimicrobial activity of extracellular crystalline silver nanoparticles (AgNPs) from *Morganella* sp." under the mentorship of Dr. Yogesh Shouche. After graduating from University of Pune in 2008, Kaushik worked at two Drug discovery companies, Piramal Life Sciences Limited and Lupin Limited as Research Associate (2008-2010). Kaushik joined the PhD graduate program at the Graduate School of Biomedical Sciences, a joint venture of The University of Texas Health Science Center at Houston and The University of Texas MD Anderson Cancer Center, Houston in August, 2010. Kaushik, joined the lab of Dr. Michelle C. Barton to pursue his PhD dissertation project. His project focuses on understanding the role of histone reader TRIM24 in regulating metabolic reprogramming and EMT in breast cancer.